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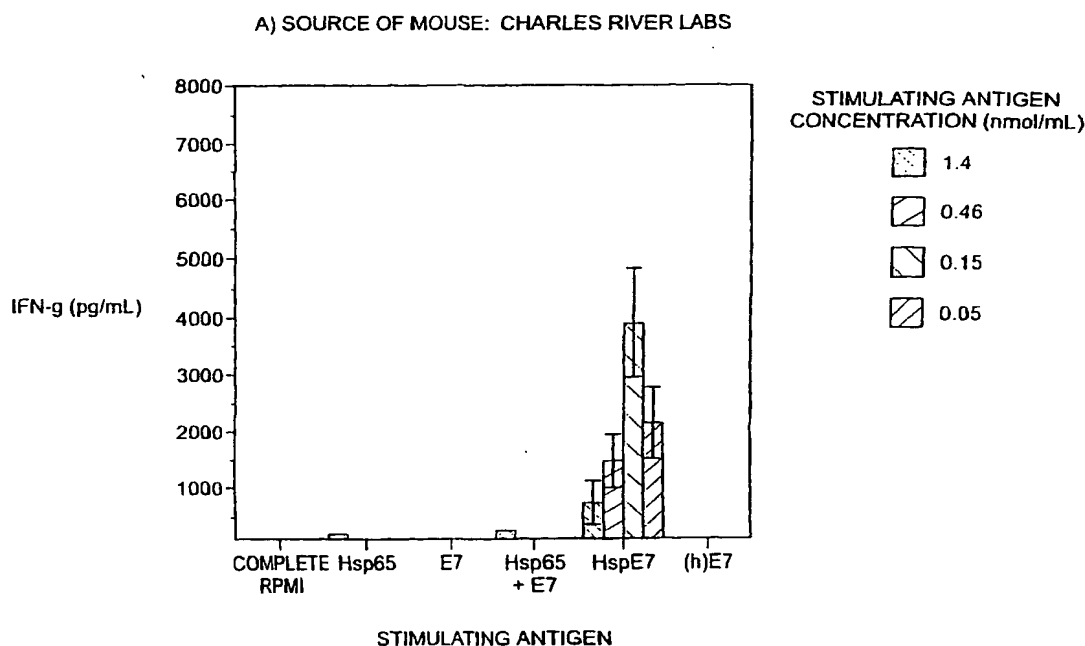
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(54) Title: **INDUCTION OF A TH1-LIKE RESPONSE IN VITRO**



(57) Abstract: The invention provides compositions and methods for stimulating a Th1-like response *in vitro*. Compositions include fusion proteins and conjugates that contain at least a portion of a heat shock protein. A Th1-like response can be elicited by contacting *in vitro* a cell sample containing naive lymphocytes with a fusion protein or conjugate of the invention. The Th1-like response can be detected by measuring IFN-gamma produced by the cell sample.



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INDUCTION OF A TH1-LIKE RESPONSE IN VITRO

Cross Reference to Related Applications

This application claims priority from U.S. Provisional Application
5 No. 60/143,757, filed July 8, 1999. The content of this application is incorporated
herein by reference in its entirety.

Field of the Invention

The invention relates to fusion proteins and methods of stimulating a Th1-
10 like response *in vitro*.

Background

T lymphocytes can generally be divided into two classes based upon expression
of the CD4 and CD8 antigens. The immune response mediated by CD4+ T cells is
15 restricted by class II major histocompatibility complex (MHC) molecules. CD4+ T
cells, also known as helper T lymphocytes, carry out their helper functions via the
secretion of lymphokines. The immune response mediated by CD8+ T cells is
restricted by class I MHC molecules. CD8+ T cells, also known as cytolytic T
lymphocytes (CTLs), carry out cell mediated cytotoxicity and also secrete some
20 lymphokines upon activation.

CD4+ T cells can be further divided into Th1 and Th2 subsets. Th1 cells
participate in cell mediated immunity by producing lymphokines, such as interferon
(IFN)-gamma and tumor necrosis factor (TNF)-beta, that activate cell mediated
immunity. Th2 cells provide help for humoral immunity by secreting lymphokines that
25 stimulate B cells, such as IL-4 and IL-5. Antigenic stimuli that activate either the Th1
or Th2 pathway can inhibit the development of the other. For example, IFN-gamma
produced by a stimulated Th1 cell can inhibit the formation of Th2 cells, and IL-4
produced by a stimulated Th2 cell can inhibit the formation of Th1 cells.

Certain disease conditions, such as cancer, allergy, and parasitic infections, are
30 characterized by a predominantly Th2 response. Under certain circumstances, the
induction of the Th1 response, typified by the production of IFN-gamma, may
ameliorate these conditions.

Summary of the Invention

The invention is based on the discovery that a cell sample containing naive lymphocytes can be stimulated *in vitro* to exhibit a Th1-like response.

Accordingly, the invention features a method of determining whether a fusion protein stimulates a Th1-like response by: (a) providing a cell sample containing naive lymphocytes *in vitro*; (b) providing a fusion protein containing (i) a heat shock protein (Hsp) or a fragment thereof at least eight amino acid residues in length, fused to (ii) a heterologous polypeptide at least eight amino acid residues in length; (c) contacting the cell sample with the fusion protein; and (d) determining whether the fusion protein stimulates a Th1-like response in the cell sample.

“Naive lymphocytes” are lymphocytes that have not been exposed to the fusion protein (*in vivo* or *in vitro*) prior to their use in a method the invention. An “Hsp” is a polypeptide consisting of a sequence that is at least 40% identical to that of a protein whose expression is induced or enhanced in a cell exposed to stress, e.g., heat shock. A “fusion protein” is a non-naturally occurring polypeptide containing amino acid sequences derived from at least two different proteins.

The Hsp used in the method can be selected from the group consisting of Hsp65, Hsp40, Hsp10, Hsp60, and Hsp71. Additionally, the fusion protein can contain the full amino acid sequence of any of Hsp65, Hsp40, Hsp10, Hsp60, or Hsp71. In some embodiments, the fusion protein contains a fragment of an Hsp, e.g., amino acids 1-200 of Hsp65 of *Mycobacterium bovis*.

The heterologous polypeptide can contain a sequence identical to at least eight consecutive amino acids of (i) a protein of a human pathogen, e.g., a virus, or (ii) a tumor associated antigen. Examples of viruses include human papilloma virus (HPV), herpes simplex virus (HSV), hepatitis B virus (HBV), hepatitis C virus (HCV), cytomegalovirus (CMV), Epstein-Barr virus (EBV), influenza virus, measles virus, and human immunodeficiency virus (HIV). The heterologous polypeptide can contain an HPV E6 antigen, e.g., HPV16 E6, an HPV E7 antigen, e.g., HPV16 E7, or a fragment of any of these antigens that is at least eight amino acid residues in length.

In one example, the fusion protein contains *Mycobacterium bovis* BCG Hsp65 and HPV 16 E7.

The cell sample used in the methods of the invention can contain cells derived from a spleen, lymph node, peripheral blood, bone marrow, thymus, lung, respiratory

tract, or anogenital mucosa. In preferred embodiments, the cells are splenocytes or lymph node cells.

The stimulation of a Th1-like response can be determined by detecting the presence of a lymphokine produced by the cell sample, e.g. IFN-gamma or TNF-beta.

5 In one embodiment, the method also includes the steps of: (e) providing a second cell sample containing naive lymphocytes; (f) contacting the second cell sample with a second fusion protein; and (g) determining whether the second fusion protein stimulates a Th1-like response in the second cell sample. In this example, the first fusion protein contains the sequence of a full-length, naturally occurring Hsp, and the
10 second fusion protein contains at least eight amino acids but less than all of the sequence of a naturally occurring Hsp.

 In another aspect, the invention features a method of screening a compound by: (a) providing a cell sample containing naive lymphocytes *in vitro*; (b) providing a fusion protein containing (i) a Hsp or a fragment thereof at least eight amino acid
15 residues in length, fused to (ii) a heterologous polypeptide at least eight amino acid residues in length; (c) contacting the cell sample with the compound and the fusion protein; and (d) determining whether the cell sample exhibits a Th1-like response following the contacting step. In this method, a decrease in the Th1-like response in the presence of the compound compared to in the absence of the compound indicates
20 that the compound inhibits a Th1-like response by the cell sample.

 The invention also includes a method of screening a compound by: (a) providing a cell sample containing naive lymphocytes *in vitro*; (b) providing a fusion protein containing (i) a Hsp or a fragment thereof at least eight amino acid residues in length, fused to (ii) a heterologous polypeptide at least eight amino acid residues in
25 length; (c) contacting the cell sample with the compound and the fusion protein; and (d) determining whether the cell sample exhibits a Th1-like response following the contacting step. In this method, an increase in the Th1-like response in the presence of the compound compared to in the absence of the compound indicates that the compound promotes a Th1-like response by the cell sample.

30 In another aspect, the invention features a method of determining whether a hybrid compound stimulates a Th1-like response by: (a) providing a cell sample containing naive lymphocytes *in vitro*; (b) providing a hybrid compound that is non-naturally occurring and contains (i) a non-peptide compound having a molecular weight

of less than 1,500, covalently linked to (ii) a polypeptide of at least eight amino acids in length, wherein the hybrid compound is made by covalently linking the non-peptide compound to the polypeptide; (c) contacting the cell sample with the hybrid compound; and (d) determining whether the hybrid compound stimulates a Th1-like response in the cell sample. In one embodiment, the non-peptide compound has a molecular weight of at least 100.

In another aspect, the invention features a method of determining whether a hybrid compound stimulates a Th1-like response by: (a) producing a hybrid compound by covalently linking a non-peptide compound to a polypeptide of at least eight amino acids in length; (b) providing a cell sample containing naive lymphocytes *in vitro*; (c) contacting the cell sample with the hybrid compound; and (d) determining whether the hybrid compound stimulates a Th1-like response in the cell sample. In one embodiment, the non-peptide compound has a molecular weight between 100 and 1,500.

In another aspect, the invention features a method of determining whether a fusion protein stimulates a Th1-like response by: (a) providing a cell sample containing naive lymphocytes *in vitro*; (b) providing a fusion protein comprising (i) a first polypeptide at least eight amino acids in length, fused to (ii) a second polypeptide at least eight amino acids in length; (c) contacting the cell sample with the fusion protein; and (d) detecting a Th1-like response exhibited by the cell sample following the contacting step. In one embodiment, the detected Th1-like response is greater than a Th1-like response exhibited by a second cell sample containing naive lymphocytes when the second cell sample is contacted with either the first polypeptide, the second polypeptide, or a mixture of the first polypeptide and the second polypeptide. In one example, the detected Th1-like response is at least two times greater than the Th1-like response exhibited by the second cell sample. In another example, the detected Th1-like response is at least five times greater than the Th1-like response exhibited by the second cell sample.

In another aspect, the invention provides a fusion protein containing (i) a Hsp10 protein or a fragment thereof at least eight amino acid residues in length, and (ii) a heterologous polypeptide at least eight amino acids in length. The Hsp10 protein of the fusion protein can be a mycobacterial protein, e.g., *Mycobacterium tuberculosis* Hsp10 protein. The heterologous polypeptide can contain a sequence identical to at least eight

consecutive amino acids of a protein of a human virus, e.g., HPV. In one example, the heterologous polypeptide contains HPV16 E7.

In another aspect, the invention provides a fusion protein containing (i) a Hsp40 protein or a fragment thereof at least eight amino acid residues in length, and (ii) a heterologous polypeptide at least eight amino acids in length. The Hsp40 protein of the fusion protein can be a mycobacterial protein, e.g., *Mycobacterium tuberculosis* Hsp40 protein. The heterologous polypeptide can contain a sequence identical to at least eight consecutive amino acids of a protein of a human virus, e.g., HPV. In one example, the heterologous polypeptide contains HPV16 E7.

In another aspect, the invention provides a fusion protein containing (i) a Hsp71 protein or a fragment thereof at least eight amino acid residues in length, and (ii) a heterologous polypeptide at least eight amino acids in length. The Hsp71 protein of the fusion protein can be a mycobacterial protein, e.g., *Mycobacterium tuberculosis* Hsp71 protein. The heterologous polypeptide can contain a sequence identical to at least eight consecutive amino acids of a protein of a human virus, e.g., HPV. In one example, the heterologous polypeptide contains HPV16 E7.

In another aspect, the invention features a method of determining whether a compound stimulates a Th1-like response by: (a) providing a cell sample containing naive lymphocytes *in vitro*; (b) providing a compound; (c) contacting the cell sample with the compound; and (d) detecting a Th1-like response exhibited by the cell sample following the contacting step.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present application, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

Other features and advantages of the invention will be apparent from the following detailed description, and from the claims.

Brief Description of the Drawings

Figures 1A-1B show the sequence of plasmid pET65 coding for expression of Hsp65.

5 Figure 2 shows the sequence of plasmid pET/E7 (NH) coding for expression of E7.

Figure 3 shows the sequence of plasmid pET/H/E7 coding for expression of (h)E7.

Figures 4A-4B show the sequence of plasmid pET65C/E7-1N coding for expression of HspE7.

10 Figures 5A-5B show the sequence of plasmid pETMT40E7 coding for expression of MT40-E7.

Figure 6 shows the sequence of plasmid pET/OVA coding for expression of ovalbumin (OVA).

15 Figures 7A-7C show the sequence of plasmid pET65H/OVA coding for expression of HspOVA.

Figure 8 shows the sequence of plasmid pGEX/K coding for expression of GST.

Figure 9 shows the sequence of plasmid pGEX/K/E7 coding for expression of GST-E7.

20 Figures 10A-10B show the sequence of plasmid pET/E7/5'65 coding for expression of E7-L-BCG65.

Figure 11 shows the sequence of plasmid pET65F1/E7 coding for expression of BCG65(F1)-E7.

Figure 12 shows the sequence of plasmid pETESE7 coding for expression of TB10-E7.

25 Figures 13A-13B show the sequence of plasmid pET/E7/71 coding for expression of E7-TB71.

Figures 14A-14B show the sequence of plasmid pET/E7/71' coding for expression of a fusion protein.

30 Figures 15A-15B show the sequence of plasmid pET/SP65c-E7 coding for expression of SP65(2)-E7.

Figures 16A-16B show the sequence of plasmid pETAf60E7 coding for expression of AF60-E7.

Figures 17A-17B show enhanced IFN-gamma release by splenocytes from C57BL/6 mice obtained from the Charles River Laboratory (Fig. 17A) and the Jackson Laboratory (Fig. 17B) upon exposure to HspE7.

Figures 18A-18C show enhanced IFN-gamma release by splenocytes from Balb/c (Fig. 18A), C57BL/6 (Fig. 18B), and C3HeB/FeJ (Fig. 18C) mice upon exposure to HspE7.

Figure 19 shows enhanced IFN-gamma release by splenocytes upon exposure to fusion proteins containing an antigen and a stress protein but not upon exposure to a fusion protein containing an antigen and a protein other than a stress protein.

Figures 20A-20B show enhanced IFN-gamma release by splenocytes upon exposure to fusion proteins containing stress proteins of different types, stress proteins from different organisms, or a fragment of a stress protein.

Figure 21 shows enhanced IFN-gamma release by lymph node cells and splenocytes upon exposure to fusion proteins containing an antigen and a stress protein.

Figures 22A-22B show a time course of tumor incidence (Fig. 22A) and tumor volume (Fig. 22B) in mice injected with TC-1 tumor cells followed by an injection with either saline, HspE7, SP65(2)-E7, or AF60-E7.

Figures 23A-23B show a time course of tumor incidence (Fig. 23A) and tumor volume (Fig. 23B) in mice injected with TC-1 tumor cells followed by an injection with either saline, HspE7, MT40-E7, E7-MT71, or TB10-E7.

Detailed Description

The invention relates to methods of stimulating *in vitro* a Th1-like response in a cell sample containing naive lymphocytes. These methods are useful for assessing the ability of a protein, e.g., a fusion protein containing an Hsp linked to a heterologous polypeptide, to function as a stimulator of a Th1-like response. Additionally, the method can be used to identify compounds that can regulate a Th1-like response. Various materials and procedures suitable for use in the methods of the invention are discussed below.

The terms stress protein and heat shock protein (Hsp) are used synonymously herein. An Hsp is a polypeptide consisting of a sequence that is at least 40% identical to that of a protein whose expression is induced or enhanced in a cell exposed to stress. Turning to stress proteins generally, cells respond to a stressor (typically heat shock

treatment) by increasing the expression of a group of genes commonly referred to as stress, or heat shock, genes. Heat shock treatment involves exposure of cells or organisms to temperatures that are one to several degrees Celsius above the temperature to which the cells are adapted. In coordination with the induction of such genes, the levels of corresponding stress proteins increase in stressed cells. As used herein, a "stress protein," also known as a "heat shock protein" or "Hsp," is a protein that is encoded by a stress gene, and is therefore typically produced in significantly greater amounts upon the contact or exposure of the stressor to the organism. A "stress gene," also known as "heat shock gene" is used herein as a gene that is activated or otherwise detectably upregulated due to the contact or exposure of an organism (containing the gene) to a stressor, such as heat shock, hypoxia, glucose deprivation, heavy metal salts, inhibitors of energy metabolism and electron transport, and protein denaturants, or to certain benzoquinone ansamycins. Nover, L., *Heat Shock Response*, CRC Press, Inc., Boca Raton, FL (1991). "Stress gene" also includes homologous genes within known stress gene families, such as certain genes within the Hsp70 and Hsp90 stress gene families, even though such homologous genes are not themselves induced by a stressor. Each of the terms stress gene and stress protein as used in the present specification may be inclusive of the other, unless the context indicates otherwise.

An antigen can be any compound, peptide or protein to which an immune response is desired. Antigens of particular interest are tumor-associated antigens, allergens of any origin, and proteins from viruses, mycoplasma, bacteria, fungi, protozoa and other parasites.

Fusion Proteins

The invention provides Hsp fusion proteins. As used herein, a "fusion protein" is a non-naturally occurring polypeptide containing at least two amino acid sequences which generally are from two different proteins. The amino acid sequence of the full length fusion protein is not identical to the amino acid sequence of a naturally occurring protein or a fragment thereof. An Hsp fusion protein contains an Hsp or a fragment thereof at least eight amino acids in length linked to a heterologous polypeptide. An "Hsp polypeptide" refers to a polypeptide consisting of a sequence that is at least 40% identical to that of a protein whose expression is induced or enhanced in a cell exposed to stress, e.g., heat shock. A "heterologous polypeptide" refers to a polypeptide that is

fused to the Hsp protein or fragment thereof. The heterologous polypeptide is preferably at least eight amino acids in length. In some embodiments, the heterologous polypeptide is at least 10, 20, 50, 100, 150, 180, 200, or 300 amino acids in length. The heterologous polypeptide generally is not part or all of a naturally occurring Hsp.

5 However, the fusion protein can also be a fusion between a first Hsp and a second, different, Hsp, or between all or portion of an Hsp fused to all or a portion of the same Hsp (as long as the resultant fusion is not identical to a naturally occurring protein). The Hsp polypeptide can be attached to the N-terminus or C-terminus of the heterologous polypeptide. Preferably the fusion protein is a purified protein.

10 The preferred Hsp fusion protein has one Hsp polypeptide linked to one heterologous polypeptide, but other conformations are within the invention. In one embodiment, the fusion protein comprises at least two copies of the heterologous polypeptide, e.g., HPV16 E7. In another embodiment, the fusion protein contains at least two copies of the Hsp polypeptide, e.g., Hsp65. Additionally, the fusion protein
15 can contain at least two different heterologous polypeptides, e.g., two or more fragments of a single antigenic protein representing different epitopes or fragments of two or more different antigenic proteins derived from the same or different tumors or pathogens, and/or at least two different Hsp polypeptides.

The Hsp and heterologous polypeptide can be directly fused without a linker
20 sequence. In preferred embodiments, the C-terminus of the Hsp can be directly fused to the N-terminus of the heterologous polypeptide or the C-terminus of the heterologous polypeptide can be directly fused to the N-terminus of the Hsp.

Alternatively, Hsp and heterologous polypeptides can be linked to each other via a peptide linker sequence. Preferred linker sequences (1) should adopt a flexible
25 extended conformation, (2) should not exhibit a propensity for developing an ordered secondary structure which could interact with the functional Hsp and heterologous polypeptide domains, and (3) should have minimal hydrophobic or charged character, which could promote interaction with the functional protein domains. Typical surface amino acids in flexible protein regions include Gly, Asn and Ser. Permutations of
30 amino acid sequences containing Gly, Asn and Ser would be expected to satisfy the above criteria for a linker sequence. Other neutral or near-neutral amino acids, such as Thr and Ala, can also be used in the linker sequence. Any other amino acid can also be used in the linker. A linker sequence length of fewer than 20 amino acids can be used

to provide a suitable separation of functional protein domains, although longer linker sequences may also be used.

The Hsp fusion protein may be further fused to another amino acid sequence that facilitates the purification of the fusion protein. One useful fusion protein is a GST fusion protein in which the Hsp-heterologous polypeptide sequences are fused to the C-terminus or N-terminus of the GST sequence. Another useful fusion protein is a poly-histidine (His) fusion protein in which the Hsp-heterologous polypeptide sequences are fused to either the C-terminus or N-terminus of the poly-histidine sequence, e.g. His x 6. In another embodiment, the fusion protein contains the chitin-binding region of intein, thereby permitting the purification of the fusion protein by chitin beads (Hoang et al. (1999) *Gene* 1999 237:361-71). In another embodiment, the fusion protein contains a signal sequence from another protein. In certain host cells (e.g., mammalian host cells), expression and/or secretion of the Hsp fusion protein can be increased through use of a heterologous signal sequence. For example, the gp67 secretory sequence of the baculovirus envelope protein can be used as a heterologous signal sequence (Current Protocols in Molecular Biology, Ausubel et al., eds., John Wiley & Sons, 1992). Other examples of eukaryotic signal sequences include the secretory sequences of melittin and human placental alkaline phosphatase (Stratagene; La Jolla, California). Prokaryotic signal sequences useful for increasing secretion by a prokaryotic host cell include the phoA secretory signal (Molecular Cloning, Sambrook et al., second edition, Cold Spring Harbor Laboratory Press, 1989) and the protein A secretory signal (Pharmacia Biotech; Piscataway, New Jersey).

Fusion proteins of the invention, e.g., a fusion protein of Hsp65 and HPV16 E7, can be produced by standard recombinant techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together, in any order, in-frame in accordance with conventional techniques. Such techniques can include employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. Correct linkage of the two nucleic acids requires that the product of the linkage encode a chimeric protein consisting of a Hsp moiety and a heterologous polypeptide moiety. In another embodiment, the fusion gene can be synthesized by conventional techniques, including automated DNA synthesizers. Alternatively, PCR amplification of gene

fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments, which are subsequently annealed and reamplified to generate a chimeric gene sequence (see, e.g., Current Protocols in Molecular Biology, Ausubel et al. eds., John Wiley & Sons: 1992).

5 Expression vectors encoding fusion proteins containing a heterologous polypeptide and either an Hsp or a protein other than an Hsp can be prepared by the above procedures. Examples of Hsp fusion proteins can be found in international patent application WO 99/07860, incorporated herein by reference, that describes vector construction, expression and purification of *Mycobacterium bovis* BCG Hsp65 -
10 HPV16 E7 (HspE7) fusion protein as well as of HPV16 E7 (E7), histidine tagged HPV16 E7 (hE7), and *M. bovis* BCG Hsp65 (Hsp65). Additional examples of nucleic acids encoding an Hsp optionally linked to a heterologous polypeptide, e.g., an HPV antigen, are described in WO 89/12455, WO 94/29459, WO 98/23735, and references cited therein, the contents of which are herein incorporated by reference.

15 A variety of heat shock proteins have been isolated, cloned, and characterized from a diverse array of organisms (Mizzen, Biotherapy 10:173-189, 1998). Any Hsp or fragment thereof may be suitable for use in the fusion polypeptides and conjugates of the invention. For example, Hsp70, Hsp60, Hsp20-30, and Hsp10 are among the major determinants recognized by host immune responses to infection by *Mycobacterium*
20 *tuberculosis* and *Mycobacterium leprae*. In addition, Hsp65 of Bacille Calmette Guerin (BCG), a strain of *Mycobacterium bovis*, was found to be an effective stimulatory agent, as described in the examples below.

 Families of stress genes and proteins for use in the present invention are well known in the art and include, for example, Hsp100-200, Hsp100, Hsp90, Lon, Hsp70,
25 Hsp60, TF55, Hsp40, FKBP, cyclophilins, Hsp20-30, ClpP, GrpE, Hsp10, ubiquitin, calnexin, and protein disulfide isomerases. See, e.g., Macario, Cold Spring Harbor Laboratory Res. 25:59-70, 1995; Parsell et al., Rev. Genet. 27:437-496, 1993; and U.S. Patent No. 5,232,833. Preferred Hsps include Hsp65, Hsp40, Hsp10, Hsp60, and Hsp71.

30 The Hsp portion of the fusion protein can include either a full length Hsp or a fragment of an Hsp at least eight amino acids in length. In some embodiments, the Hsp fragment is greater than 10 amino acids in length, and preferably is at least 20, 50, 100, 150, 180, 200, or 300 amino acids in length. In one embodiment, the Hsp portion of the

fusion protein consists of amino acids 1-200 of Hsp65 of *Mycobacterium bovis*. Other portions of Hsp65 and other Hsps can be used in a fusion protein to elicit a Th1-like response *in vitro*. Other preferred Hsps include Hsp40 of *M. tuberculosis*, Hsp10 of *M. tuberculosis*, Hsp65 of *Streptococcus pneumoniae*, and Hsp60 of *Aspergillus fumigatus*.

5 Heterologous polypeptides can contain any amino acid sequence useful for stimulating an immune response, *in vitro* and/or *in vivo*. Preferably, the heterologous polypeptide contains an MHC-binding epitope, e.g., an MHC class I or MHC class II binding epitope. The heterologous polypeptide can contain sequences found in a protein produced by a human pathogen, e.g., viruses, bacteria, mycoplasma, fungi, protozoa, and other parasites, or sequences found in the protein of a tumor associated antigen (TAA). Examples of viruses include human papilloma virus (HPV), herpes simplex virus (HSV), hepatitis B virus (HBV), hepatitis C virus (HCV), cytomegalovirus (CMV), Epstein-Barr virus (EBV), influenza virus, measles virus, and human immunodeficiency virus (HIV). Examples of tumor associated antigens include
10 MAGE1, MAGE2, MAGE3, BAGE, GAGE, PRAME, SSX-2, Tyrosinase, MART-1, NY-ESO-1, gp100, TRP-1, TRP-2, A2 melanotope, BCR/ABL, Procinase-3/Myeloblastin, HER2/neu, CEA, P1A, HK2, PAPA, PSA, PSCA, PSMA, pg75, MUM-1, MUC-1, E6, E7, GnT-V, Beta-catenin, CDK4 and
15 P15.

20 HPV antigens from any strain of HPV are suitable for use in the fusion polypeptide. HPV expresses six or seven non-structural and two structural proteins. Viral capsid proteins L1 and L2 are the late structural proteins. L1 is the major capsid protein, the amino acid sequence of which is highly conserved among different HPV types. There are seven early non-structural proteins. Proteins E1, E2, and E4 play an
25 important role in virus replication. Protein E4 also plays a role in virus maturation. The role of E5 is less well known. Proteins E6 and E7 are oncoproteins critical for viral replication, as well as for host cell immortalization and transformation. Fusion proteins of the invention can contain either the entire sequence of an HPV protein or a fragment thereof, e.g., a fragment of at least 8 amino acids. In one embodiment, the
30 HPV antigenic sequence is derived from a "high risk" HPV, such as HPV16 or HPV18 E7 protein. The HPV antigenic sequence can include an MHC-binding epitope, e.g., an MHC class I and/or an MHC class II binding epitope.

In addition to Hsp fusion proteins, other fusion proteins can be used in the *in vitro* assay described herein. These non-Hsp fusion proteins contain a first polypeptide at least eight amino acids in length, fused to a second polypeptide at least eight amino acids in length, wherein the first and second polypeptides are derived from different proteins (preferably naturally occurring proteins). The fusion protein itself does not have the sequence of a naturally occurring protein.

In the fusion protein of the invention, neither the first nor second polypeptide is an amino acid sequence that is commonly used for protein purification or detection, e.g., GST or poly-histidine.

In order to produce the fusion protein, a nucleic acid encoding the fusion protein can be introduced into a host cell, e.g., a bacterium, a primary cell, or an immortalized cell line using an expression vector. The recombinant cells are then used to produce the fusion protein. The transfection can be transient or stable, the later sometimes accomplished by homologous recombination.

The nucleotide sequence encoding a fusion protein will usually be operably linked to one or more regulatory sequences, selected on the basis of the host cells to be used for expression. The term "regulatory sequence" refers to promoters, enhancers and other expression control elements (e.g., polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel (1990) *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA, the content of which is incorporated herein by reference. Regulatory sequences include those that direct constitutive expression of a nucleotide sequence in many types of host cells, those that direct expression of the nucleotide sequence only in certain host cells (e.g., tissue-specific regulatory sequences), and those that direct expression in a regulatable manner (e.g., only in the presence of an inducing agent). It will be appreciated by those skilled in the art that the design of the expression vector may depend on such factors as the choice of the host cell to be transformed, the level of expression of fusion protein desired, and the like.

Recombinant expression vectors can be designed for expression of fusion proteins in prokaryotic or eukaryotic cells. For example, fusion proteins can be expressed in bacterial cells such as *E. coli*, insect cells (e.g., in the baculovirus expression system), yeast cells or mammalian cells. Some suitable host cells are discussed further in Goeddel (1990) *Gene Expression Technology: Methods in*

Enzymology 185, Academic Press, San Diego, CA. Examples of vectors for expression in yeast *S. cerevisiae* include pYepSec1 (Baldari *et al.* (1987) *EMBO J.* 6:229-234), pMFa (Kurjan and Herskowitz (1982) *Cell* 30:933-943), pJRY88 (Schultz *et al.* (1987) *Gene* 54:113-123), and pYES2 (Invitrogen Corporation, San Diego, CA). Baculovirus
5 vectors available for expression of fusion proteins in cultured insect cells (*e.g.*, Sf 9 cells) include the pAc series (Smith *et al.* (1983) *Mol. Cell. Biol.* 3:2156-2165) and the pVL series (Lucklow and Summers (1989) *Virology* 170:31-39).

Examples of mammalian expression vectors include pCDM8 (Seed (1987) *Nature* 329:840) and pMT2PC (Kaufman *et al.* (1987), *EMBO J.* 6:187-195). When
10 intended for use in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40.

In addition to the regulatory control sequences discussed above, the recombinant expression vector can contain additional nucleotide sequences. For
15 example, the recombinant expression vector may encode a selectable marker gene to identify host cells that have incorporated the vector. Moreover, to facilitate secretion of the fusion protein from a host cell, in particular mammalian host cells, the recombinant expression vector can encode a signal sequence linked to the amino-terminus of the fusion protein, such that upon expression, the fusion protein is synthesized with the
20 signal sequence fused to its amino terminus. This signal sequence directs the fusion protein into the secretory pathway of the cell and is then usually cleaved, allowing for release of the mature fusion protein (*i.e.*, the fusion protein without the signal sequence) from the host cell. Use of a signal sequence to facilitate secretion of proteins or peptides from mammalian host cells is known in the art.

25 Vector DNA can be introduced into prokaryotic or eukaryotic cells *via* conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" refer to a variety of art-recognized techniques for introducing foreign nucleic acid (*e.g.*, DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection,
30 lipofection, electroporation, microinjection and viral-mediated transfection. Suitable methods for transforming or transfecting host cells can be found in Sambrook *et al.* (*Molecular Cloning: A Laboratory Manual*, 2nd Edition, Cold Spring Harbor Laboratory Press (1989)), and other laboratory manuals.

Often only a small fraction of mammalian cells integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (*e.g.*, resistance to antibiotics) can be introduced into the host cells along with the gene encoding the fusion protein. Preferred selectable markers include those that confer resistance to drugs such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding the fusion protein or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (*e.g.*, cells that have incorporated the selectable marker gene will survive, while the other cells die).

Alternatively, a recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

In addition to the recombinant techniques described above, a fusion protein of the invention can be formed by linking two polypeptides, *e.g.*, a Hsp and a heterologous polypeptide, to form a conjugate. Methods of forming Hsp conjugates are described in WO 89/12455, WO 94/29459, WO 98/23735, and WO 99/07860, the contents of which are herein incorporated by reference. As used herein, an Hsp "conjugate" comprises an Hsp that has been covalently linked to a heterologous polypeptide via the action of a coupling agent. A conjugate thus comprises two separate molecules that have been coupled one to the other. The term "coupling agent," as used herein, refers to a reagent capable of coupling one polypeptide to another polypeptide, *e.g.*, a Hsp to a heterologous polypeptide. Any bond which is capable of linking the components such that the linkage is stable under physiological conditions for the time needed for the assay (*e.g.*, at least 12 hours, preferably at least 72 hours) is suitable. The link between two components may be direct, *e.g.*, where a Hsp is linked directly to a heterologous polypeptide, or indirect, *e.g.*, where a Hsp is linked to an intermediate, *e.g.*, a backbone, and that intermediate is also linked to the heterologous polypeptide. A coupling agent should function under conditions of temperature, pH, salt, solvent system, and other reactants that substantially retain the chemical stability of the Hsp, the backbone (if present), and the heterologous polypeptide.

A coupling agent can link components, *e.g.*, a Hsp and a heterologous polypeptide, without the addition of the coupling agent to the resulting fusion protein. Other coupling agents result in the addition of the coupling agent to the resulting fusion

protein. For example, coupling agents can be cross-linking agents that are homo- or hetero-bifunctional, and wherein one or more atomic components of the agent is retained in the composition. A coupling agent that is not a cross-linking agent can be removed entirely following the coupling reaction, so that the molecular product is composed entirely of the Hsp, the heterologous polypeptide, and a backbone moiety (if present).

Many coupling agents react with an amine and a carboxylate, to form an amide, or an alcohol and a carboxylate to form an ester. Coupling agents are known in the art, see, e.g., M. Bodansky, "Principles of Peptide Synthesis", 2nd ed., referenced herein, and T. Greene and P. Wuts, "Protective Groups in Organic Synthesis," 2nd Ed, 1991, John Wiley, NY. Coupling agents should link component moieties stably, but such that there is minimal or no denaturation or deactivation of the Hsp or the heterologous polypeptide.

The conjugates of the invention can be prepared by coupling a Hsp to a heterologous polypeptide using methods known in the art. A variety of coupling agents, including cross-linking agents, can be used for covalent conjugation. Examples of cross-linking agents include N,N'-dicyclohexylcarbodiimide (DCC; Pierce), N-succinimidyl-S-acetyl-thioacetate (SATA), N-succinimidyl-3-(2-pyridyldithio)propionate (SPDP), ortho-phenylenedimaleimide (o-PDM), and sulfosuccinimidyl 4-(N-maleimidomethyl) cyclohexane-1-carboxylate (sulfo-SMCC). See, e.g., Karpovsky *et al.* (1984) *J. Exp. Med.* 160:1686 and Liu *et al.* (1985) *Proc. Natl. Acad. Sci. USA* 82:8648. Other methods include those described by Paulus (1985) *Behring Ins. Mitt.* 78:118-132; Brennan *et al.* (1985) *Science* 229:81-83; and Glennie *et al.* (1987) *J. Immunol.* 139: 2367-2375. A large number of coupling agents for peptides and proteins, along with buffers, solvents, and methods of use, are described in the Pierce Chemical Co. catalog, pages T-155 –T-200, 1994 (3747 N. Meridian Rd., Rockford IL, 61105, U.S.A.; Pierce Europe B.V., P.O. Box 1512, 3260 BA Oud Beijerland, The Netherlands), which catalog is hereby incorporated by reference.

DCC is a useful coupling agent (Pierce #20320; Rockford, IL). It promotes coupling of the alcohol NHS in DMSO (Pierce #20684), forming an activated ester which can be cross-linked to polylysine. DCC (N,N'-dicyclohexylcarbodiimide) is a carboxy-reactive cross-linker commonly used as a coupling agent in peptide synthesis,

and has a molecular weight of 206.32. Another useful cross-linking agent is SPDP (Pierce #21557), a heterobifunctional cross-linker for use with primary amines and sulfhydryl groups. SPDP has a molecular weight of 312.4 and a spacer arm length of 6.8 angstroms, is reactive to NHS-esters and pyridyldithio groups, and produces

5 cleavable cross-linking such that upon further reaction, the agent is eliminated so the Hsp can be linked directly to a backbone or heterologous polypeptide. Other useful conjugating agents are SATA (Pierce #26102) for introduction of blocked SH groups for two-step cross-linking, which are deblocked with hydroxylamine-HCl (Pierce #26103), and sulfo-SMCC (Pierce #22322), reactive towards amines and sulfhydryls.

10 Other cross-linking and coupling agents are also available from Pierce Chemical Co. (Rockford, IL). Additional compounds and processes, particularly those involving a Schiff base as an intermediate, for conjugation of proteins to other proteins or to other compositions, for example to reporter groups or to chelators for metal ion labeling of a protein, are disclosed in EP 243,929 A2 (published Nov. 4, 1987).

15 Polypeptides that contain carboxyl groups can be joined to lysine ϵ -amino groups in the heterologous polypeptide either by preformed reactive esters (such as N-hydroxy succinimide ester) or esters conjugated *in situ* by a carbodiimide-mediated reaction. The same applies to Hsps containing sulfonic acid groups, which can be transformed to sulfonyl chlorides that react with amino groups. Hsps that have

20 carboxyl groups can be joined to amino groups on the polypeptide by an *in situ* carbodiimide method. Hsps can also be attached to hydroxyl groups of serine or threonine residues, or to sulfhydryl groups of cysteine residues.

In addition to conjugates of two polypeptides, e.g., a Hsp and a heterologous polypeptide, hybrid compounds can be constructed containing a non-peptide compound

25 covalently linked to a polypeptide at least eight amino acids in length. The polypeptide component of this hybrid compound can be any of the heterologous polypeptides described herein as a component of a Hsp fusion protein or conjugate. Examples of the non-peptide component of this hybrid compound include polynucleotides, polynucleotide analogs, nucleotides, nucleotide analogs, organic or inorganic

30 compounds having a molecular weight less than about 5,000 grams per mole, preferably between about 1,500 and 100 grams per mole, and salts, esters, and other pharmaceutically acceptable forms of such non-peptide compounds.

In Vitro Assays for Th1-Like Activity

Cell samples containing naive lymphocytes are prepared from any mammal, e.g., a mouse, rat, rabbit, goat, or human, and are plated at an appropriate density in one or more tissue culture plates. A naive lymphocyte is a lymphocyte that has not been
5 exposed (either *in vivo* or *in vitro*) to the fusion protein (or to either of the polypeptides that are joined to make the fusion protein) prior to the cell's use in the *in vitro* assay. The cell sample can be derived from any of various primary or secondary lymphoid organs or tissues of an animal, e.g., spleen, lymph node, peripheral blood, bone marrow, or thymus. The sample may also be derived from any tissue in the body
10 containing lymphoid cells, such as the lung, respiratory tract (including pharynx, larynx, trachea, bronchi, etc), and anogenital mucosa. The cell sample can include naive lymphocytes selected from NK cells, NK T cells, $\alpha\beta$ T cells and $\gamma\delta$ T cells. The cell sample can be either unfractionated or enriched for a particular cell type or cell types. In addition to naive lymphocytes, the cell sample can optionally include naive
15 antigen presenting cells such as macrophages, dendritic cells, and/or B cells. The cell sample can optionally include cell lines, e.g., a transformed T cell line or a T cell clone.

The cell sample is exposed *in vitro* to a fusion protein or a conjugate described herein. Following a period of incubation between the cell sample and the fusion protein or conjugate, e.g., 6, 12, 24, 36, 48, 60, 72, or 96 hours, a determination is made
20 as to whether a Th1-like response has been elicited in the cell sample. A Th1-like response can be detected, for example, by measuring the production of particular lymphokines, e.g., IFN-gamma or TNF-beta, by the cell sample. Alternatively, a Th1-like response can be detected by assaying for cell surface marker expression, such as SLAM (signaling lymphocytic activation molecule), or for cytokine expression, using a
25 variety of techniques (for example, flow cytometry).

In one example, pooled, unfractionated splenocyte cultures containing naive lymphocytes are prepared from a mouse and are plated in tissue culture plates. Methods of isolating and culturing splenocytes are described in Current Protocols in Immunology, Coligan et al., eds., John Wiley & Sons, 2000. Cultures of splenocytes
30 are then exposed to different concentrations of a test protein, e.g., a recombinant Hsp fusion protein, Hsp, the antigen alone, or another antigen-containing fusion protein, for a time that is sufficient to elicit a measurable IFN-gamma response against a standard antigen-stress protein fusion protein such as, for example, HspE7, described in patent

application WO 99/07860 and employed in the Examples below. Following exposure of the cell sample to the test protein, the IFN-gamma level in the extracellular medium is determined using a suitable assay such as an IFN-gamma ELISA.

Results of the assays described below reveal that IFN-gamma release elicited by exposure of splenocytes or lymph node cells to an Hsp fusion protein is much more substantial than that induced by exposure to the antigen itself, the Hsp itself, an admixture of antigen and Hsp, or a fusion between antigen and a protein other than a Hsp.

The assay of the invention can be used to evaluate a preparation of an Hsp fusion protein (e.g., as a quality control assay) or compare different preparations of Hsp fusion proteins. The measurements taken in the assay constitute a method for identifying a particularly active batch or to eliminate substandard batches of fusion protein preparations. The assay may also be used to optimize production procedures, storage regimes, etc. In cases in which a maximal Th1-like response to a particular antigen is desired, the assays can be used to test different fusions between the antigen and different types of Hsps or Hsps of different origins. Furthermore, the assay can be used to test a series of different candidate antigens, to identify the antigen that gives rise to the most pronounced Th1-like response when fused to a Hsp.

The assay can also be used to identify regions in an antigen sequence or an Hsp sequence that are primarily responsible for eliciting a Th1-like response and thus have therapeutic potential. To identify such active regions in an antigen, fusions containing individual subregions of the antigen fused to an Hsp can be prepared and tested in the assay of the invention. To identify active regions in an Hsp, fusions containing individual subregions of the Hsp fused to the antigen can be prepared and tested. These determinations will provide the basis for the construction of shortened fusion proteins comprising subregions of antigen and/or Hsp that are sufficient to elicit a Th1-like response. Fusions containing subregions of a Hsp and/or subregions of an antigen can be tested by comparing the elicited Th1-like response to that induced by a full length fusion protein with known activity, e.g., HspE7.

The fusion proteins described herein are useful in assays for screening compounds for their effectiveness in stimulating a Th1-like response. For example, the Hsp fusion proteins that were found to stimulate IFN-gamma secretion in the *in vitro*

assay can be used as controls to test candidate compounds for their ability to produce the same effect.

The system described herein for stimulating a Th1-like response *in vitro* can be used to generate activated Th1 cells *ex vivo* for reimplantation into an individual. This
5 may be useful for treating conditions characterized by a dominant Th2 immune response and an insufficient Th1 response.

The assay can also be used to identify compounds that can regulate a Th1-like response. Compounds can be screened for their ability to inhibit an Hsp-fusion protein-induced Th1-like response, or to promote a Th1-like response in a manner similar to a
10 Hsp fusion protein, or to enhance the Th1-like response induced by a Hsp fusion protein (or any other protein found to act in a manner comparable to a Hsp fusion protein). Inhibitory compounds may be useful to treat conditions characterized by an inappropriate Th1 response, e.g., inflammatory and autoimmune diseases. Potential inhibitors (e.g., of binding of antigen-stress protein fusion proteins to antigen-
15 presenting cells or of stress protein fusion-enhanced antigen processing) can be screened as follows. A cell sample comprising naive lymphocytes is mixed with a fusion protein or conjugate that is known to induce a Th1-like response, e.g., IFN-gamma secretion. Compounds to be screened as potential inhibitors are added to the cell culture either before, after, or simultaneous to the addition of the fusion protein or
20 conjugate. The effect of the compound on the induction of a Th1-like response, e.g., as measured by IFN-gamma release, can be determined by comparing the response to that obtained when the fusion protein or conjugate alone is added to the cell sample.

In a similar manner, compounds can be screened for their ability to promote a Th1-like response. Any compound can be screened for its ability to regulate a Th1-like
25 response, including both peptides and non-peptide chemicals. These compounds include, but are not limited to, peptides, peptidomimetics (e.g., peptoids), amino acids, amino acid analogs, polynucleotides, polynucleotide analogs, nucleotides, nucleotide analogs, organic or inorganic compounds having a molecular weight less than about 5,000 grams per mole, and salts, esters, and other pharmaceutically acceptable forms of
30 such compounds. In this case, a cell sample comprising naive lymphocytes is contacted with a test compound. The effect of the test compound on the induction of a Th1-like response, e.g., as measured by IFN-gamma release, is then measured and compared to a control (no test sample) or compared to an Hsp fusion known to stimulate a Th1-like

response. This assay can be used to identify novel compounds that can be used to stimulate a Th1-like response. Preferably the Th1-like response stimulated by the compound is at least 25%, e.g., at least 40%, 50%, 60%, 70%, or 80%, the level of the maximum response induced by an HspE7 fusion protein. In one embodiment, the compound is preferably not a naturally occurring compound. In another embodiment, the compound is a peptide, wherein the peptide does not correspond to the fragment of a naturally occurring protein.

The following are examples of the practice of the invention. They are not to be construed as limiting the scope of the invention in any way.

Examples

Example 1: Bacterial Growth and Cell Lysis for Production of Recombinant Proteins

E. coli strains BL21(DE3) or BLR(DE3) (Novagen) were used as the host for all recombinant protein production, with the exception of pET65, which was transformed into BL21(DE3) pLysS (Novagen). BL21(DE3) pLysS cells harboring pET65 were grown in 2xYT media (20 g/L tryptone; 10 g/L yeast extract, 20 g/L NaCl; Milli-Q™ quality water) containing 30 µg/ml kanamycin and 34 µg/ml chloramphenicol, while all other transformants were grown in 2xYT media containing 30 µg/ml kanamycin. All bacterial cultures were grown in 2 L shaker flasks at 200-400 rpm to OD₆₀₀=0.5 and then induced with 0.5 mM IPTG for 3 hours at 37°C. Cells were then harvested by centrifugation at 4°C and 4,000 – 8,000 g for 5 minutes, then suspended in 300 ml of Lysis Buffer (10 mM TRIS·HCl, 10 mM 2-mercaptoethanol, pH 7.5), lysozyme was added to 200 µg/mL, and the suspension mixed and frozen at -70°C.

To purify the recombinant protein, the cells were thawed using a 37°C waterbath and proteinase inhibitors were added (2 µg/ml aprotinin, 2 µg/ml leupeptin, 2 µg/ml pepstatin and 2 mM PMSF). The cell suspension was split into 50 mL samples, stored on ice, and sonicated 3–4 times for 30 seconds at Power-Level 5 - 8 (Sonicator 450, Branson, Corp.). The supernatant was separated from the pellet by centrifugation at 35,000 – 60,000 g for 10 – 20 minutes at 4°C. For soluble proteins, the supernatant was kept and processed as the Soluble Fraction. For proteins found in inclusion bodies,

the supernatant was discarded and the pellet was washed with Lysis Buffer (optionally containing 1 M urea, 1 %(v/v) Triton X-100). The resulting mixture was then centrifugation at 35,000 – 60,000 g for 10 –20 minutes at 4°C and the supernatant discarded. The pellet was dissolved in Lysis Buffer containing 8 M urea. This mixture
5 was then centrifuged at 4°C for 10 - 20 minutes at 35,000 – 60,000 g and the pellet was discarded and the supernatant stored at -70°C as the Inclusion Body fraction.

Example 2: Production of Recombinant *M. bovis* BCG Hsp65 (Hsp65)

A plasmid encoding Hsp65 was constructed as follows. The *M. bovis* BCG
10 Hsp65 coding sequence was PCR amplified from pRIB1300 (van Eden *et al.* (1988) Nature 331:171-173) using the following primers. The forward primer (w046: 5' TTC GCC ATG GCC AAG ACA ATT GCG 3'; SEQ ID NO:1) contains an ATG start codon at an NcoI site. The reverse primer (w078: 5' TTC TCG GCT AGC TCA GAA ATC CAT GCC 3'; SEQ ID NO:2) contains an Nhe I site downstream of a TGA stop
15 codon. The PCR product was digested with NcoI and NheI, purified and ligated to pET28a (Novagen) which had been cut with NcoI and NheI. Plasmid pET65 encodes the *M. bovis* BCG Hsp65 protein, abbreviated Hsp65. The nucleotide sequence (SEQ ID NO:3) coding for expression of Hsp65 (SEQ ID NO:4) is shown in Figs. 1A-1B.

The Hsp65 protein was purified as follows. The Soluble Fraction was prepared
20 as described above from *E. coli* BL21(DE3) pLysS cells transformed with plasmid pET65. The *M. bovis* BCG Hsp65 protein (Hsp65) present in the Soluble Fraction was purified by the following chromatographic steps: SP-Sepharose (200 ml column, Amersham Pharmacia), Q-Sepharose (200 ml column, Amersham Pharmacia), Sephacryl S-300 (500 ml column, Amersham Pharmacia) and ceramic hydroxyapatite
25 (HAP; 100 ml column, Biorad). Purified Hsp65 was exchanged into Dulbecco's modified phosphate buffered saline (DPBS)/15% (v/v) glycerol and stored at -70°C.

Example 3: Production of Recombinant HPV16 E7 (E7)

A plasmid encoding HPV16 E7 was constructed as follows. The HPV16 E7
30 coding sequence was PCR-amplified from pSK/HPV16 (ATCC) using primers w280 and w134 (w280: CCA GCT GTA ACC ATG GAT GGA GAT (SEQ ID NO:5) and w134: AGC CAT GAA TTC TTA TGG TTT CTG (SEQ ID NO:6)). The PCR product was digested with restriction enzyme Nco I and EcoR I and purified from an agarose

gel. The purified PCR product was ligated to pET28a that had been previously digested with the same enzymes. The ligation reaction was used to transform *E. coli* DH5alpha and putative clones containing the HPV16 E7 gene insert were selected based on diagnostic restriction digestion. This initial restriction analysis was confirmed by DNA sequence analysis of entire gene, promoter and termination regions. DNA of the confirmed construct, named pET/E7 (NH), was then introduced by electroporation into *E. coli* strain BL21(DE3). The nucleotide sequence (SEQ ID NO:7) coding for expression of E7 (SEQ ID NO:8) is shown in Fig. 2.

The HPV16 E7 protein was purified as follows. The Soluble Fraction was prepared as described above from *E. coli* BL21(DE3) cells transformed with plasmid pET/E7 (NH). The HPV 16 E7 protein was purified by the following chromatographic steps: Q-Sepharose (100 ml column, Amersham Pharmacia); Superdex 200 (26/60 column, Amersham Pharmacia); and Ni-chelating Sepharose (100 ml, Amersham Pharmacia) under denaturing conditions with serial washings containing 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100, and the pooled fractions containing HPV E7 protein were then dialyzed overnight against 30 mM TRIS-HCl, 1 M NaCl, 1 mM 2-mercaptoethanol, pH 7.5. The dialyzed protein was further purified by Ni-chelating Sepharose (75 ml, Amersham Pharmacia) under denaturing conditions with serial washings containing 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100. The purity of the protein was checked by SDS-PAGE, the appropriate fractions pooled and dialyzed overnight at 4°C against DPBS/10 % (v/v) glycerol.

Example 4: Production of Recombinant Histidine-tagged HPV 16 E7 ((h)E7)

A plasmid encoding (h)E7 was constructed as follows. The HPV16 E7 coding sequence was PCR amplified from HPV16 genomic DNA (pSK/HPV16) using the following primers. The forward primer (w133: 5' AAC CCA GCT GCT AGC ATG CAT GGA GAT 3'; SEQ ID NO:9) contains an NheI site upstream of an ATG start codon. The reverse primer (w134: 5' AGC CAT GAA TTC TTA TGG TTT CTG 3'; SEQ ID NO:10) contains an EcoRI site downstream of a TAA stop codon. The PCR product was digested with NheI and EcoRI, purified and ligated to pET28a which had been cut with NheI and EcoRI. pET/H/E7 which encodes the HPV16 E7 protein containing an N-terminal histidine tag, abbreviated (h)E7, was used to transform *E. coli*

BL21(DE3) cells. The nucleotide sequence (SEQ ID NO:11) coding for expression of (h)E7 (SEQ ID NO:12) is shown in Fig. 3.

The (h)E7 protein was purified as follows. The Inclusion Body fraction was prepared as described above from *E. coli* BL21(DE3) cells transformed with plasmid pET/H/E7. The N-terminal histidine-tagged HPV16 E7 protein ((h)E7) present in the Inclusion Body fraction was purified using the following chromatographic steps: Ni-chelating Sepharose (60 ml, Amersham Pharmacia) under denaturing conditions with serial washings containing 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100. Bound (h)E7 was refolded on the resin and eluted by a 50-500 mM imidazole gradient. Purified (h)E7 was dialyzed against DPBS/25% (v/v) glycerol.

Example 5: Production of Recombinant HPV 16 E7 - *M. bovis* BCG 65 Fusion Protein (HspE7)

A plasmid encoding HspE7 was constructed as follows. The *M. bovis* BCG Hsp65 coding sequence was PCR amplified from pRIB1300 using the same forward primer (w046) as for pET65. The reverse primer (w076: 5' CGC TCG GAC GCT AGC TCA CAT ATG GAA ATC CAT GCC 3'; SEQ ID NO:13) contains an NdeI site upstream and an NheI site downstream of a TGA stop codon. The PCR product was digested with NcoI and NheI, purified and ligated to pET28a which had been cut with NcoI and NheI.

The HPV16 E7 coding sequence was PCR amplified from HPV16 genomic DNA (pSK/HPV16) using the following primers. The forward primer (w151: 5' CCA GCT GTA CAT ATG CAT GGA GAT 3'; SEQ ID NO:14) contains an ATG start codon at an NdeI site. The reverse primer (w134: 5' AGC CAT GAA TTC TTA TGG TTT CTG 3'; SEQ ID NO:15) contains an EcoRI site downstream of a TAA stop codon. The PCR product was digested with NdeI and EcoRI, purified and ligated to pET65C which had been cut with Nde I and EcoRI and the resulting plasmid (pET65C/E7-1N) was transformed into *E. coli* BL21(DE3) cells. pET65C/E7-1N encodes a fusion protein consisting of Hsp65 linked via its C-terminus to HPV16 E7, abbreviated HspE7. The nucleotide sequence (SEQ ID NO:16) coding for expression of HspE7 (SEQ ID NO:17) is shown in Figs. 4A-4B.

The HspE7 protein was purified as follows. The Soluble Fraction was prepared as described above from *E. coli* BL21(DE3) cells transformed with plasmid pET65C/E7-1N. Hsp65-HPV16 E7 fusion protein (HspE7) present in the Soluble Fraction was purified by the following chromatographic steps: 0-15% ammonium sulfate precipitation, Ni-chelating Sepharose (100 ml column, Amersham Pharmacia) and Q-Sepharose (100 ml column, Amersham Pharmacia). Endotoxin was removed by extensive washing with 1% (v/v) Triton X-100 on a Ni-chelating Sepharose column in the presence of 6M guanidine-HCl (Gu-HCl). Purified HspE7 was exchanged into DPBS/15% (v/v) glycerol and stored at -70°C.

Example 6: Production of Recombinant *M. tuberculosis* Hsp40 – HPV 16 E7 Fusion Protein (MT40-E7)

pETMT40E7 is a plasmid encoding chimeric recombinant protein MT40E7 composed of *Mycobacterium tuberculosis* (strain H37RV - ATCC 27294) hsp40 protein with hu HPV16 (ATCC 45113) E7 protein attached at the C-terminus of Hsp40. The plasmid was transformed into *E. coli* BL21(DE3) cells for protein production and purification. The nucleotide sequence (SEQ ID NO:18) coding for expression of MT40-E7 (SEQ ID NO:19) is shown in Figs. 5A-5B.

The MT40-E7 protein was purified as follows. The Inclusion Body fraction was prepared as described above from *E. coli* BL21(DE3) cells transformed with plasmid pETMT40E7. MT40-E7 protein was purified using the following chromatographic steps: Q-Sepharose (100 ml column, Amersham Pharmacia), Ni-chelating Sepharose (70 ml, Amersham Pharmacia) under native conditions with serial washings containing 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100. The purity of the protein was checked by SDS-PAGE, the appropriate fractions pooled and dialyzed overnight at 4°C against DPBS/25% (v/v) glycerol.

Example 7: Ovalbumin (OVA)

Ovalbumin (Lot # 37H7010) was purchased from Sigma Chemicals and purified by chromatography using 20 mL of Con A Sepharose (Amersham-Pharmacia). Fractions containing the purified product were pooled and dialyzed overnight against DPBS.

Example 8: Production of Recombinant *M. bovis* BCG Hsp65-Ovalbumin Fusion Protein (HspOva)

A plasmid encoding HspOva was constructed as follows. The full length
5 chicken ovalbumin-coding sequence was excised from pET/OVA with Nhe I and EcoR
I digestion and purified from an agarose gel. The sequence coding for expression of
OVA is shown in Fig. 6. The purified product was ligated to pET65H previously
digested with the same enzymes. The ligation reaction was used to transform *E. coli*
DH5alpha and putative clones containing the chicken ovalbumin gene insert were
10 selected based on diagnostic restriction digestion. This initial restriction analysis was
confirmed by DNA sequence analysis of the entire fusion gene, promoter and
termination regions. DNA of the confirmed construct, named pET65H/OVA, was used
to transform *E. coli* BL21(DE3). The nucleotide sequence (SEQ ID NO:20) coding for
expression of HspOVA (SEQ ID NO:21) is shown in Figs. 7A-7C.

15 The HspOva protein was purified as follows. The Inclusion Body fraction was
prepared as described above from *E. coli* BL21(DE3) cells transformed with plasmid
pET65H/OVA. The HspOva fusion protein present in the Inclusion Body fraction was
purified using the following chromatographic steps: Q-Sepharose (100 ml column,
Amersham Pharmacia) and Ni-chelating Sepharose (60 ml, Amersham Pharmacia)
20 under denaturing conditions with serial washings containing 2% (v/v) Triton X-100
followed by serial washing to remove residual Triton X-100. The purity of the protein
was checked by SDS-PAGE, the appropriate fractions pooled and dialyzed overnight at
4°C against DPBS/15% (v/v) glycerol, followed by a dialysis against DPBS/2.5 % (w/v)
sucrose.

25 Example 9: Production of Recombinant Glutathione-S-Transferase (GST)

A plasmid encoding Gst was constructed as follows. The kanamycin resistance-
coding sequence was excised from pET28a DNA with AlwN I and Xho I digestion and
purified from an agarose gel. The purified product was ligated to pGEX-4T-2 that had
30 been previously digested with the same enzymes. The ligation reaction was used to
transform *E. coli* DH5alpha and putative clones containing the kanamycin resistance
gene insert were selected based on diagnostic restriction digestion. This initial
restriction analysis was confirmed by DNA sequence analysis of the entire insert

coding sequence, promoter and termination regions. DNA of the confirmed construct, named pGEX/K, was used to transform *E. coli* strain BL21(DE3). The nucleotide sequence (SEQ ID NO:22) coding for expression of GST (SEQ ID NO:23) is shown in Fig. 8.

5 The GST protein was purified as follows. The Soluble fraction was prepared as described above from *E. coli* BL21(DE3) cells transformed with plasmid pGEX/K. The GST protein present in the Soluble Fraction was purified by Glutathione-Agarose Chromatography as follows. Approximately 20 mL of Glutathione-Agarose (Sigma-Aldrich; Cat. #: G4510) was equilibrated with DPBS, and mixed and incubated
10 overnight with the sample at room temperature on a shaker. The next morning, the resin was packed into a column and serially washed with DPBS. Endotoxin was removed by washing with 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100. Finally, the protein was eluted using 10 mM glutathione (reduced form), 50 mM TRIS-HCl, pH 8.0.

15

Example 10: Production of Recombinant Glutathione-S-Transferase – HPV 16 E7 Fusion Protein (GST-E7)

A plasmid encoding GST-E7 was constructed as follows. The HPV16 E7 coding sequence was excised from pETOVA/E7 with BamH I and EcoR I digestion
20 and purified from an agarose gel. The purified product was ligated to pGEX/K that had been previously digested with the same enzymes. The ligation reaction was used to transform *E. coli* DH5alpha and putative clones containing the HPV16-E7 gene insert were selected based on diagnostic restriction digestion. This initial restriction analysis was confirmed by DNA sequence analysis of entire fusion gene, promoter and
25 termination regions. DNA of the confirmed construct, named pGEX/K/E7, was used to transform *E. coli* strain BL21(DE3). The nucleotide sequence (SEQ ID NO:24) coding for expression of GST-E7 (SEQ ID NO:25) is shown in Fig. 9.

The GST-E7 protein was purified as follows. Bacteria containing the expression vector pGEX/K/E7 were grown and the protein purified using the affinity
30 chromatography procedure essentially as described above for GST.

Example 11: Production of Recombinant HPV 16 E7 – Linker – *M. bovis* BCG Hsp65 Fusion Protein (E7-L-BCG65)

A plasmid encoding E7-L-BCG65 was constructed as follows. The HPV16 E7-coding sequence was PCR-amplified from pSK/HPV16 (ATCC) using primers w280 and w396 (w280: CCA GCT GTA ACC ATG GAT GGA GAT (SEQ ID NO:26) and w396: GCC ATG GTA CTA GTT GGT TTC TGA GAA (SEQ ID NO:27)). The PCR product was digested with restriction enzyme Nco I and Spe I and purified from an agarose gel. The purified PCR product was ligated to pET5'65 (pET5'65 is pET65 with a polyglycine linker sequence inserted at the 5' end of the *M. bovis* BCG hsp65 sequence) that had been previously digested with the same enzymes. The ligation reaction was used to transform *E. coli* DH5alpha and putative clones containing the HPV16 E7 gene insert were selected based on diagnostic restriction digestion. This initial restriction analysis was confirmed by DNA sequence analysis of entire fusion gene, promoter and termination regions. DNA of confirmed construct, named pET/E7/5'65, was used to transform *E. coli* strain BLR(DE3). The nucleotide sequence (SEQ ID NO:28) coding for expression of E7-L-BCG65 (SEQ ID NO:29) is shown in Figs. 10A-10B.

The E7-L-BCG65 protein was purified as follows. The Soluble Fraction was prepared as described above from *E. coli* BLR(DE3) cells transformed with plasmid pET/E7/5'65. The E7-L-BCG65 fusion protein present in the Soluble Fraction was purified using the following chromatographic steps: Butyl Sepharose (100 ml, Amersham-Pharmacia), Q-Sepharose (100 ml column, Amersham Pharmacia), Superdex 200 Gel Filtration (26/60 column, Amersham Pharmacia), and Ni-chelating Sepharose Fast Flow Chromotography (60 ml, Amersham Pharmacia) under denaturing conditions with serial washings containing 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100. The purity of the protein was checked by SDS-PAGE, the appropriate fractions pooled and dialyzed overnight at 4°C against DPBS. In order to reduce the amount of endotoxin contained in the sample, it was further purified using a pre-packed 1 ml column of DetoxiGel™ (Pierce, Rockford, IL, USA) according to the manufacturer's instructions.

Example 12: Production of Recombinant HPV 16 E7 – *M. bovis* BCG Hsp65
Fragment Fusion Protein (BCG65(F1)-E7)

A plasmid encoding BCG65(F1)-E7 was constructed as follows. The first 600 amino terminal base pairs of *M. bovis* BCG hsp65 gene were PCR-amplified from
5 pET65C/E7-1N using primers w046 and w293 (w046: TTC GCC ATG GCC AAG
ACA ATT GCG (SEQ ID NO:30) and w293: GTA CCC CGA CAT ATG GCC CTT
GTC GAA CCG CAT AC(SEQ ID NO:31)). The PCR product was digested with the
restriction enzymes Nco I and Nde I and purified from an agarose gel. The purified
10 PCR product was ligated to pET65C/E7-1N that had been previously digested with the
same enzymes. The ligation reaction was used to transform *E. coli* DH5alpha and
putative clones containing the truncated BCG65 gene were selected based on diagnostic
restriction digestion. This initial restriction analysis was confirmed by DNA sequence
analysis of the entire fusion gene, promoter and termination regions. The confirmed
plasmid construct, named pET65F1/E7, was used to transform *E. coli* strain
15 BLR(DE3). The nucleotide sequence (SEQ ID NO:32) coding for expression of
BCG65(F1)-E7 (SEQ ID NO:33) is shown in Fig. 11.

The BCG65(F1)-E7 protein was purified as follows. The Inclusion Body
fraction was prepared as described above from *E. coli* BLR(DE3) cells transformed
with plasmid pET65F1/E7. The BCG65(F1)-E7 fusion protein present in the Inclusion
20 Body fraction was purified using the following chromatographic steps: Source 15Q
Sephacrose (Amersham-Pharmacia) and Ni-chelating Sepharose (60 ml, Amersham
Pharmacia) under denaturing conditions with serial washings containing 2% (v/v)
Triton X-100 followed by serial washing to remove residual Triton X-100. The purity
of the protein was checked by SDS-PAGE, the appropriate fractions pooled and
25 dialyzed overnight at 4°C against DPBS.

Example 13: Production of Recombinant *M. tuberculosis* Hsp10 – HPV 16 E7 Fusion
Protein (TB10-E7)

Expression plasmid pETESE7 contains a chimeric gene composed of the
30 *Mycobacterium tuberculosis* strain H37RV (ATCC 27294) groES (hsp10) coding
sequence fused, at its 3' end, to the HPV16 (ATCC 45113) E7 coding. The chimeric
gene was cloned into expression vector pET28a and transformed into *E. coli*
BL21(DE3) cells for protein production and purification. The nucleotide sequence

(SEQ ID NO:34) coding for expression of TB10-E7 (SEQ ID NO:35) is shown in Fig. 12.

The TB10-E7 protein was purified as follows. The Inclusion Body fraction was prepared as described above from *E. coli* BL21(DE3) cells transformed with plasmid pETESE7. The TB10-E7 fusion protein present in the Inclusion Body fraction was purified using the following chromatographic steps: DEAE Sepharose (100 ml column, Amersham Pharmacia), Source 15Q Sepharose (100 ml column, Amersham Pharmacia) and Ni-chelating Sepharose (60 ml, Amersham Pharmacia) under denaturing conditions with serial washings containing 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100. The purity of the protein was checked by SDS-PAGE, the appropriate fractions pooled and dialyzed overnight at 4°C against DPBS/10 % (v/v) glycerol.

Example 14: Production of Recombinant HPV 16 E7 – *M. tuberculosis* Hsp71 Fusion Protein (E7-TB71)

A plasmid encoding E7-TB71 was constructed as follows. The *M. tuberculosis* hsp71 gene was PCR-amplified from clone pY3111/8 (Mehlert and Young (1989) Mol. Microbiol. 3:125-130) using primers w048 and w079 (w048: 5'-TTC ACC ATG GCT CGT GCG GTC GGG (SEQ ID NO:36) and w079: ACC TCC GCG TCC ACA GCT AGC TCA GCC (SEQ ID NO:37)). The PCR product was digested with Nco I and Nhe I, gel-purified and ligated to pET28a digested with the same enzymes to generate pET/71.

The HPV16 E7-coding sequence was PCR-amplified from pSK/HPV16 (ATCC) using primers w280 and w344 (w280: CCA GCT GTA ACC ATG GAT GGA GAT (SEQ ID NO:38) and w344: GGA TCA GAC ATG GCC ATG GCT GGT TTC TG (SEQ ID NO:39)). The PCR product was digested with restriction enzyme Nco I and purified from an agarose gel. The purified PCR product was ligated to pET/71 DNA that had been previously digested with Nco I and CIAP to remove 5' phosphate. The ligation reaction was used to transform *E. coli* DH5alpha and putative clones containing the HPV16 E7 gene insert were selected based on diagnostic restriction digestion. This initial restriction analysis was confirmed by DNA sequence analysis of entire fusion gene, promoter and termination regions. The confirmed construct, named pET/E7/71, was used to transform *E. coli* strain BL21(DE3). The nucleotide sequence

(SEQ ID NO:40) coding for expression of E7-TB71 (SEQ ID NO:41) is shown in Figs. 13A-13B. The resulting construct, pET/E7/71, was further modified (to complete sequences at the 3' end of the hsp71 gene) by replacement of a Kpn I to Nhe I fragment containing sequences from the 3' end of the hsp71 gene by a Kpn I- and Nhe I-digested PCR fragment amplified from pY3111/8 using primers w391 and w392 (w391: GAG GGT GGT TCG AAG GTA CC (SEQ ID NO:42) and w392: TTT GAT TTC GCT AGC TCA CTT GGC CTC(SEQ ID NO:43)). The resulting final plasmid, pET/E7/71', expresses HPV16 E7 fused to the amino-terminus of full-length Hsp71 protein and was used to transform *E. coli* strain BL21(DE3). The nucleotide sequence (SEQ ID NO:44) coding for expression of the fusion protein (SEQ ID NO:45) of pET/E7/71' is shown in Figs. 14A-14B.

The E7-TB71 protein was purified as follows. The Inclusion Body fraction was prepared as described above from *E. coli* BL21(DE3) cells transformed with plasmid pET/E7/71'. The E7-TB71 fusion protein present in the Inclusion Body fraction was purified using the following chromatographic steps: Q-Sepharose (100 ml column, Amersham Pharmacia) and Ni-chelating Sepharose (80 ml, Amersham Pharmacia) under native conditions with serial washings containing 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100. The purity of the protein was checked by SDS-PAGE, the appropriate fractions pooled and dialyzed overnight at 4°C against DPBS/10 %(v/v) glycerol.

Example 15: Production of Recombinant *Streptococcus pneumoniae* HSP65(2) – HPV 16 E7 Fusion Protein (SP65(2)-E7)

A plasmid encoding SP65(2)-E7 was constructed as follows. The *Streptococcus pneumoniae* hsp65 gene was PCR-amplified from plasmid pETP60-2 (PCT patent application WO 99/35720) using primers w384 and w385 (w384: GCA GCC CCA TGG CAA AAG AAA (SEQ ID NO:46) and w385: GCT CGA ATT CGG TCA GCT AGC TCC GCC CAT (SEQ ID NO:47)). The PCR product was digested with Nco I and EcoR I, gel-purified and ligated to pET28a digested with the same enzymes to generate pET/SP65-2C.

The HPV16 E7-coding sequence was PCR-amplified from pSK/HPV16 (ATCC) using primers w133 and w134 (w133: AAC CCA GCT GCT AGC ATG CAT GGA GAT (SEQ ID NO:48) and w134: AGC CAT GAA TTC TTA TGG TTT CTG

(SEQ ID NO:49)). The PCR product was digested with restriction enzymes Nhe I and EcoR I and purified from an agarose gel. The purified PCR product was then ligated to pET/SP65-2C that had been previously digested with Nhe I and EcoR I. The ligation reaction was used to transform *E. coli* DH5alpha and putative clones containing the HPV16 E7 insert were selected based on diagnostic restriction digestion. This initial restriction analysis was confirmed by DNA sequence analysis of entire fusion gene, promoter and termination regions. DNA of the confirmed construct, named pET/SP65c-E7, was used to transform *E. coli* strain BLR(DE3). The nucleotide sequence (SEQ ID NO:50) coding for expression of SP65(2)-E7 (SEQ ID NO:51) is shown in Figs. 15A-15B.

The SP65(2)-E7 protein was purified as follows. The Inclusion Body fraction was prepared as described above from *E. coli* BLR(DE3) cells transformed with plasmid pET/SP65c-E7. The SP65(2)-E7 fusion protein present in the Inclusion Body fraction was purified using the following chromatographic steps: Q-Sepharose (100 ml column, Amersham Pharmacia) and Ni-chelating (60 ml, Amersham Pharmacia) under denaturing conditions with serial washings containing 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100. The purity of the protein was checked by SDS-PAGE, the appropriate fractions pooled and dialyzed overnight at 4°C against DPBS.

Example 16: Recombinant Production of *Aspergillus fumigatus* Hsp60- HPV 16 E7 Fusion Protein (AF60-E7)

pETAF60E7 is a plasmid encoding a recombinant protein, AF60-E7, composed of the *Aspergillus fumigatus* (ATCC 26933) Hsp60 protein (without leader) (obtained as described in PCT/CA99/01152) fused at its C-terminus to the HPV16 (ATCC 45113) E7 protein sequence. Plasmid pETAF60E7 was used to transform *E. coli* BL21(DE3) cells for protein production and purification. The nucleotide sequence (SEQ ID NO:52) coding for expression of AF60-E7 (SEQ ID NO:53) is shown in Figs. 16A-16B.

The AF60-E7 protein was purified as follows. The Inclusion Body fraction was prepared as described above from *E. coli* BL21(DE3) cells transformed with plasmid pETAF60E7. AF60-E7 protein was purified using the following chromatographic steps: Source 15Q Sepharose (Amersham-Pharmacia) and Ni-chelating Sepharose (60

ml, Amersham Pharmacia) under denaturing conditions with serial washings containing 2% (v/v) Triton X-100 followed by serial washing to remove residual Triton X-100. The purity of the protein was checked by SDS-PAGE, the appropriate fractions pooled and dialyzed overnight at 4°C against DPBS.

5

Example 17: Stimulation of IFN-Gamma Release by a Hsp65-HPV E7 (HspE7)

Fusion Protein

Pooled, unfractionated splenocytes were prepared from untreated naive C57BL/6 mice obtained from two different sources (Charles River Laboratory and Jackson Laboratory) and were plated in complete medium (complete RPMI) at 6×10^5 cells/well in flat bottom 96-well tissue culture plates. Replicate cultures (5) were incubated for 72 hours with 0.05 to 1.4 nmol/mL concentrations of recombinant *M. bovis* BCG Hsp65 (Hsp65), HPV16 E7 (E7) or histidine-tagged E7 ((h)E7), an admixture of *M. bovis* BCG Hsp65 and HPV16 E7 (Hsp65 + E7), or *M. bovis* BCG Hsp65 - HPV16 E7 fusion protein (HspE7). Subsequent to incubation, cells were pelleted, and supernatants were transferred to IFN-gamma capture ELISA plates.

After incubation, the replicate samples were harvested, pooled in eppendorf tubes and pelleted at 1200 rpm for 7 minutes in Beckman GS-6R centrifuge (300 x g). The supernatants were removed into cryovials and frozen at -70°C until time of analysis.

Maxisorp ELISA plates (Nunc cat# 442404A) were coated overnight at 4°C with 1 µg/mL purified rat anti-mouse IFN-gamma (PharMingen cat. no 18181D) in 0.1 M NaHCO₃ buffer, pH 8.2. The plates were washed with 0.05% Tween 20 in PBS then blocked with 3% BSA (albumin fraction V: Amersham cat. no 10857) in DPBS (blocking buffer) for 2 hours. After the plates were washed, recombinant mouse IFN-gamma (8000, 4000, 2000, 1000, 500, 250, 125, 62.5 pg/mL in complete RMPI) was placed in triplicate onto each ELISA plate. Sample supernatants were removed from -70°C, thawed quickly at 37°C, and placed undiluted onto the ELISA plates in duplicate. The samples were then serially diluted by seven, 3-fold dilutions in complete RPMI followed by incubation at 4°C overnight. Background ELISA values were established by measuring eight wells containing all reagents except the target antigen.

Detection of bound murine IFN-gamma was accomplished using 1 µg/mL of a rat anti-mouse IFN-gamma biotin conjugate (PharMingen cat. no 18112D) in blocking

buffer. Following washing, bound biotin-conjugated antibody was detected using a 1:1000 dilution of a streptavidin-alkaline phosphatase conjugate (Caltag cat. no SA1008). The plates were washed as before followed by the addition of a chromogenic substrate, p-nitrophenyl phosphate (pNPP; Sigma cat# N-2765) at 1 mg/mL in diethanolamine buffer, pH 9.5. After 30 minutes incubation, the color reaction was stopped using 50 μ L of 100 mM EDTA, pH 8.0. The absorbance was measured at 410 nm using a Dynatech MR5000 ELISA plate reader equipped with Biolinx 2.0 software. The levels of IFN-gamma detected in test samples were extrapolated from the standard curves generated on each of the respective ELISA plates. Data is expressed as IFN-gamma released (pg/mL \pm SD).

Results of assays are shown in Figs. 17A-17B. The averages from five replicates are shown along with the standard deviation. Substantial secretion of IFN-gamma was elicited by exposure of splenocytes to 0.05, 0.15, 0.46 and 1.4 nmol/mL HspE7. Hsp65 alone, E7 alone, hE7 alone, and an admixture of Hsp65 and E7 were virtually incapable of stimulating IFN-gamma release. Similar results were obtained with splenocytes prepared from mice obtained from the Charles River Laboratory (Fig. 17A) and from the Jackson Laboratory (Fig. 17B).

Example 18: Stimulation of IFN-Gamma Release by a HspE7 Fusion Protein in Splenocyte Cultures from Mice Having Different Genetic Backgrounds

Experiments similar to those presented in Example 17 were carried out using splenocytes from mice (from Jackson Laboratory) of three different haplotypes: C57BL/6 (H-2^b); Balb/c (H-2^d); and C3HeB/FeJ (H-2^k). The relative effects of the fusion protein on the different splenocyte preparations were similar, although there were differences in the absolute amounts of IFN-gamma released: the observed order being Balb/c (highest; Fig. 18A), C57BL/6 (intermediate; Fig. 18B), and C3HeB/FeJ (lowest; Fig. 18C). As in Example 17, substantially increased IFN-gamma release was induced by HspE7, but not by E7 alone, Hsp65 alone, or an admixture of E7 and Hsp65.

Example 19: Stimulation of IFN-Gamma Release by Fusion Proteins is Independent of the Nature of the Linked Antigen but Requires a Linked Stress Protein Moiety

Experiments were performed as discussed under the previous examples. It was observed that stimulation of naive splenocytes by (h)E7 or Hsp65 (*M. bovis* BCG) produced negligible IFN-gamma release, but that fusion proteins containing E7 and Hsp65 (*M. bovis* BCG) or Hsp40 (*M. tuberculosis*) substantially enhanced IFN-gamma release (Fig. 19). Virtually no induction of IFN-gamma release was mediated by a fusion protein containing E7 and glutathione-S-transferase (GST). When a fusion protein including an ovalbumin fragment and an Hsp (*M. bovis* BCG Hsp65) was tested, high levels of IFN-gamma release were detected. The IFN-gamma release mediated by the HspOVA fusion protein exceeded that resulting from addition of OVA alone to the cell culture. These results demonstrate that the induced release of IFN-gamma is not dependent on the presence of the E7 antigen in the fusion protein, but that other antigens fused to an Hsp can similarly enhance IFN-gamma production.

Example 20: Stimulation of IFN-Gamma Release by E7 Fusion Proteins Having Different Stress Protein Moieties

Experiments were performed as discussed under the previous examples. HPV16 E7 was fused to different Hsps, i.e., *M. tuberculosis* Hsp10 (TB10-E7), *M. bovis* BCG Hsp65 (HspE7), *Streptococcus pneumoniae* Hsp65 (2) (SP65(2)-E7), and *Aspergillus fumigatus* Hsp60 (AF60-E7). Furthermore, in two cases (E7-L-BCG65 and E7-TB71) the Hsp (*M. bovis* BCG Hsp65 and *M. tuberculosis* Hsp71, respectively) was added to the carboxy terminus of the E7 antigen instead of to the amino terminus as in the other fusions.

Additionally, one construct was tested, in which the E7 antigen was linked to the amino terminal one third (residues 1-200) of the *M. bovis* BCG Hsp65 sequence (BCG65(F1)-E7), rather than an intact Hsp. It was observed (Figs. 20A-20B) that stimulation of IFN-gamma release occurred upon exposure of splenocytes to all the different fusion proteins, although differences in the magnitude of the responses were noted. Thus, fusions containing different Hsps, including Hsp65 from different organisms as well as different types of Hsps, were capable of eliciting enhanced IFN-gamma release. Furthermore, fusions containing a stress protein at either the amino terminal end or at the carboxy terminal end of the E7 antigen were active. Finally,

BCG65(F1)-E7, containing amino acids 1-200 of *M. bovis* BCG Hsp65, induced IFN-gamma secretion in a manner similar to the full-length Hsp65 sequence (HspE7).

Example 21: Stimulation of IFN-Gamma Release by HspE7 Fusion Protein in Lymph

5 Node Cell Cultures

To test for their ability to induce IFN-gamma release, various concentrations of the HspE7 proteins (diluted to the desired starting concentration in complete medium, defined as RPMI 1640 with 10% fetal calf serum) were added as replicate samples (3 to 5 replicates) to flat bottom 96-well tissue culture plates. For the cellular component of the assay, three inguinal lymph nodes were aseptically removed from untreated
10 C57BL/6 mice and placed in 5 ml of Hank's balanced salt solution supplemented with 5% fetal calf serum (medium). Following their transfer to a sterile 0.22 micron nylon mesh, a sterile syringe plunger was used to disperse the cells through the mesh. Medium was used to rinse the cells, yielding a pooled, unfractionated single cell
15 suspension. Cells were washed once, resuspended in complete medium and added to wells at 6×10^5 cells/well, to a final volume of 0.2 ml. Cultures were exposed to the HspE7 protein in medium or to medium alone for 72 hours at 37°C in a 5% CO₂ atmosphere. Following incubation, replicate cultures were pooled, cells pelleted by centrifugation and supernatants either measured for IFN-gamma content by ELISA
20 according to the procedure described in Example 17, or frozen immediately at -70°C for later analysis.

Fig. 21 shows the results of the above experiment, comparing induction of IFN-gamma release by lymph node cells and by splenocytes. The fusion protein was found to elicit a release of IFN-gamma in both cell types. The IFN-gamma release elicited by
25 the fusion protein greatly exceeded that induced by Hsp65 alone.

Example 22: Regression of Pre-Established Tumors *in vivo* Induced by Administration of Hsp Fusion Proteins

Human papilloma virus type 16 (HPV16) is an infectious agent associated with
30 the induction of cervical cancer and its premalignant precursor, cervical intraepithelial neoplasia. The following experiments use Hsp - HPV16 E7 fusion proteins of the invention to target immune recognition as part of a strategy to eliminate HPV16 E7-expressing host cells.

The H-2^b murine epithelial cell-derived tumor line, TC-1 (co-transformed with HPV16 E6 and E7 and activated human Ha-*ras*), was obtained from T.C. Wu of Johns Hopkins University (Baltimore, MD). The use of TC-1 cells in assays similar to those used herein is described in PCT patent application WO 99/07860. TC-1 was

5 maintained in complete medium, consisting of: RPMI 1640 (ICN, cat no. 1260354) supplemented with 10% FBS (Hyclone, cat no. SH30071); 2 mM L-Glutamine (ICN, cat no. 16-801-49); 10 mM HEPES (ICN, cat no. 16-884-49); 0.1mM MEM Non Essential Amino Acid Solution (Gibco BRL, cat no. 11140-050); 1 mM MEM Sodium Pyruvate (Gibco BRL, cat no. 11360-070); 50 μ M 2-Mercaptoethanol (Sigma, cat no. M-7522); and 50 mcg/mL Gentamycin Sulfate (Gibco BRL, cat no. 15750-011). The
10 medium was also supplemented with G418 (0.4 mg/mL active, Gibco BRL, cat no. 11811-023) and Hygromycin B (0.2 mg/mL active, Calbiochem, cat no. 400051).

Since the TC-1 cell line was derived from a C57BL/6 mouse, this mouse strain was used as the host in these experiments. Female C57BL/6 mice of approximately 8
15 to 10 weeks of age were purchased from Charles River Canada (St-Constant, Quebec, Canada) and housed using filter top cages (four animals per cage).

TC-1 cells were prepared for implantation as follows. TC-1 cells were seeded at a density of $2 - 5 \times 10^4$ cells/mL and incubated for two to four days until 70 to 90% confluent. Cells were trypsinized using a 30 second exposure to 0.25% Trypsin (10x
20 stock, Gibco cat. no. 1505-065, diluted to 1x with DPBS), then diluted four-fold with supplemented complete medium. Following trypsinization, TC-1 cells were pelleted at 4°C at 1000 rpm (250x g) for 4 minutes, the supernatant removed by aspiration and 30 mL of cold DPBS added. The cells were then pelleted at 4°C at 700 rpm (100x g) for 4 minutes, the supernatant removed by aspiration, and a minimal amount (approx.
25 5 mL) of cold DPBS added. The final cell density for injection was adjusted to 6.5×10^5 viable cells per mL, as measured by the trypan blue dye exclusion method. At least 90% of the cells used for TC-1 inoculations were viable. The cells were stored on ice for immediate injection into mice.

TC-1 cells were implanted as follows. Between 24 to 72 hours prior to
30 implantation, the hind flank of each mouse was shaved. TC-1 cells were prepared as described above and held on ice until injected. All injections were performed within two hours of cell trypsinization. The cells were swirled gently in the centrifuge tube and drawn into a 1 mL syringe (Becton-Dickinson, cat. no. 309602) without a needle.

A 25 gauge needle (Becton-Dickinson, cat. no. 305122) was then attached and any air bubbles were expelled. The shaved skin was raised gently and the needle was inserted bevel side up just beneath the skin surface. Cells (1.3×10^5) were injected in a 0.2 mL volume for all studies. A fresh syringe and needle was used for every fifth injection.

5 Fusion proteins were injected as follows. On treatment days, the fusion proteins HspE7, SP65(2)-E7, AF60-E7, E7-TB71 (shown in Figs. 23A and 23B as E7-MT71), MT40-E7 and TB10-E7 (prepared as described above) were removed from -70°C storage and thawed in a 37°C water bath. Dulbecco's phosphate buffered saline (DPBS) (4°C) was added to obtain the protein concentration desired for injection. The
10 diluted fusion protein was held on ice until drawn into a 1 mL syringe (Becton-Dickinson, cat no. 309602) with a 30 gauge needle (Becton-Dickinson, cat no. 3095106). The same syringe was used to inject 0.2 mL of fusion protein into each mouse within a dose group; the syringe was refitted with a fresh needle for every fifth injection. Mice were injected subcutaneously in the scruff of the neck, as high on the
15 neck as possible.

Tumor incidence (TI) was measured as follows. TI was generally recorded three times per week, beginning eight days after tumor implantation and continuing for eight weeks. Mice were assessed for the presence or absence of subcutaneous tumor by palpation and visual observation of the tumor injection site.

20 Tumor volume was measured as follows. Volumes of palpable subcutaneous tumor nodules were measured beginning on approximately Day 8 post implantation. The two longest orthogonal dimensions were measured using a Fowler Sylvac Ultra-Cal Mark III digital caliper with computerized data collection. Data points were tabulated in a Microsoft Excel spreadsheet. Tumor nodule measurements were
25 extrapolated to mm^3 using the formula $V = W^2 \times L \times 0.5$ (where V represents volume, W represents width and L represents length) and are presented as average tumor volume \pm standard error of the mean. The Student's t test function of Excel (two-tailed, unpaired samples, equal variances) was used to test the significance ($p < 0.05$) of the difference of the means of tumor volumes in each group.

30 Seven different HPV16 E7 fusion proteins linked to various hsps were tested for their ability to regress a tumor *in vivo*.

In the first experiment, C57BL/6 mice (18 per group) were inoculated subcutaneously with 1.3×10^5 TC-1 cells in the right hind flank (Day 0). After 7 days,

groups of mice were treated with 0.2 mL of either DPBS (saline), 115 ug HspE7, 100 ug SP65(2)-E7, or 100 ug AF60-E7. The doses of the two latter proteins were chosen based on the same molar equivalent of E7 contained in HspE7. The mice were monitored for the presence or absence of tumor in addition to tumor volume. The data are represented as percent tumor incidence (TI) per group (Fig. 22A) and tumor volume, expressed as average tumor volume \pm standard error of the mean (Fig. 22B).

As indicated in Fig. 22A, the majority of animals had detectable tumor by Day 8 post implantation and by Day 13 tumor was evident in 94 to 100% of the mice. After this timepoint, TI in all of the mice declined until day 25 when the incidence for the DPBS-treated animals stabilized to approximately 50% for the remainder of the observation period. In contrast, the animals treated with fusion proteins showed a comparatively sharp decline in TI until day 28, when none of the animals had detectable tumor. This complete absence of tumor was observed for the remainder of the observation period for most of these animals. The complete regression of tumor in the animals treated with the fusion proteins was also clearly seen when measured by tumor volume. Figure 22B shows that by day 28, the average tumor volume of the animals treated with the fusion proteins was not detectable. By comparison, the average tumor volume of those animals treated with DPBS rose steadily from day 25 onwards.

In the second experiment, C57BL/6 mice (18 per group) were inoculated subcutaneously with 1.3×10^5 TC-1 cells in the right hind flank (Day 0). After 7 days, groups of mice were treated with 0.2 mL of either DPBS (saline), 100 ug HspE7, 100 ug MT40-E7, 100 ug E7-TB71 (shown in Figs. 23A and 23B as E7-MT71), or 100 ug TB10-E7. The mice were monitored for the presence or absence of tumor in addition to tumor volume. The data are represented as percent tumor incidence (TI) per group (Fig. 23A) and tumor volume, expressed as average tumor volume \pm standard error of the mean (Fig. 23 B).

As in Figure 22A, a majority (approximately 95%) of the animals had visible and palpable tumors on day 8 post tumor implantation (Fig. 23A). By day 19, a decrease in TI was apparent. Following this, a sharp decrease in TI for all of the fusion protein-treated animals was observed such that by day 33, practically all of the animals were tumor-free. In contrast, the TI of the mice treated with DPBS had stabilized to approximately 75%. Fig. 23B shows the average tumor volumes of the mice treated

40

What is claimed is:

1. A method of determining whether a fusion protein stimulates a Th1-like response, the method comprising:

- 5 (a) providing a cell sample comprising naive lymphocytes *in vitro*;
- (b) providing a fusion protein comprising (i) a heat shock protein (Hsp) or a fragment thereof at least eight amino acid residues in length, fused to (ii) a heterologous polypeptide at least eight amino acid residues in length;
- (c) contacting the cell sample with the fusion protein; and
- 10 (d) determining whether the fusion protein stimulates a Th1-like response in the cell sample.

2. The method of claim 1, wherein the Hsp is selected from the group consisting of Hsp65, Hsp40, Hsp10, Hsp60, and Hsp71.

15

3. The method of claim 2, wherein the fusion protein comprises a polypeptide selected from the group consisting of Hsp65, Hsp40, Hsp10, Hsp60, and Hsp71.

20

4. The method of claim 1, wherein the fusion protein comprises amino acids 1-200 of Hsp65 of *Mycobacterium bovis*.

25

5. The method of claim 1, wherein the heterologous polypeptide comprises a sequence identical to at least eight consecutive amino acids of (i) a protein of a human pathogen or (ii) a tumor associated antigen.

30

6. The method of claim 1, wherein the heterologous polypeptide comprises a sequence identical to at least eight consecutive amino acids of a protein of a human virus.

7. The method of claim 6, wherein the virus is selected from the group consisting of human papilloma virus (HPV), herpes simplex virus (HSV), hepatitis B

virus (HBV), hepatitis C virus (HCV), cytomegalovirus (CMV), Epstein-Barr virus (EBV), influenza virus, measles virus, and human immunodeficiency virus (HIV).

8. The method of claim 7, wherein the heterologous polypeptide comprises
5 HPV E6.

9. The method of claim 7, wherein the heterologous polypeptide comprises
HPV E7.

10. The method of claim 1, wherein the heterologous polypeptide comprises
10 HPV 16 E7 or a fragment thereof at least eight amino acid residues in length.

11. The method of claim 1, wherein the heterologous polypeptide comprises
15 HPV 16 E6 or a fragment thereof at least eight amino acid residues in length.

12. The method of claim 10, wherein the fusion protein comprises
Mycobacterium bovis Hsp65 and HPV 16 E7.

13. The method of claim 1, wherein the cell sample comprises cells derived
20 from a spleen, lymph node, peripheral blood, bone marrow, thymus, lung, respiratory
tract, or anogenital mucosa

14. The method of claim 1, wherein the cell sample comprises splenocytes
or lymph node cells.

15. The method of claim 1, wherein the detecting step comprises detecting
25 IFN-gamma produced by the cell sample.

16. The method of claim 1, comprising the further steps of
30 (e) providing a second cell sample comprising naive lymphocytes;
(f) contacting the second cell sample with a second fusion protein; and
(g) determining whether the second fusion protein stimulates a Th1-like
response in the second cell sample,

wherein the first fusion protein comprises the sequence of a full-length, naturally occurring Hsp, and the second fusion protein comprises at least eight amino acids but less than all of the sequence of a naturally occurring Hsp.

- 5 17. A method of screening a compound, the method comprising:
- (a) providing a cell sample comprising naive lymphocytes *in vitro*;
- (b) providing a fusion protein comprising (i) a Hsp or a fragment thereof at least eight amino acid residues in length, fused to (ii) a heterologous polypeptide at least eight amino acid residues in length;
- 10 (c) contacting the cell sample with the compound and the fusion protein; and
- (d) determining whether the cell sample exhibits a Th1-like response following the contacting step,

 wherein a decrease in the Th1-like response in the presence of the compound
15 compared to in the absence of the compound indicates that the compound inhibits a Th1-like response by the cell sample.

 18. The method of claim 17, wherein the determining step comprises detecting IFN-gamma produced by the cell sample.

20

 19. The method of claim 17, wherein the cell sample comprises cells derived from a spleen, lymph node, peripheral blood, bone marrow, thymus, lung, respiratory tract, or anogenital mucosa

25 20. The method of claim 17, wherein the cell sample comprises splenocytes or lymph node cells.

 21. The method of claim 17, wherein the Hsp is selected from the group consisting of Hsp65, Hsp40, Hsp10, Hsp60, and Hsp71.

30

 22. The method of claim 21, wherein the fusion protein comprises a polypeptide selected from the group consisting of Hsp65, Hsp40, Hsp10, Hsp60, and Hsp71.

23. The method of claim 17, wherein the heterologous polypeptide comprises HPV E6.

5 24. The method of claim 17, wherein the heterologous polypeptide comprises HPV E7.

25. The method of claim 17, wherein the fusion protein comprises *Mycobacterium bovis* Hsp65 and HPV 16 E7.

10 26. A method of screening a compound, the method comprising:
(a) providing a cell sample comprising naive lymphocytes *in vitro*;
(b) providing a fusion protein comprising (i) a Hsp or a fragment thereof at
least eight amino acid residues in length, fused to (ii) a heterologous polypeptide at
15 least eight amino acid residues in length;

(c) contacting the cell sample with the compound and the fusion protein;
and

(d) determining whether the cell sample exhibits a Th1-like response
following the contacting step,

20 wherein an increase in the Th1-like response in the presence of the compound
compared to in the absence of the compound indicates that the compound promotes a
Th1-like response by the cell sample.

27. The method of claim 26, wherein the determining step comprises
25 detecting IFN-gamma produced by the cell sample.

28. The method of claim 26, wherein the cell sample comprises cells derived
from a spleen, lymph node, peripheral blood, bone marrow, thymus, lung, respiratory
tract, or anogenital mucosa

30 29. The method of claim 26, wherein the cell sample comprises splenocytes
or lymph node cells.

30. The method of claim 26, wherein the Hsp is selected from the group consisting of Hsp65, Hsp40, Hsp10, Hsp60, and Hsp71.

31. The method of claim 30, wherein the fusion protein comprises a polypeptide selected from the group consisting of Hsp65, Hsp40, Hsp10, Hsp60, and Hsp71.

32. The method of claim 26, wherein the heterologous polypeptide comprises HPV E6.

33. The method of claim 26, wherein the heterologous polypeptide comprises HPV E7.

34. The method of claim 26, wherein the fusion protein comprises *Mycobacterium bovis* BCG Hsp65 and HPV 16 E7.

35. A method of determining whether a hybrid compound stimulates a Th1-like response, the method comprising:

- (a) providing a cell sample comprising naive lymphocytes *in vitro*;
- (b) providing a hybrid compound that is non-naturally occurring and comprises (i) a non-peptide compound having a molecular weight of less than 1,500, covalently linked to (ii) a polypeptide of at least eight amino acids in length, wherein the hybrid compound is made by covalently linking the non-peptide compound to the polypeptide;
- (c) contacting the cell sample with the hybrid compound; and
- (d) determining whether the hybrid compound stimulates a Th1-like response in the cell sample.

36. The method of claim 35, wherein the non-peptide compound has a molecular weight of at least 100.

37. A method of determining whether a hybrid compound stimulates a Th1-like response, the method comprising:

- (a) producing a hybrid compound by covalently linking a non-peptide compound to a polypeptide of at least eight amino acids in length;
- (b) providing a cell sample comprising naive lymphocytes *in vitro*;
- (c) contacting the cell sample with the hybrid compound; and
- 5 (d) determining whether the hybrid compound stimulates a Th1-like response in the cell sample.

38. The method of claim 37, wherein the non-peptide compound has a molecular weight between 100 and 1,500.

10

39. A method of determining whether a fusion protein stimulates a Th1-like response, the method comprising:

- (a) providing a cell sample comprising naive lymphocytes *in vitro*;
- (b) providing a fusion protein comprising (i) a first polypeptide at least eight
- 15 amino acids in length, fused to (ii) a second polypeptide at least eight amino acids in length;
- (c) contacting the cell sample with the fusion protein; and
- (d) detecting a Th1-like response exhibited by the cell sample following the contacting step.

20

40. The method of claim 39, wherein the detected Th1-like response is greater than a Th1-like response exhibited by a second cell sample comprising naive lymphocytes when the second cell sample is contacted with either the first polypeptide, the second polypeptide, or a mixture of the first polypeptide and the second

25 polypeptide.

41. The method of claim 40, wherein the detected Th1-like response is at least two times greater than the Th1-like response exhibited by the second cell sample.

30

42. The method of claim 40, wherein the detected Th1-like response is at least five times greater than the Th1-like response exhibited by the second cell sample.

43. A fusion protein comprising (i) a Hsp10 protein or a fragment thereof at least eight amino acid residues in length, and (ii) a heterologous polypeptide at least eight amino acids in length.

5 44. The fusion protein of claim 43, comprising a Hsp10 protein.

45. The fusion protein of claim 44, wherein the Hsp10 protein is a mycobacterial protein.

10 46. The fusion protein of claim 45, comprising the *Mycobacterium tuberculosis* Hsp10 protein.

47. The fusion protein of claim 43, wherein the heterologous polypeptide comprises a sequence identical to at least eight consecutive amino acids of a protein of
15 a human virus.

48. The fusion protein of claim 47, wherein the human virus is HPV.

49. The fusion protein of claim 48, wherein the heterologous polypeptide
20 comprises HPV16 E7.

50. A fusion protein comprising (i) a Hsp40 protein or a fragment thereof at least eight amino acid residues in length, and (ii) a heterologous polypeptide at least eight amino acids in length.

25

51. The fusion protein of claim 50, comprising a Hsp40 protein.

52. The fusion protein of claim 51, wherein the Hsp40 protein is a mycobacterial protein.

30

53. The fusion protein of claim 52, comprising the *Mycobacterium tuberculosis* Hsp40 protein.

54. The fusion protein of claim 50, wherein the heterologous polypeptide comprises a sequence identical to at least eight consecutive amino acids of a protein of a human virus.

5 55. The fusion protein of claim 54, wherein the human virus is HPV.

56. The fusion protein of claim 55, wherein the heterologous polypeptide comprises HPV16 E7.

10 57. A fusion protein comprising (i) a Hsp71 protein or a fragment thereof at least eight amino acid residues in length, and (ii) a heterologous polypeptide at least eight amino acids in length.

58. The fusion protein of claim 57, comprising a Hsp71 protein.

15

59. The fusion protein of claim 58, wherein the Hsp71 protein is a mycobacterial protein.

20 60. The fusion protein of claim 59, comprising the *Mycobacterium tuberculosis* Hsp71 protein.

61. The fusion protein of claim 57, wherein the heterologous polypeptide comprises a sequence identical to at least eight consecutive amino acids of a protein of a human virus.

25

62. The fusion protein of claim 61, wherein the human virus is HPV.

63. The fusion protein of claim 62, wherein the heterologous polypeptide comprises HPV16 E7.

30

64. A method of determining whether a compound stimulates a Th1-like response, the method comprising:

(a) providing a cell sample comprising naive lymphocytes *in vitro*;

- (b) providing a compound;
- (c) contacting the cell sample with the compound; and
- (d) detecting a Th1-like response exhibited by the cell sample following the contacting step.

5

1/37

3/1
 atg gcc aag aca att gcg tac gac gaa gag
 M A K T I A Y D E E
 63/21
 gcc ctg gcc gat gog gta aag gtg aca ttg
 A L A D A V K V T L
 123/41
 aag aag tgg ggt gcc ccc acg atc acc aac
 K K W G A P T I T N
 183/61
 ctg gag gat cgg tac gag aag atc ggc gcc
 L E D P Y E K I G A
 243/81
 gat gac gtc gcc ggt gac ggc acc acg acg
 D D V A G D G T T T
 303/101
 gag gcc ctg cgc aac gtc gcg gcc gcc gcc
 E G L R N V A A G A
 363/121
 aag gcc gtg gag aag gtc acc gag acc ctg
 K A V E K V T E T L
 423/141
 gag cag att gcg gcc acc gca gog att tgg
 E Q I A A T A A I S
 483/161
 gcc gag gcg atg gac aag gtg ggc aac gag
 A E A M D K V G N E
 543/181
 ttt ggg ctg cag ctg gag ctg acc gag ggt
 F G L Q L E L T E G
 603/201
 tac ttc gtg acc gac cgg gag cot cag gag
 Y F V T D P E R Q E
 663/221
 gtc acc tcc aag gtg tcc act gtc aag gat
 V S S K V S T V K D
 723/241
 gcc ggt aag cgg ctg ctg atc atc gcc gag
 A G K P L L I I A E
 783/261
 gtc gtc aac aag atc cgc ggc acc ttc aag
 V V N K I R G T F K
 843/281
 gac cgc cgc aag gcg atg ctg cag gat atg
 D R R K A M L Q D M
 903/301
 gaa gag gtc gcc ctg acg ctg gag aac gcc
 E E V G L T L E N A
 963/321
 gtc gtg gtc acc aag gac gag acc acc atc
 V V V T K D E T T I
 1023/341
 gcc gga cga gtg gcc cag atc cgc cag gag
 A G R V A Q I R Q E
 1083/361
 gag aag ctg cag gag cgg ctg gcc aag ctg
 E K L Q E R L A K L
 1143/381
 gcc gcc acc gag gtc gaa ctg aag gag cgc
 A A T E V E L K E R
 1203/401
 gcc aag gcc gcc gtc gag gag gcc atc gtc
 A K A A V E E G I V
 33/11
 gcc cgt cgc ggc ctg gag cgg gcc ttg aac
 A R R G L E R G L N
 93/31
 gcc ccc aag gcc cgc aac gtc gtc ctg gaa
 G P K G R N V V L E
 153/51
 gat ggt gtg tcc atc gcc aag gag atc gag
 D G V S I A K E I E
 213/71
 gag ctg gtc aaa gag gta gcc aag aag acc
 E L V K E V A K K T
 273/91
 gcc acc gtg ctg gcc cag gcg ttg gtt cgc
 A T V L A Q A L V R
 333/111
 aac cgg ctg ggt ctg aaa cgc gcc atc gaa
 N P L G L K R G I E
 393/131
 ctg aag gcc gcc aag gag gtc gag acc aag
 L K G A K E V E T K
 453/151
 gcc ggt gac cag tcc atc ggt gac ctg atc
 A G D Q S I G D L I
 513/171
 gcc gtc atc acc gtc gag gag tcc aac acc
 G V I T V E E S N T
 573/191
 atg cgg ttc gac aag gcc tac atc tgg ggg
 M R F D K G Y I S G
 633/211
 gcc gtc ctg gag gac ccc tac atc ctg ctg
 A V L E D P Y I L L
 693/231
 ctg ctg cgg ctg ctg gag aag gtc atc gga
 L L P L L E K V I G
 753/251
 gac gtc gag gcc gag gcc ctg tcc acc ctg
 D V E G E A L S T L
 813/271
 tgg gtg gcc gtc aag ggt ccc gcc ttc gcc
 S V A V K A P G F G
 873/291
 gcc att ctg acc ggt ggt cag gtg atc acc
 A I L T G G Q V I S
 933/311
 gac ctg tgg ctg cta gcc aag gcc cgc aag
 D L S L L G K A R K
 993/331
 gtc gag gcc gcc ggt gac acc gac gcc atc
 V E G A G D T D A I
 1053/351
 atc gag aac agc gac tcc gac tac gac cgt
 I E N S D S D Y D R
 1113/371
 gcc ggt ggt gtc gcg gtc atc aag gcc ggt
 A G G V A V I K A G
 1173/391
 aag cac cgc atc gag gat gcg gtt cgc aat
 K H R I E D A V R N
 1233/411
 gcc ggt ggt ggt gtc acg ctg ttg caa gcc
 A G G G V T L L Q A

FIG. 1A

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1263/421
GCC CCG ACC CTG GAC GAG CTG AAG CTC GAA GGC GAC GAG GCG ACC GGC GGC AAC ATC GTG
A P T L D E L K L E G D E A T G A N I V

1323/441
AAG GTG GCG CTG GAG GCC CCG CTG AAG CAG ATC GCC TTC AAC TCC GGG CTG GAG CCG GGC
K V A L E A P L K Q I A F N S G L E P G

1383/461
GTG GTG GCC GAG AAG GTG CCG AAC CTG CCG GCT GGC CAC GGA CTG AAC GCT CAG ACC GGT
V V A E K V R N L P A G H G L N A Q T G

1443/481
GTC TAC GAG GAT CTG CTC GCT GCC GGC GTT GCT GAC CCG GTC AAG GTG ACC COT TCG GCG
V Y E D L L A A G V A D P V K V T R S A

1503/501
CTG CAG AAT GCG GCG TCC ATC GCG GGG CTG TTC CTG ACC ACC GAG GCC GTC GTT GCC GAC
L Q N A A S I A G L F L T T E A V V A D

1563/521
AAG CCG GAA AAG GAG AAG GCT TCC GTT CCC GGT GGC GGC GAC ATG GGT GGC ATG GAT TTC
K P E K E K A S V P G G G D M G G M D F

1623/541
TGA
*

1293/431
GCC GAC GAG GCG ACC GGC GGC AAC ATC GTG
G D E A T G A N I V

1353/451
ATC GCC TTC AAC TCC GGG CTG GAG CCG GGC
I A F N S G L E P G

1413/471
GCT GGC CAC GGA CTG AAC GCT CAG ACC GGT
A G H G L N A Q T G

1473/491
GCT GAC CCG GTC AAG GTG ACC COT TCG GCG
A D P V K V T R S A

1533/511
TTC CTG ACC ACC GAG GCC GTC GTT GCC GAC
F L T T E A V V A D

1593/531
GGT GGC GGC GAC ATG GGT GGC ATG GAT TTC
G G G D M G G M D F

FIG. 1B

3/37

3/1		33/11
ATG GAT GGA GAT ACA CCT ACA TTG CAT GAA TAT ATG TTA GAT TTG CAA CCA GAG ACA ACT		
M D G D T P T L H E Y M L D L Q P E T T		
63/21		93/31
GAT CTC TAC TGT TAT GAG CAA TTA AAT GAC AGC TCA GAG GAG GAG GAT GAA ATA GAT GGT		
D L Y C Y E Q L N D S S E E E D E I D G		
123/41		153/51
CCA GCT GGA CAA GCA GAA CCG GAC AGA GGC CAT TAC AAT ATT GTA ACC TTT TGT TGC AAG		
P A G Q A E P D R A H Y N I V T F C C K		
183/61		213/71
TGT GAC TCT ACG CTT CCG TTG TGC GTA CAA AGC ACA CAC GTA GAC ATT CGT ACT TTG GAA		
C D S T L R L C V Q S T H V D I R T L E		
243/81		273/91
GAC CTG TTA ATG GGC ACA CTA GGA ATT GTG TGC CCC ATC TGT TCT CAG AAA CCA TAA		
D L L M G T L G I V C P I C S Q K P *		

FIG. 2

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3/1	ATG GGC AGC AGC CAT CAT CAT CAT CAT CAC	33/11	AGC AGC GGC CTG GTG CCG CGC GGC AGC CAT
63/21	M G S S H H H H H H S S G L V P R G S H	93/31	
123/41	ATG gct agc ATG CAT GGA GAT ACA CCT ACA TTG CAT GAA TAT ATG TTA GAT TTG CAA CCA	153/51	M A S M H G D T P T L H E Y M L D L Q P
183/61	GAG ACA ACT GAT CTC TAC TGT TAT GAG CAA TTA AAT GAC AGC TCA GAG GAG GAG GAT GAA	213/71	E T T D L Y C Y E Q L N D S S E E E D E
243/81	ATA GAT GGT CCA GCT GGA CAA GCA GAA CCG GAC AGA GGC CAT TAC AAT ATT GTA ACC TTT	273/91	I D G P A G Q A E P D R A H Y N I V T F
303/101	TGT TGC AAG TGT GAC TCT ACG CTT CGG TTG TGC GTA CAA AGC ACA CAC GTA GAC ATT CGT	333/111	C C K C D S T L R L C V Q S T H V D I R
363/121	ACT TTG GAA GAC CTG TTA ATG GGC ACA CTA GGA ATT GTG TGC CCC ATC TGT TCT CAG AAA		T L E D L L M G T L G I V C P I C S Q K
	CCA TAA		
	P *		

FIG. 3

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3/1
 atg gcc aag aca att cgc tac gac gaa gag gcc cgt cgc ggc ctc gag cgg gcc ttg aac
 M A K T I A Y D E E A R R G L E R G L N
 63/21
 gcc ctc gcc gat cgc gta aag gtg aca ttg gcc ccc aag ggc cgc aac gtc gtc ctg gaa
 A L A D A V K V T L G P K G R N V V L E
 123/41
 aag aag tgg ggt gcc ccc acg atc acc aac gat ggt gtg tcc atc gcc aag gag atc gag
 K K W G A P T I T N D G V S I A K E I E
 183/61
 ctg gag gat cgc tac gag aag atc ggc gcc gag ctg gtc aaa gag gta gcc aag aag acc
 L E D P Y E K I G A E L V K E V A K K T
 243/81
 gat gac gtc gcc ggt gac ggc acc acg acg gcc acc gtg ctg gcc cag ggc ttg gtt cgc
 D D V A G D G T T T A T V L A Q A L V R
 303/101
 gag gcc ctg cgc aac gtc cgc gcc gcc gcc aac cgc ctc ggt ctc aaa cgc gcc atc gaa
 E G L R N V A A G A N P L G L K R G I E
 363/121
 aag gcc gtg gag aag gtc acc gag acc ctg ctg aag ggc gcc aag gag gtc gag acc aag
 K A V E K V T E T L L K G A K E V E T K
 423/141
 gag cag att cgc gcc acc gca cgc att tcc gcc ggt gac cag tcc atc ggt gac ctg atc
 E Q I A A T A A I S A G D Q S I G D L I
 483/161
 gcc gag cgc atg gac aag gtg ggc aac gag ggc gtc atc acc gtc gag gag tcc aac acc
 A E A M D K V G N E G V I T V E E S N T
 543/181
 ttt ggc ctg cag ctc gag ctc acc gag ggt atg cgc ttc gac aag ggc tac atc tcc ggg
 F G L Q L E L T E G M R F D K G Y I S G
 603/201
 tac ttc gtg acc gac cgc gag cgt cag gag cgc gtc ctg gag gac ccc tac atc ctg ctg
 Y F V T D P E R Q E A V L E D P Y I L L
 663/221
 gtc acc tcc aag gtg tcc act gtc aag gat ctg ctg cgc ctg ctc gag aag gtc atc gga
 V S S K V S T V K D L L P L L E K V I G
 723/241
 gcc ggt aag cgc ctg ctg atc atc gcc gag gac gtc gag ggc gag cgc ctg tcc acc ctg
 A G K P L L I I A E D V E G E A L S T L
 783/261
 gtc gtc aac aag atc cgc ggc acc ttc aag tcc gtg cgc gtc aag gct ccc gcc ttc gcc
 V V N K I R G T F K S V A V K A P G F G
 843/281
 gac cgc cgc aag ggc atg ctg cag gat atg gcc att ctc acc ggt ggt cag gtg atc agc
 D R R K A M L Q D M A I L T G G Q V I S
 903/301
 gaa gag gtc ggc ctg acg ctg gag aac gcc gac ctg tcc ctg cta gcc aag gcc cgc aag
 E E V G L T L E N A D L S L L G K A R K
 963/321
 gtc gtg gtc acc aag gac gag acc acc atc gtc gag gcc gcc ggt gac acc gac gcc atc
 V V V T K D E T T I V E G A G D T D A I
 1023/341
 gcc gga cga gtg gcc cag atc cgc cag gag atc gag aac agc gac tcc gac tac gac cgt
 A G R V A Q I R Q E I E N S D S D Y D R
 1083/361
 gag aag ctg cag gag cgc ctg gcc aag ctg gcc ggt ggt gtc cgc gtg atc aag gcc ggt
 E K L Q E R L A K L A G G V A V I K A G
 1143/381
 gcc gcc acc gag gtc gaa ctc aag gag cgc aag cac cgc atc gag gat ggc gtt cgc aat
 A A T E V E L K E R K H R I E D A V R N
 1203/401
 gcc aag gcc gcc gtc gag gag gcc atc gtc gcc ggt ggc ggt gtg acg ctg ttg caa gcc
 A K A A V E E G I V A G G G V T L L Q A

FIG. 4A

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1263/421      1293/431
GCC CCG ACC CTG GAC GAG CTG AAG CTC GAA GGC GAC GAG GCG ACC GGC GCC AAC ATC GTG
A P T L D E L K L E G D E A T G A N I V
1323/441      1353/451
AAG GTG GCG CTG GAG GCC CCG CTG AAG CAG ATC CCC TTC AAC TCC GGG CTG GAG CCG GGC
K V A L E A P L K Q I A F N S G L E P G
1383/461      1413/471
GTG GTG GCC GAG AAG GTG CCG AAC CTG CCG GCT GGC CAC GGA CTG AAC GCT CAG ACC GGT
V V A E K V R N L P A G H G L N A Q T G
1443/481      1473/491
GTC TAC GAG GAT CTG CTC GCT GCC GGC GTT GCT GAC CCG GTC AAG GTG ACC CGT TCG GCG
V Y E D L L A A G V A D P V K V T R S A
1503/501      1533/511
CTG CAG AAT GCG GCG TCC ATC GCG GGC CTG TTC CTG ACC ACC GAG GCC GTC GTT GCC GAC
L Q N A A S I A G L F L T T E A V V A D
1563/521      1593/531
AAG CCG GAA AAG GAG AAG GCT TCC GTT CCC GGT GGC GGC GAC ATG GGT GGC ATG GAT TTC
K P E K E K A S V P G G G D M G G M D F
1623/541      1653/551
cat atg cat gga gat aca cct aca ttg cat gaa tat atg tta gat ttg caa cca gag aca
H M H G D T P T L H E Y M L D L Q P E T
1683/561      1713/571
act gat ctc tac tgt tat gag caa tta aat gac agc tca gag gag gag gat gaa ata gat
T D L Y C Y E Q L N D S S E E E D E I D
1743/581      1773/591
ggt cca gct gga caa gca gaa ccg gac aga gcc cat tac aat att gta acc ttt tgt tgc
G P A G Q A E P D R A H Y N I V T F C C
1803/601      1833/611
aag tgt gac tct acg ctt cgg ttg tgc gta caa agc aca cac gta gac att cgt act ttg
K C D S T L R L C V Q S T H V D I R T L
1863/621      1893/631
gaa gac ctg tta atg ggc aca cta gga att gtg tgc ccc atc tgt tct cag aaa cca TAA
E D L L M G T L G I V C P I C S Q K P *

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FIG. 4B

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1303/421      1333/431
GAG CAA TTA AAT GAC AGC TCA GAG GAG GAG GAT GAA ATA GAT GGT CCA GCT GGA CAA GCA
E Q L N D S S E E E D E I D G P A G Q A
1363/441      1393/451
GAA CCG GAC AGA GCC CAT TAC AAT ATT GTA ACC TTT TGT TGC AAG TGT GAC TCT ACG CTT
E P D R A H Y N I V T F C C K C D S T L
1423/461      1453/471
CGG TTG TGC GTA CAA AGC ACA CAC GTA GAC ATT CGT ACT TTG GAA GAC CTG TTA ATG GGC
R L C V Q S T H V D I R T L E D L L M G
1483/481      1513/491
ACA CTA GGA ATT GTG TGC CCC ATC TGT TCT CAG AAA CCA TAG
T L G I V C P I C S Q K P *

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FIG. 5B

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43/1	ATG GCC CAA AGG GAA TGG GTC GAA AAA GAC	73/11	TTC TAC CAG GAG CTG GGC GTC TCC TCT GAT
M A Q R E W V E K D		F Y Q E L G V S S D	
103/21	GCC AGT CCT GAA GAG ATC AAA CGT GCC TAT	133/31	CGG AAG TTG GCG CGC GAC CTG CAT CCG GAC
A S P E E I K R A Y		R K L A R D L H F D	
163/41	GCG AAC CCG GGC AAC CCG GCC GCC GGC GAA	193/51	CGG TTC AAG GCG GTT TCG GAG GCG CAT AAC
A N P G N P A A G E		R F K A V S E A H N	
223/61	GTG CTG TCG GAT CCG GCC AAG CGC AAG GAG	253/71	TAC GAC GAA ACC CGC CGC CTG TTC GCC GGC
V L S D P A K R K E		Y D E T R R L F A G	
283/81	GGC GGG TTC GGC GGC CGT CGG TTC GAC AGC	313/91	GGC TTT GGG GGC GGG TTC GGC GGT TTC GGG
G G F G G R R F D S		G F G G G F G G F G	
343/101	GTC GGT GGA GAC GGC GCC GAG TTC AAC CTC	373/111	AAC GAC TTG TTC GAC GCC GCC AGC CGA ACC
V G G D G A E F N L		N D L F D A A S R T	
403/121	GGC GGT ACC ACC ATC GGT GAC TTG TTC GGT	433/131	GGC TTG TTC GGA CGC GGT GGC AGC GCC CGT
G G T T I G D L F G		G L F G R G G S A R	
463/141	CCC AGC CGC CCG CGA CGC GGC AAC GAC CTG	493/151	GAG ACC GAG ACC GAG TTG GAT TTC GTG GAG
P S R P R R G N D L		E T E T E L D F V E	
523/161	GCC GCC AAG GGC GTG GCG ATG CCG CTG CGA	553/171	TTA ACC AGC CCG GCG CCG TGC ACC AAC TGC
A A K G V A M P L R		L T S P A P C T N C	
583/181	CAT GGC AGC GGG GCC CGG CCA GGC ACC AGC	613/191	CCA AAG GTG TGT CCC ACT TGC AAC GGG TCG
H G S G A R P G T S		P K V C P T C N G S	
643/201	GGC GTG ATC AAC CGC AAT CAG GGC GCG TTC	673/211	GGC TTC TCC GAG CCG TGC ACC GAC TGC CGA
G V I N R N Q G A F		G F S E P C T D C R	
703/221	GGT AGC GGC TCG ATC ATC GAG CAC CCC TGC	733/231	GAG GAG TGC AAA GGC ACC GGC GTG ACC ACC
G S G S I I E H P C		E E C K G T G V T T	
763/241	CGC ACC CGA ACC ATC AAC GTG CCG ATC CCG	793/251	CCC GGT GTC GAG GAT GGG CAG CGC ATC CGG
R T R T I N V R I P		P G V E D G Q R I R	
823/261	CTA GCC GGT CAG GGC GAG GOC GGG TTG CGC	853/271	GGC GCT CCC TCG GGG GAT CTC TAC GTG ACG
L A G Q G E A G L R		G A P S G D L Y V T	
883/281	GTG CAT CTG CCG CCC GAC AAG ATC TTC GGC	913/291	CGC GAC GGC GAC GAC CTC ACC GTC ACC GTT
V H V R P D K I F G		R D G D D L T V T V	
943/301	CCG GTC AGC TTC ACC GAA TTG GCT TTG GGC	973/311	TCG ACG CTG TCG GTG CCT ACC CTG GAC GGC
P V S F T E L A L G		S T L S V P T L D G	
1003/321	ACG GTC GGG GTC CCG GTG CCC AAA GGC ACC	1033/331	GCT GAC GGC CGC ATT CTG CGT GTG CGC GGA
T V G V R V P K G T		A D G R I L R V R G	
1063/341	CGC GGT GTG CCC AAG CGC AGT GGG GGT AGC	1093/351	GGC GAC CTA CTT GTC ACC GTG AAG GTG GCC
R G V P K R S G G S		G D L L V T V K V A	
1123/361	GTG CCG CCC AAT TTG GCA GGC GCC GCT CAG	1153/371	GAA GCT CTG GAA GCC TAT GCG GCG GCG GAG
V P P N L A G A A Q		E A L E A Y A A A E	
1183/381	CGG TCC AGT GGT TTC AAC CCG CCG GCC GGA	1213/391	TGG GCA GGT AAT CGC ATG CAT GGA GAT ACA
R S S G F N P R A G		W A G N R M H G D T	
1243/401	CCT ACA TTG CAT GAA TAT ATG TTA GAT TTG	1273/411	CAA CCA GAG ACA ACT GAT CTC TAC TGT TAT
P T L H E Y M L D L		Q P E T T D L Y C Y	

FIG. 5A

SUBSTITUTE SHEET (RULE 26)

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3/1
 ATG GGC AGC AGC CAT CAT CAT CAT CAT CAC AGC AGC GGC CTG GTG CCG CGC GGC AGC CAT
 M G S S H H H H H H S S G L V P R G S H
 53/21
 ATG gct agc atg ggc tcc atc ggc gca gca agc atg gaa ttt tgt ttt gat gta ttc aag
 M A S M G S I G A A S M E F C F D V F K
 123/41
 gag ctc aaa gtc cac cat gcc aat gag aac atc ttc tac tgc ccc att gcc atc atg tca
 E L K V H H A N E N I F Y C F I A I M S
 183/61
 gct cta gcc atg gta tac ctg ggt gca aaa gac agc acc agg aca cag ata aat aag gtt
 A L A M V Y L G A K D S T R T Q I N K V
 243/81
 gtt cgc ttt gat aaa ctt cca gga ttc gga gac agt att gaa gct cag tgt ggc aca tct
 V R F D K L P G F G D S I E A Q C G T S
 303/101
 gta aac gtt cac tct tca ctt aga gac atc ctc aac caa atc acc aaa cca aat gat gtt
 V N V H S S L R D I L N Q I T K P N D V
 363/121
 tat ctg ttc agc ctt gcc agt aga ctt tat gct gaa gag aga tac cca atc ctg cca gaa
 Y S F S L A S R L Y A E E R Y P I L P E
 423/141
 tac ttg cag tgt gtg aag gaa ctg tat aga gga ggc ttg gaa cct atc aac ttt caa aca
 Y L Q C V K E L Y R G G L E P I N F Q T
 483/161
 gct gca gat caa gcc aga gag ctc atc aat tcc tgg gta gaa agt cag aca aat gga att
 A A D Q A R E L I N S W V E S Q T N G I
 543/181
 atc aga aat gtc ctt cag cca agc tcc gtg gat tct caa act gca atg gtt ctg gtt aat
 I R N V L Q P S S V D S Q T A M V L V N
 603/201
 gcc att gtc ttc aaa gga ctg tgg gag aaa aca ttt aag gat gaa gac aca caa gca atg
 A I V F K G L W E K T F K D E D T Q A M
 663/221
 cct ttc aga gtg act gag caa gaa agc aaa cct gtg cag atg atg tac cag att ggt tta
 P F R V T E Q E S K P V Q M M Y Q I G L
 723/241
 ttt aga gtg gca tca atg gct tct gag aaa atg aag atc ctg gag ctt cca ttt gcc agt
 P R V A S M A S E K M K I L E L P F A S
 783/261
 ggg aca atg agc atg ttg gtg ctg ttg cct gat gaa gtc tca ggc ctt gag cag ctt gag
 G T M S M L V L L P D E V S G L E Q L E
 843/281
 agt ata atc aac ttt gaa aaa ctg act gaa tgg acc agt tct aat gtt atg gaa gag agg
 S I I N F E K L T E W T S S N V M E S R
 903/301
 aag atc aaa gtg tac tta cct cgc atg aag atg gag gaa aaa tac aac ctc aca tct gtc
 K I K V Y L P R M K M E E K Y N L T S V
 963/321
 tta atg gct atg ggc att act gac gtg ttt agc tct tca gcc aat ctg tct ggc atc tcc
 L M A M G I T D V F S S S A N L S G I S
 1023/341
 tca gca gag agc ctg aag ata tct caa gct gtc cat gca gca cat gca gaa atc aat gaa
 S A E S L K I S Q A V H A A H A E I N E
 1083/361
 gca ggc aga gag gtg gta ggg tca gca gag gct gga gtg gat gct gca agc gtc tct gaa
 A G R E V V G S A E A G V D A A S V S E
 1143/381
 gaa ttt agg gct gac cat cca ttc ctc ttc tgt atc aag cac atc gca acc aac gcc gtt
 E F R A D H P F L F C I K H I A T N A V
 1203/401
 ctc ttc ttt ggc aga tgt gtt gga tcc taa
 L P P G R C V G S

FIG. 6

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3/1
 atg ggc agc agc cat cat cat cat cat cac
 M G S S H H H H H H S S G L V P R G S H
 63/21
 atg gcc aag aca att gcg tac gac gaa gag
 M A K T I A Y D E E A R R G L E R G L N
 123/41
 gcc ctg gcc gat gcg gta aag gtg aca ttg
 A L A D A V K V T L G P K G R N V V L E
 183/61
 aag aag tgg ggt gcc ccc acg atc acc aac
 K K W G A P T I T N D G V S I A K E I E
 243/81
 ctg gag gat ccg tac gag aag atc ggc gcc
 L E D P Y E K I G A E L V K E V A K K T
 303/101
 gat gac gtc gcc ggt gac ggc acc acg acg
 D D V A G D G T T T A T V L A Q A L V R
 363/121
 gag gcc ctg cgc aac gtc gcg gcc gcc gcc
 E G L R N V A A G A N P L G L K R G I E
 423/141
 aag gcc gtg gag aag gtc acc gag acc ctg
 K A V E K V T E T L L K G A K E V E T K
 483/161
 gag cag att gcg gcc acc gca gcg att tgg
 E Q I A A T A A I S A G D Q S I G D L I
 543/181
 gcc gag gcg atg gac aag gtg gcc aac gag
 A E A M D K V G N E G V I T V E E S N T
 603/201
 ttt ggc ctg cag ctg gag ctg acc gag ggt
 F G L Q L E L T E G M R P D K G Y I S G
 663/221
 tac ttc gtg acc gac cgc gag cgt cag gag
 Y F V T D P E R Q E A V L E D P Y I L L
 723/241
 gtc agc tcc aag gtg tcc act gtc aag gat
 V S S K V S T V K D L L P L L E K V I G
 783/261
 gcc ggt aag ccg ctg ctg atc atc gcc gag
 A G K P L L I I A E D V E G E A L S T L
 843/281
 gtc gtc aac aag atc cgc gcc acc ttc aag
 V V N K I R G T F K S V A V K A P G F G
 903/301
 gac cgc cgc aag gcg atg ctg cag gat atg
 D R R K A M L Q D M A I L T G G Q V I S
 963/321
 gaa gag gtc gcc ctg acg ctg gag aac gcc
 E E V G L T L E N A D L S L L G K A R K
 1023/341
 gtc gtg gtc acc aag gac gag acc acc atc
 V V V T K D E T T I V E G A G D T D A I
 1083/361
 gcc gaa cga gtg gcc cag atc cgc cag gag
 A G R V A Q I R Q E I E N S D S D Y D R
 1143/381
 gag aag ctg cag gag cgc ctg gcc aag ctg
 E K L Q E R L A K L A G G V A V I K A G
 1203/401
 gcc gcc acc gag gtc gaa ctg aag gag cgc
 A A T E V E L K E R K H R I E D A V R N
 33/11
 agc agc ggc ctg gtg ccg cgc gcc agc cat
 S S G L V P R G S H
 93/31
 gcc cgt cgc gcc ctg gag cgc gcc ttg aac
 A R R G L E R G L N
 153/51
 gcc ccc aag gcc cgc aac gtc gtc ctg gaa
 G P K G R N V V L E
 213/71
 gat ggt gtg tcc atc gcc aag gag atc gag
 D G V S I A K E I E
 273/91
 gag ctg gtc aaa gag gta gcc aag aag acc
 E L V K E V A K K T
 333/111
 gcc acc gtc ctg gcc cag gcg ttg gtt cgc
 A T V L A Q A L V R
 393/131
 aac ccg ctg ggt ctg aaa cgc gcc atc gaa
 N P L G L K R G I E
 453/151
 ctg aag gcc gcc aag gag gtc gag acc aag
 L K G A K E V E T K
 513/171
 gcg ggt gac cag tcc atc ggt gac ctg atc
 A G D Q S I G D L I
 573/191
 gcc gtc atc acc gtc gag gag tcc aac acc
 G V I T V E E S N T
 633/211
 atg ccg ttc gac aag gcc tac atc tgg ggg
 M R P D K G Y I S G
 693/231
 cgc gtc ctg gag gac ccc tac atc ctg ctg
 A V L E D P Y I L L
 753/251
 ctg ctg ccg ctg ctg gag aag gtc atc gga
 L L P L L E K V I G
 813/271
 gac gtc gag gcc gag cgc ctg tcc acc ctg
 D V E G E A L S T L
 873/291
 tgg gtg cgc gtc aag gct ccc gcc ttc gcc
 S V A V K A P G F G
 933/311
 gcc att ctg acc ggt ggt cag gtg atc acc
 A I L T G G Q V I S
 993/331
 gac ctg tgg ctg cta gcc aag gcc cgc aag
 D L S L L G K A R K
 1053/351
 gtc gag gcc gcc ggt gac acc gac gcc atc
 V E G A G D T D A I
 1113/371
 atc gag aac agc gac tcc gac tac gac cgt
 I E N S D S D Y D R
 1173/391
 gcc ggt ggt gtc cgc gtg atc aag gcc ggt
 A G G V A V I K A G
 1233/411
 aag cac cgc atc gag gat gcg gtt cgc aat
 K H R I E D A V R N

FIG. 7A

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1263/421
GCC AAG GCC GCC GTC GAG GAG GGC ATC CTC
A K A A V E E G I V
1323/441
GCC CCG ACC CTG GAC GAG CTG AAG CTC GAA
A P T L D E L K L E
1383/461
AAG GTG GCG CTG GAG GCC CCG CTG AAG CAG
K V A L E A P L K Q
1443/481
GTG GTG GCC GAG AAG GTG CCG AAC CTG CCG
V V A E K V R N L P
1503/501
GTC TAC GAG GAT CTG CTC GCT GCC GGC GTT
V Y E D L L A A G V
1563/521
CTG CAG AAT GCG GCG TCC ATC GCG GGC CTG
L Q N A A S I A G L
1623/541
AAG CCG GAA AAG GAG AAG GCT TCC GTT CCC
K P E K E K A S V P
1683/561
gct agc ATG ggc tcc atc ggc gca gca agc
A S M G S I G A A S
1743/581
ctc aaa gtc cac cat gcc aat gag aac atc
L K V H H A N E N I
1803/601
cta gcc atg gta tac ctg ggt gca aaa gac
L A M V Y L G A K D
1863/621
cgc ttt gat aaa ctt cca gga ttc gga gac
R F D K L P G F G D
1923/641
aac gtt cac tct tca ctt aga gac atc ctc
N V H S S L R D I L
1983/661
tcg ttc agc ctt gcc agt aga ctt tat gct
S F S L A S R L Y A
2043/681
ttg cag tgt gtg aag gaa ctg tat aga gga
L Q C V K E L Y R G
2103/701
gca gat caa gcc aga gag ctc atc aat tcc
A D Q A R E L I N S
2163/721
aga aat gtc ctt cag cca agc tcc gtg gat
R N V L Q P S S V D
2223/741
att gtc ttc aaa gga ctg tgg gag aaa aca
I V F K G L W E K T
2283/761
ttc aga gtg act gag caa gaa agc aaa cct
F R V T E Q E S K P
2343/781
aga gtg gca tca atg gcc tct gag aaa atg
R V A S M A S E K M
2403/801
aca atg agc atg ttg gtg ctg ttg cct gat
T M S M L V L L P D
2463/821
ata atc aac ttt gaa aaa ctg act gaa tgg
I I N F E K L T E W
2523/841
atc aaa gtg tac tta cct cgc atg aag atg
I K V Y L P R M K M
2583/861
atg gct atg ggc att act gac gtg ttt agc
M A M G I T D V F S

1293/431
GCC GGT CCG GGT GTG ACG CTG TTG CAA GCG
A G G G V T L L Q A
1353/451
GGC GAC GAG GCG ACC GGC GCC AAC ATC GTG
G D E A T G A N I V
1413/471
ATC GCC TTC AAC TCC GGG CTG GAG CCG GGC
I A F N S G L E P G
1473/491
GCT GGC CAC GGA CTG AAC GCT CAG ACC GGT
A G H G L N A Q T G
1533/511
GCT GAC CCG GTC AAG GTG ACC CGT TCG GCG
A D P V K V T R S A
1593/531
TTC CTG ACC ACC GAG GCC GTC GTT GCC GAC
F L T T E A V V A D
1653/551
GGT GGC GGC GAC ATG GGT GGC ATG GAT TTC
G G G D M G G M D F
1713/571
atg gaa ttt tgt ttt gat gta ttc aag gag
M E F C F D V F K E
1773/591
ttc tac tgc ccc att gcc atc atg tca gcc
F Y C P I A I M S A
1833/611
agc acc agg aca cag ata aat aag gtt gtt
S T R T Q I N K V V
1893/631
agt att gaa gct cag tgt ggc aca tct gta
S I E A Q C G T S V
1953/651
aac caa atc acc aaa cca aat gat gtt tat
N Q I T K P N D V Y
2013/671
gaa gag aga tac cca atc ctg cca gaa tac
E E R Y P I L P E Y
2073/691
ggc ttg gaa cct atc aac ttt caa aca gct
G L E P I N F Q T A
2133/711
tgg gta gaa agt cag aca aat gga att atc
W V E S Q T N G I I
2193/731
tct caa act gca atg gtt ctg gtt aat gcc
S Q T A M V L V N A
2253/751
ttt aag gat gaa gac aca caa gca atg cct
F K D E D T Q A M P
2313/771
gtg cag atg atg tac cag att ggt tta ttt
V Q M M Y Q I G L F
2373/791
aag atc ctg gag ctt cca ttt gcc agt ggg
K I L E L P F A S G
2433/811
gaa gtc tca ggc ctt gag cag ctt gag agt
E V S G L E Q L E S
2493/831
acc agt tct aat gtt atg gaa gag agg aag
T S S N V M E E R K
2553/851
gag gaa aaa tac aac ctc aca tct gtc tta
E E K Y N L T S V L
2613/871
tct tca gcc aat ctg tct ggc atc tcc tca
S S A N L S G I S S

11/37

2643/881
gca gag agc ctg aag ata tct caa gct gtc cat gca gca cat gca gaa atc aat gaa gca
A E S L K I S Q A V H A A H A E I N E A

2703/901
ggc aga gag gtg gta ggg tca gca gag gct gga gtg gat gct gca agc gtc tct gaa gaa
G R E V V G S A E A G V D A A S V S E E

2763/921
ttt agg gct gac cat cca ttc ctc ttc tgt atc aag cac atc gca acc aac gcc gtt ctc
P R A D H P F L F C I K H I A T N A V L

2823/941
ttc ttt ggc aga tgt gtt gga tcc TAA
P F G R C V G S *

2673/891
cat gca gca cat gca gaa atc aat gaa gca
H A A H A E I N E A

2733/911
gga gtg gat gct gca agc gtc tct gaa gaa
G V D A A S V S E E

2793/931
aag cac atc gca acc aac gcc gtt ctc
K H I A T N A V L

FIG. 7C

1/1
atg tcc cct ata cta ggt tat tgg aaa att aag ggc ctt gtg caa ccc act cga ctt ctt
M S P I L G Y W K I K G L V Q P T R L L

61/21
ttg gaa tat ctt gaa gaa aaa tat gaa gag cat ctg tat gag cgc gat gaa ggt gat aaa
L E Y L E E K Y E E H L Y E R D E G D K

121/41
tgg cga aac aaa aag ttt gaa ttg ggt ttg gag ttt ccc aat ctt cct tat tat att gat
W R N K K F E L G L Z F P N L P Y Y I D

181/61
ggc gat gtt aaa tta aca cag tct atg gcc atc ata cgt tat ata gct gac aag cac aac
G D V K L T Q S M A I I R Y I A D K H N

241/81
atg ttg ggt ggt tgt cca aaa gag cgt gca gag att tca atg ctt gaa gga gcg gtt ttg
M L G G C P K E R A E I S M L E G A V L

301/101
gat att aga tac ggt gtt tgg aga att gca tat agt aaa gac ttt gaa act ctc aaa gtt
D I R Y G V S R I A Y S K D F E T L K V

361/121
gat ttt ctt agc aag cta cct gaa atg ctg aaa atg ttc gaa gat cgt tta tgt cat aaa
D F L S K L P E M L K M F E D R L C H K

421/141
aca tat tta aat ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat
T Y L N G D H V T H P D F M L Y D A L D

481/161
gtc gtc tta tac atg gac cca atg tgc ctg gat gcg ttc cca aaa tta gtt tgt ttt aaa
V V L Y M D P M C L D A F P K L V C F K

541/181
aaa cgt att gaa gct atc cca caa att gat aag tac ttg aaa tcc agc aag tat ata gca
K R I E A I P Q I D K Y L K S S K Y I A

601/201
tgg cct ttg cag ggc tgg caa gcc acg ttt ggt ggt ggc gac cat cct cca aaa tgg gat
W P L Q G W Q A T F G G G D H P P K S D

661/221
ctg gtt ccg cgt gga tcc cca gga att ccc ggg tgg act cga gca cca cca cca cca cca
L V P R G S F G I P G S T R A P P P P P

721/241
ctg aga tcc gcc tgc TAA
L R S G C *

31/11
aag ggc ctt gtg caa ccc act cga ctt ctt
K G L V Q P T R L L

91/31
cat ctg tat gag cgc gat gaa ggt gat aaa
H L Y E R D E G D K

151/51
gag ttt ccc aat ctt cct tat tat att gat
Z F P N L P Y Y I D

211/71
cgt tat ata gct gac aag cac aac
I I R Y I A D K H N

271/91
gag att tca atg ctt gaa gga gcg gtt ttg
E I S M L E G A V L

331/111
tat agt aaa gac ttt gaa act ctc aaa gtt
Y S K D F E T L K V

391/131
aaa atg ttc gaa gat cgt tta tgt cat aaa
K M F E D R L C H K

451/151
cct gac ttc atg ttg tat gac gct ctt gat
P D F M L Y D A L D

511/171
gtc gtc tta tac atg gac cca atg tgc ctg gat gcg ttc cca aaa tta gtt tgt ttt aaa
V V L Y M D P M C L D A F P K L V C F K

571/191
aag tac ttg aaa tcc agc aag tat ata gca
K Y L K S S K Y I A

631/211
ggc gac cat cct cca aaa tgg gat
G G G D H P P K S D

691/231
act cga gca cca cca cca cca cca
A C T C G A G C A C C A C C A C C A C C A

FIG. 8

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1/1
atg tcc cct ata cta ggc tat tgg aaa att aag ggc ctt gtg caa ccc act cga ctt ctt
M S P I L G Y W K I K G L V Q P T R L L
61/21
ttg gaa tat ctc gaa gaa aaa tat gaa gag cat ttg tat gag cgc gat gaa ggt gat aaa
L E Y L E E K Y E E H L Y E R D E G D K
121/41
tgg cga aac aaa aag ttt gaa ttg ggt ttg gag ttt ccc aat ctt cct tat tat att gat
W R N K K F E L G L E F P N L P Y Y I D
181/61
ggt gat gtt aaa tta aca cag tct atg gcc atc ata cgt tat ata gct gac aag cac aac
G D V K L T Q S M A I I R Y I A D K H N
241/81
atg ttg ggt ggt tgt cca aaa gag cgt gca gag att tca atg ctt gaa gga gcg gtt ttg
M L G G C P K E R A E I S M L E G A V L
301/101
gat att aga tac ggt gtt tgg aga att gca tat agt aaa gac ttt gaa act ctc aaa gtt
D I R Y G V S R I A Y S K D F E T L K V
361/121
gat ttt ctt agc aag cta cct gaa atg ctg aaa atg ttc gaa gat cgt tta tgt cat aaa
D F L S K L P E M L K M F E D R L C H K
421/141
aca tat tta aat ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat
T Y L N G D H V T H P D F M L Y D A L D
481/161
gtt gtt tta tac atg gac cca atg tgc ctg gat gcg ttc cca aaa tta gtt tgt ttt aaa
V V L Y M D P M C L D A P P K L V C F K
541/181
aaa cgt att gaa gct atc cca caa att gat aag tac ttg aaa tcc agc aag tat ata gca
K R I E A I P Q I D K Y L K S S K Y I A
601/201
tgg cct ttg cag ggc tgg caa gcc acg ttt ggt ggt ggc gac cat cct cca aaa tgg gat
W P L Q G W Q A T F G G G D H P P K S D
661/221
ctg gtt ccg cgt gga tcc ATG CAT GGA GAT ACA CCT ACA TTG CAT GAA TAT ATG TTA GAT
L V P R G S M H G D T P T L H E Y M L D
721/241
TTG CAA CCA GAG ACA ACT GAT CTC TAC TGT TAT GAG CAA TTA AAT GAC AGC TCA GAG GAG
L Q P E T T D L Y C Y E Q L N D S S E E
781/261
GAG GAT GAA ATA GAT GGT CCA GCT GGA CAA GCA GAA CCG GAC AGA GCC CAT TAC AAT ATT
E D E I D G P A G Q A E P D R A H Y N I
841/281
GTA ACC TTT TGT TGC AAG TGT GAC TCT ACG CTT CGG TTG TGC GTA CAA AGC ACA CAC GTA
V T P C C K C D S T L R L C V Q S T H V
901/301
GAC ATT CGT ACT TTG GAA GAC CTG TTA ATG GGC ACA CTA GGA ATT GTG TGC CCC ATC TGT
D I R T L E D L L M G T L G I V C P I C
961/321
TCT CAG AAA CCA TAA
S Q K P *

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FIG. 9

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3/1
 ATG GAT GGA GAT ACA CCT ACA TTG CAT GAA TAT ATG TTA GAT TTG CAA CCA GAG ACA ACT
 M D G D T P T L H E Y M L D L Q P E T T
 63/21
 GAT CTC TAC TGT TAT GAG CAA TTA AAT GAC AGC TCA GAG GAG GAG GAT GAA ATA GAT GGT
 D L Y C Y E Q L N D S S E E E D E I D G
 123/41
 CCA GCT GGA CAA GCA GAA CCG GAC AGA GCC CAT TAC AAT ATT GTA ACC TTT TGT TGC AAG
 P A G Q A E P D R A H Y N I V T F C C K
 183/61
 TGT GAC TCT ACG CTT CGG TTG TGC GTA CAA AGC ACA CAC GTA GAC ATT CGT ACT TTG GAA
 C D S T L R L C V Q S T H V D I R T L E
 243/81
 GAC CTG TTA ATG GGC ACA CTA GGA ATT GTG TGC CCC ATC TGT TCT CAG AAA CCA ACT AGT
 D L L M G T L G I V C P I C S Q K P T S
 303/101
 GGT GGC GGT GGC GGC GGA TCC CAC ATG GCC AAG ACA ATT GCG TAC GAC GAA GAG GCC CGT
 G G G G G G S H M A K T I A Y D E E A R
 363/121
 CGC GGC CTC GAG CGG GGC TTG AAC GCC CTC GCC GAT GCG GTA AAG GTG ACA TTG GGC CCC
 R G L E R G L N A L A D A V K V T L G P
 423/141
 AAG GGC CGC AAC GTC GTC CTG GAA AAG AAG TGG GGT GCC CCC ACG ATC ACC AAC GAT GGT
 K G R N V V L E K K W G A P T I T N D G
 483/161
 GTG TCC ATC GCC AAG GAG ATC GAG CTG GAG GAT CCG TAC GAG AAG ATC GGC GCC GAG CTG
 V S I A K E I E L E D P Y E K I G A E L
 543/181
 GTC AAA GAG GTA GCC AAG AAG ACC GAT GAC GTC GCC GGT GAC GGC ACC ACG ACG GCC ACC
 V K E V A K K T D D V A G D G T T T A T
 603/201
 GTG CTG GCC CAG GCG TTG GTT CGC GAG GGC CTG CGC AAC GTC GCG GCC GGC GCC AAC CCG
 V L A Q A L V R E G L R N V A A G A N P
 663/221
 CTC GGT CTC AAA CGC GGC ATC GAA AAG GCC GTG GAG AAG GTC ACC GAG ACC CTG CTC AAG
 L G L K R G I E K A V E K V T E T L L K
 723/241
 GGC GCC AAG GAG GTC GAG ACC AAG GAG CAG ATT GCG GCC ACC GCA GCG ATT TCG GCG GGT
 G A K E V E T K E Q I A A T A A I S A G
 783/261
 GAC CAG TCC ATC GGT GAC CTG ATC GCC GAG GCG ATG GAC AAG GTG GGC AAC GAG GGC GTC
 D Q S I G D L I A E A M D K V G N E G V
 843/281
 ATC ACC GTC GAG GAG TCC AAC ACC TTT GGG CTG CAG CTC GAG CTC ACC GAG GGT ATG CCG
 I T V E E S N T F G L Q L E L T E G M R
 903/301
 TTC GAC AAG GGC TAC ATC TCG GGG TAC TTC GTG ACC GAC CCG GAG CGT CAG GAG GCG GTC
 F D K G Y I S G Y F V T D P E R Q E A V
 963/321
 CTG GAG GAC CCC TAC ATC CTG CTG GTC AGC TCC AAG GTG TCC ACT GTC AAG GAT CTG CTG
 L E D P Y I L L V S S K V S T V K D L L
 1023/341
 CCG CTG CTC GAG AAG GTC ATC GGA GCC GGT AAG CCG CTG CTG ATC ATC GCC GAG GAC GTC
 P L L E K V I G A G K P L L I I A E D V
 1083/361
 GAG GGC GAG GCG CTG TCC ACC CTG GTC GTC AAC AAG ATC CCG GGC ACC TTC AAG TCG GTG
 E G E A L S T L V V N K I R G T F K S V
 1143/381
 GCG GTC AAG GCT CCC GGC TTC GGC GAC CCG CCG AAG GCG ATG CTG CAG GAT ATG GCC ATT
 A V K A P G F G D R R K A M L Q D M A I
 1203/401
 CTC ACC GGT GGT CAG GTG ATC AGC GAA GAG GTC GGC CTG ACG CTG GAG AAC GCC GAC CTG
 L T G G Q V I S E E V G L T L E N A D L

FIG. 10A

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1263/421	1293/431
TCG CTG CTA GGC AAG GCC CGC AAG GTC GTG	GTC ACC AAG GAC GAG ACC ACC ATC GTC GAG
S L L G K A R K V V	V T K D E T T I V E
1323/441	1353/451
GGC GCC GGT GAC ACC GAC GCC ATC GCC GGA	CGA GTG GCC CAG ATC CGC CAG GAG ATC GAG
G A G D T D A I A G	R V A Q I R Q E I E
1383/461	1413/471
AAC AGC GAC TCC GAC TAC GAC CGT GAG AAG	CTG CAG GAG CCG CTG GCC AAG CTG GCC GGT
N S D S D Y D R E K	L Q E R L A K L A G
1443/481	1473/491
GGT GTC GCG GTG ATC AAG GCC GGT GCC GCC	ACC GAG GTC GAA CTC AAG GAG CGC AAG CAC
G V A V I K A G A A	T E V E L K E R K H
1503/501	1533/511
CGC ATC GAG GAT GCG GTT CGC AAT GCC AAG	GCC GCC GTC GAG GAG GGC ATC GTC GCC GGT
R I E D A V R N A K	A A V E E G I V A G
1563/521	1593/531
GGG GGT GTG ACG CTG TTG CAA GCG GCC CCG	ACC CTG GAC GAG CTG AAG CTC GAA GGC GAC
G G V T L L Q A A P	T L D E L K L E G D
1623/541	1653/551
GAG GCG ACC GGC GCC AAC ATC GTG AAG GTG	GCG CTG GAG GCC CCG CTG AAG CAG ATC GCC
E A T G A N I V K V	A L E A P L K Q I A
1683/561	1713/571
TTC AAC TCC GGG CTG GAG CCG GGC GTG GTG	GCC GAG AAG GTG CCG AAC CTG CCG GCT GGC
F N S G L E P G V V	A E K V R N L P A G
1743/581	1773/591
CAC GGA CTG AAC GGT CAG ACC GGT GTC TAC	GAG GAT CTG CTC GCT GCC GGC GTT GCT GAC
H G L N A Q T G V Y	E D L L A A G V A D
1803/601	1833/611
CCG GTC AAG GTG ACC CGT TCG GCG CTG CAG	AAT GCG GCG TCC ATC GCG GGG CTG TTC CTG
P V K V T R S A L Q	N A A S I A G L F L
1863/621	1893/631
ACC ACC GAG GCC GTC GTT GCC GAC AAG CCG	GAA AAG GAG AAG GCT TCC GTT CCC GGT GGC
T T E A V V A D K P	E K E K A S V P G G
1923/641	
GGC GAC ATG GGT GGC ATG GAT TTC TGA	
G D M G G M D F *	

FIG. 10B

15/37

3/1
 ATG GCC AAG ACA ATT GCG TAC GAC GAA GAG
 M A K T I A Y D E E
 63/21
 GCC CTC GCC GAT GCG GTA AAG GTG ACA TTG
 A L A D A V K V T L
 123/41
 AAG AAG TGG GGT GCC CCC ACG ATC ACC AAC
 K K W G A P T I T N
 183/61
 CTG GAG GAT CCG TAC GAG AAG ATC GGC GCC
 L E D P Y E K I G A
 243/81
 GAT GAC GTC GCC GGT GAC GGC ACC ACG ACG
 D D V A G D G T T T
 303/101
 GAG GGC CTG CGC AAC GTC GCG GCC GGC GCC
 E G L R N V A A G A
 363/121
 AAG GCC GTG GAG AAG GTC ACC GAG ACC CTG
 K A V E K V T E T L
 423/141
 GAG CAG ATT GCG GCC ACC GCA GCG ATT TCG
 E Q I A A T A A I S
 483/161
 GCC GAG GCG ATG GAC AAG GTG GGC AAC GAG
 A E A M D K V G N E
 543/181
 TTT GGG CTG CAG CTC GAG CTC ACC GAG GGT
 F G L Q L E L T E G
 603/201
 GAT ACA CCT ACA TTG CAT GAA TAT ATG TTA
 D T P T L H E Y M L
 663/221
 TGT TAT GAG CAA TTA AAT GAC AGC TCA GAG
 C Y E Q L N D S S E
 723/241
 CAA GCA GAA CCG GAC AGA GCC CAT TAC AAT
 Q A E P D R A H Y N
 783/261
 ACG CTT CCG TTG TGC GTA CAA AGC ACA CAC
 T L R L C V Q S T H
 843/281
 ATG GGC ACA CTA GGA ATT GTG TGC CCC ATC
 M G T L G I V C P I C

33/11
 GCC CGT CGC GGC CTC GAG CCG GGC TTG AAC
 A R R G L E R G L N
 93/31
 GGC CCC AAG GGC CGC AAC GTC GTC CTG GAA
 G P K G R N V V L E
 153/51
 GAT GGT GTG TCC ATC GCC AAG GAG ATC GAG
 D G V S I A K E I E
 213/71
 GAG CTG GTC AAA GAG GTA GCC AAG AAG ACC
 E L V K E V A K K T
 273/91
 GCC ACC GTG CTG GCC CAG GCG TTG GTT CCG
 A T V L A Q A L V R
 333/111
 AAC CCG CTC GGT CTC AAA CCG GGC ATC GAA
 N P L G L K R G I E
 393/131
 CTC AAG GGC GCC AAG GAG GTC GAG ACC AAG
 L K G A K E V E T K
 453/151
 GCG GGT GAC CAG TCC ATC GGT GAC CTG ATC
 A G D Q S I G D L I
 513/171
 GGC GTC ATC ACC GTC GAG GAG TCC AAC ACC
 G V I T V E E S N T
 573/191
 ATG CCG TTC GAC AAG GGC CAT ATG CAT GGA
 M R F D K G H M H G
 633/211
 GAT TTG CAA CCA GAG ACA ACT GAT CTC TAC
 D L Q P E T T D L Y
 693/231
 GAG GAG GAT GAA ATA GAT GGT CCA GCT GGA
 E E D E I D G P A G
 753/251
 ATT GTA ACC TTT TGT TGC AAG TGT GAC TCT
 I V T F C C K C D S
 813/271
 GTA GAC ATT CGT ACT TTG GAA GAC CTG TTA
 V D I R T L E D L L
 873/291
 TGT TCT CAG AAA CCA TAA
 C S Q K P *

FIG. 11

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108/1
 ATG GCG AAG GTG AAC ATC AAG CCA CTC GAG GAC AAG ATT CTC GTG CAG GCC AAC GAG GCC
 M A K V N I K P L E D K I L V Q A N E A
 138/11
 168/21
 GAG ACC ACG ACC GCG TCC GGT CTG GTC ATT CCT GAC ACC GCC AAG GAG AAG CCG CAG GAG
 E T T T A S G L V I P D T A K E K P Q E
 198/31
 228/41
 GGC ACC GTC GTT GCC GTC GGC CCT GGC CCG TGG GAC GAG GAC GGC GAG AAG CCG ATC CCG
 G T V V A V G P G R W D E D G E K R I P
 258/51
 298/61
 CTG GAC GTT GCG GAG GGT GAC ACC GTC ATC TAC ACC AAG TAC GGC GGC ACC GAG ATC AAG
 L D V A E G D T V I Y S K Y G G T E I K
 318/71
 348/81
 TAC AAC GGC GAG GAA TAC CTG ATC CTG TCG GCA CGC GAC GTG CTG GCC GTC GTT TCC AAG
 Y N G E E Y L I L S A R D V L A V V S K
 378/91
 408/101
 ATG CAT GGA GAT ACA CCT ACA TTG CAT GAA TAT ATG TTA GAT TTG CAA CCA GAG ACA ACT
 M H G D T P T L H E Y M L D L Q P E T T
 438/111
 468/121
 GAT CTC TAC TGT TAT GAG CAA TTA AAT GAC AGC TCA GAG GAG GAG GAT GAA ATA GAT GGT
 D L Y C Y E Q L N D S S E E E D E I D G
 498/131
 528/141
 CCA GCT GGA CAA GCA GAA CCG GAC AGA GCC CAT TAC AAT ATT GTA AOC TTT TGT TGC AAG
 P A G Q A E P D R A H Y N I V T P C C K
 558/151
 588/161
 TGT GAC TCT ACG CTT CGG TTG TGC GTA CAA AGC ACA CAC GTA GAC ATT CGT ACT TTG GAA
 C D S T L R L C V Q S T H V D I R T L E
 618/171
 648/181
 GAC CTG TTA ATG GGC ACA CTA GGA ATT GTG TGC CCC ATC TGT TCT CAG AAA CCA TAG
 D L L M G T L G I V C P I C S Q K P *
 678/191

FIG. 12

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3/1	33/11
atg gat gga gat aca cct aca ttg cat gaa	tat atg tta gat ttg caa cca gag aca act
M D G D T P T L H E	Y M L D L Q P E T T
63/21	93/31
gat ctg tac tgt tat gag caa cta aat gac	agc tca gag gag gag gat gaa ata gat ggc
D L Y C Y E Q L N D	S S E E E D E I D G
123/41	153/51
cca gct gga caa gca gaa ccg gac aga gcc	cat tac aat att gta acc ttt tgt tgc aag
P A G Q A E P D R A	H Y N I V T P C C K
183/61	213/71
tgt gac tct acg ctt cgg ttg tgc gta caa	agc aca cac gta gac att cgt act ttg gaa
C D S T L R L C V Q	S T H V D I R T L E
243/81	273/91
gac ctg tta atg ggc aca cta gga att gtg	tgc ccc atc tgt tct cag aaa cca gcc atg
D L L M G T L G I V	C P I C S Q K P A M
303/101	333/111
gct cgt gcg gtc ggg atc gac ctg ggg acc	acc aac tcc gtc gtc tcc gtt ctg gaa ggt
A R A V G I D L G T	T N S V V S V L E G
363/121	393/131
ggc gac ccg gtc gtc gtc gcc aac tcc gag	ggc tcc agg acc acc ccg tca att gtc gcg
G D P V V V A N S E	G S R T T P S I V A
423/141	453/151
ttc gcc cgc aac ggt gag gtc ctg gtc gcc	cag ccc gcc aag aac cag gcg gtg acc aac
F A R N G E V L V G	Q P A K N Q A V T N
483/161	513/171
gtc gat cgc acc gtg cgc tcc gtc aag cga	cac atg gcc agc gac tcc tca gag att
V D R T V R S V K R	H M G S D W S I E I
543/181	573/191
gac gcc aag aaa tac acc gcg ccg gag atc	agg gcc ccc att ctg atg aag ctg aag ccg
D G K K Y T A P E I	S A R I L M K L K R
603/201	633/211
gac gcc gag gcc tac ctg ggt gag gac att	acc gac gcg gtt atc acc acc gcc gcc tac
D A E A Y L G E D I	T D A V I T T P A Y
663/221	693/231
ttc aat gac gcc cag cgt cag gcc acc aag	gac gcc gcc cag atc gcc gcc ctg aac gtg
F N D A Q R Q A T K	D A G Q I A G L N V
723/241	753/251
ctg ccg atc gtc aac gag ccg acc gcg gcc	gcg ctg gcc tac gcc ctg gac aag gcc gag
L R I V N E P T A A	A L A Y G L D K G E
783/261	813/271
aag gag cag cga atc ctg gtc ttc gac ttg	ggt ggt gcc act ttc gac gtt tcc ctg ctg
K E Q R I L V F D L	G G G T F D V S L L
843/281	873/291
gag atc gcc gag ggt gtg gtt gag gtc cgt	gcc act tcc ggt gac aac cac ctg gcc gcc
E I G E G V V E V R	A T S G D N H L G G
903/301	933/311
gac gac tgg gac cag ccg gtc gtc gat tgg	ctg gtg gac aag ttc aag gcc acc agc gcc
D D W D Q R V V D W	L V D K F K G T S G
963/321	993/331
atc gat ctg acc aag gac aag atg gcg atg	cag ccg ctg ccg gaa gcc gcc gag aag gca
I D L T K D K M A M	Q R L R E A A E K A
1023/341	1053/351
aag atc gag ctg agt tcc agt cag tcc acc	tcc atc aac ctg ccc tac atc acc gtc gac
K I E L S S S Q S T	S I N L P Y I T V D
1083/361	1113/371
gcc gac aag aac ccc ttg ttc tta gac gag	cag ctg acc ccg gcg gag ttc caa ccg atc
A D K N P L F L D E	Q L T R A E F Q R I
1143/381	1173/391
act cag gac ctg ctg gac ccg act ccg aag	ccg ttc cag tcc gtg atc gct gac acc gcc
T Q D L L D R T R K	P F Q S V I A D T G
1203/401	1233/411
att tcc gtg tcc gag atc gat cac gtt gtg	ctc ctg ggt ggt tcc acc ccg atg ccc ccg
I S V S E I D H V V L V G G S T R M P A	

FIG. 13A

SUBSTITUTE SHEET (RULE 26)

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1263/421	1293/431
GTG ACC GAT CTG GTC AAG GAA CTC ACC GGC	GGC AAG GAA CCC AAC AAG GGC GTC AAC CCC
V T D L V K E L T G	G K E P N K G V N P
1323/441	1353/451
GAT GAG GTT GTC GCG GTG GGA GCC GGT CTG	CAG GCC GGC GTC CTC AAG GGC GAG GTG AAA
D E V V A V G A A L	Q A G V L K G E V K
1383/461	1413/471
GAC GTT CTG CTG CTT GAT GTT ACC CCG CTG	AGC CTG GGT ATC GAG ACC AAG GGC GGG GTG
D V L L L D V T P L	S L G I E T K G G V
1443/481	1473/491
ATG ACC AGG CTC ATC GAG CGC AAC ACC ACG	ATC CCC ACC AAG CGG TCG GAG ACT TTC ACC
M T R L I E R N T T	I P T K R S E T F T
1503/501	1533/511
ACC GCC GAC GAC AAC CAA CCG TCG GTG CAG	ATC CAG GTC TAT CAG GGG GAG CGT GAG ATC
T A D D N Q P S V Q	I Q V Y Q G E R E I
1563/521	1593/531
GCC GCG CAC AAC AAG TTG CTC GGG TCC TTC	GAG CTG ACC GGC ATC CCG CCG GCG CCG CCG
A A H N K L L G S F	E L T G I P P A P R
1623/541	1653/551
GGG ATT CCG CAG ATC GAG GTC ACT TTC GAC	ATC GAC GCC AAC GGC ATT GTG CAC GTC ACC
G I P Q I E V T F D	I D A N G I V H V T
1683/561	1713/571
GCC AAG GAC AAG GGC ACC GGC AAG GAG AAC	ACG ATC CGA ATC CAG GAA GGC TCG GGC CTG
A K D K G T G K E N	T I R I Q E G S G L
1743/581	1773/591
TCC AAG GAA GAC ATT GAC CGC ATG ATC AAG	GAC GCC GAA GCG CAC GCC GAG GAG GAT CGC
S K E D I D R M I K	D A E A H A E E D R
1803/601	1833/611
AAG CGT CCG GAG GAG GCC GAT GTT CGT AAT	CAA GCC GAG ACA TTG GTC TAC CAG ACG GAG
K R R E E A D V R N	Q A E T L V Y Q T E
1863/621	1893/631
AAG TTC GTC AAA GAA CAG CGT GAG GCC GAG	GGT GGT TCG AAG GTA CCT GAA GAC ACG CTG
K F V K E Q R E A E	G G S K V P E D T L
1923/641	1953/651
AAC AAG GTT GAT GGC GCG GTG GCG GAA GCG	AAG GCG GCA CTT GGC GGA TCG GAT ATT TCG
N K V D A A V A E A	K A A L G G S D I S
1983/661	2013/671
GCC ATC AAG TCG CCG ATG GAG AAG CTG GGC	CAG GAG TCG CAG GCT CTG GGG CAA GCG ATC
A I K S A M E K L G	Q E S Q A L G Q A I
2043/681	2073/691
TAC GAA GCA GCT CAG GCT GCG TCA CAG GCC	ACT GGC GGT GCC CAC CCC GGC GGC GAG CCG
Y E A A Q A A S Q A	T G A A H P G G E P
2103/701	2133/711
GGC GGT GCC CAC CCC GGC TCG GCT GAG CTA	GCA TGA
G G A H P G S A E L A	*

FIG. 13B

19/37

3/1
 atg gat gga gat aca cct aca ttg cat gaa
 M D G D T P T L H E
 63/21
 gat ctc tac tgt tat gag caa tta aat gac
 D L Y C Y E Q L N D
 123/41
 cca gct gga caa gca gaa ccg gac aga gcc
 P A G Q A E P D R A
 183/61
 tgt gac tct acg ctc cgg ttg tgc gta caa
 C D S T L R L C V Q
 243/81
 gac ctg tta atg ggc aca cta gga att gtg
 D L L M C T L G I V
 303/101
 gct cgt gcg gtc ggg atc gac ctc ggg acc
 A R A V G I D L G T
 363/121
 ggc gac cgg gtc gtc gtc gcc aac tcc gag
 G D P V V V A N S E
 423/141
 TTC GCC CGC AAC GGT GAG GTG CTG GTC GGC
 F A R N G E V L V G
 483/161
 GTC GAT CGC ACC GTG CGC TCG GTC AAG CGA
 V D R T V R S V K R
 543/181
 GAC GGC AAG AAA TAC ACC GCG CCG GAG ATC
 D G K K Y T A P E I
 603/201
 GAC GCC GAG GCC TAC CTC GGT GAG GAC ATT
 D A E A Y L G E D I
 663/221
 TTC AAT GAC GCC CAG CGT CAG GCC ACC AAG
 F N D A Q R Q A T K
 723/241
 CTG CGG ATC GTC AAC GAG CCG ACC GCG GCC
 L R I V N E P T A A
 783/261
 AAG GAG CAG CGA ATC CTG GTC TTC GAC TTG
 K E Q R I L V F D L
 843/281
 GAG ATC GGC GAG GGT GTG GTT GAG GTC CGT
 E I G E G V V E V R
 903/301
 GAC GAC TGG GAC CAG CCG GTC GTC GAT TGG
 D D W D Q R V V D W
 963/321
 ATC GAT CTG ACC AAG GAC AAG ATG GCG ATG
 I D L T K D K M A M
 1023/341
 AAG ATC GAG CTG AGT TCG AGT CAG TCC ACC
 K I E L S S S Q S T
 1083/361
 GCC GAC AAG AAC CCG TTG TTC TTA GAC GAG
 A D K N P L F L D E
 1143/381
 ACT CAG GAC CTG CTG GAC CGC ACT CGC AAG
 T Q D L L D R T R K
 1203/401
 ATT TCG GTG TCG GAG ATC GAT CAC GTT GTG
 I S V S E I D H V V
 33/11
 tat atg tta gat ttg caa cca gag aca act
 Y M L D L Q P E T T
 93/31
 agc tca gag gag gag gat gaa ata gat ggt
 S S E E E D E T D G
 153/51
 cat tac aat att gta acc ttc tgt tgc aag
 H Y N I V T F C C K
 213/71
 agc aca cac gta gac att cgt acc ttg gaa
 S T H V D I R T L E
 273/91
 tgc ccc atc tgt tct cag aaa cca gcc atg
 C P I C S Q K P A M
 333/111
 ACC AAC TCC GTC GTC TCG GTT CTG GAA GGT
 T N S V V S V L E G
 393/131
 GGC TCC AGG ACC ACC CCG TCA ATT GTC GCG
 G S R T T P S I V A
 453/151
 CAG CCC GCC AAG AAC CAG GCG GTG ACC AAC
 Q P A K N Q A V T N
 513/171
 CAC ATG GGC AGC GAC TGG TCC ATA GAG ATT
 H M G S D W S I E I
 573/191
 AGC GCC CGC ATT CTG ATG AAG CTG AAG CGC
 S A R I L M K L K R
 633/211
 ACC GAC GCG GTT ATC ACG ACG CCC GCC TAC
 T D A V I T T P A Y
 693/231
 GAC GCC GGC CAG ATC GCC GGC CTC AAC GTG
 D A G Q I A G L N V
 753/251
 GCG ctg gcc TAC GGC CTC GAC AAG GCC GAG
 A L A Y G L D K G E
 813/271
 GGT GGT GGC ACT TTC GAC GTT TCC CTG CTG
 G G G T F D V S L L
 873/291
 GCC ACT TCG GGT GAC AAC CAC CTC GGC GGC
 A T S G D N H L G G
 933/311
 CTG GTG GAC AAG TTC AAG GGC ACC AGC GCC
 L V D K F K G T S G
 993/331
 CAG CGG CTG CCG GAA GCC GCC GAG AAG GCA
 Q R L R E A A E K A
 1053/351
 TCG ATC AAC CTG CCC TAC ATC ACC GTC GAC
 S I N L P Y I T V D
 1113/371
 CAG CTG ACC CGC GCG GAG TTC CAA CCG ATC
 Q L T R A E F Q R I
 1173/391
 CCG TTC CAG TCG GTG ATC GGT GAC ACC GGC
 P F Q S V I A D T G
 1233/411
 CTC GTG GGT GGT TCG ACC CCG ATG CCC GCG
 L Y G G S T R M P A

FIG. 14A

SUBSTITUTE SHEET (RULE 26)

20/37

1263/421
 GTG ACC GAT CTG GTC AAG GAA CTC ACC GGC
 V T D L V K E L T G
 1323/441
 GAT GAG GTT GTC GCG GTG GGA GCC GCT CTG
 D E V V A V G A A L
 1383/461
 GAC GTT CTG CTG CTT GAT GTT ACC CCG CTG
 D V L L L D V T P L
 1443/481
 ATG ACC AGG CTC ATC GAG CGC AAC ACC ACG
 M T R L I E R N T T
 1503/501
 ACC GCC GAC GAC AAC CAA CCG TCG GTG CAG
 T A D D N Q P S V Q
 1563/521
 GCC GCG CAC AAC AAG TTG CTC GGG TCC TTC
 A A H N K L L G S F
 1623/541
 GGG ATT CCG CAG ATC GAG GTC ACT TTC GAC
 G I P Q I E V T F D
 1683/561
 GCC AAG GAC AAG GGC ACC GGC AAG GAG AAC
 A K D K G T G K E N
 1743/581
 TCC AAG GAA GAC ATT GAC CGC ATG ATC AAG
 S K E D I D R M I K
 1803/601
 AAG CGT CCG GAG GAG GCC GAT GTT CGT AAT
 K R R E E A D V R N
 1863/621
 AAG TTC GTC AAA GAA CAG CGT GAG GCC GAG
 K F V K E Q R E A E
 1923/641
 AAC AAG GTT GAT GCC GCG GTG GCG GAA GCG
 N K V D A A V A E A
 1983/661
 GGC ATC AAG TCG GCG ATG GAG AAG CTG GGC
 A I K S A M E K L G
 2043/681
 TAC GAA GCA GCT CAG GCT GCG TCA CAG GCC
 Y E A A Q A A S Q A
 2103/701
 GGC GGT GCC CAC CCC GGC TCG GCT GAT GAC
 G G A H P G S A D D V V D A E V V D D G
 2163/721
 CGG GAG GCC AAG TGA
 R E A K

1293/431
 GGC AAG GAA CCC AAC AAG GGC GTC AAC CCC
 G K E P N K G V N P
 1353/451
 CAG GCC GGC CTC CTC AAG GGC GAG GTG AAA
 Q A G V L K G E V K
 1413/471
 AGC CTG GGT ATC GAG ACC AAG GGC GGG GTG
 S L G I E T K G G V
 1473/491
 ATC CCC ACC AAG CCG TCG GAG ACT TTC ACC
 I P T K R S E T F T
 1533/511
 ATC CAG GTC TAT CAG GGG GAG CGT GAG ATC
 I Q V Y Q G E R E I
 1593/531
 GAG CTG ACC GGC ATC CCG CCG GCG CCG CGG
 E L T G I P P A P R
 1653/551
 ATC GAC GCC AAC GGC ATT GTG CAC GTC ACC
 I D A N G I V H V T
 1713/571
 ACG ATC CGA ATC CAG GAA GGC TCG GGC CTG
 T I R I Q E G S G L
 1773/591
 GAC GCC GAA GCG CAC GCC GAG GAG GAT CGC
 D A E A H A E E D R
 1833/611
 CAA GCC GAG ACA TTG GTC TAC CAG ACG GAG
 Q A E T L V Y Q T E
 1893/631
 GGT GGT TCG AAG GTC CCT GAA GAC ACG CTG
 G G S K V P E D T L
 1953/651
 AAG GCG GCA CTT GGC GGA TCG GAT ATT TCG
 K A A L G G S D I S
 2013/671
 CAG GAG TCG CAG GCT CTG GGG CAA GCG ATC
 Q E S Q A L G Q A I
 2073/691
 ACT GGC GGT GCC CAC CCC GGC GGC GAG CCG
 T G A A H P G G E P
 2133/711
 GTT GTG GAC GCG GAG GTG GTC GAC GAC GGC
 G T G A E V V D D G

FIG. 14B

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3/1      33/11
8c9 gCA AAA GAA ATT AAA TTT TCA TCA GAT GGC CGT TCA GCT ATG GTC CGT GGT GTC GAT
M A K E I K F S S D A R S A M V R G V D
53/21
ATC CTT GCA GAT ACT GTT AAA GTA ACT TTG GCA CCA AAA GGT CGC AAT GTC GTT CTT GAA
I L A D T V K V T L G P K G R N V V L E
123/41
AAG TCA TTC GGT TCA CCC TTG ATT ACC AAT GAC GGT GTG ACT ATT GCC AAA GAA ATT GAA
K S F G S P L I T N D G V T I A K E I E
183/61
TTA GAA GAC CAT TTT GAA AAT ATG GGT GCC AAA TTG GTA TCA GAA GTA GCT TCA AAA ACC
L E D H F E N M G A K L V S E V A S K T
243/81
AAT GAT ATC GCA GGT GAT GGA ACT ACA ACT GCA ACT GTT TTG ACC CAA GCA ATC GTC CGT
N D I A G D G T T T A T V L T Q A I V R
303/101
GAA GCA ATC AAA AAC GTC ACA GCA GGT GCA AAT CCA ATC GGT ATT CGT CGT GGG ATT GAA
E G I K N V T A G A N P I G I R R G I E
363/121
ACA GCA GTT GCC GCA GCA GTT GAA GCT TTG AAA AAC AAC GTC ATC CCT GTT GCC AAT AAA
T A V A A A V E A L K N N V I P V A N K
423/141
GAA GCT ATC GCT CAA GTT GCA GCC GTA TCT TCT CGT TCT GAA AAA GTT GGT GAG TAC ATC
E A I A Q V A A V S S R S E K V G E Y I
483/161
TCT GAA GCA ATG GAA AAA GTT GGC AAA GAC GGT GTC ATC ACC ATC GAA GAG TCA CGT GGT
S E A M E K V G K D G V I T I E E S R G
543/181
ATG GAA ACA GAG CTT GAA GTC GTA GAA GGA ATG CAG TTT GAC CGT GGT TAC CTT TCA CAG
M E T E L E V V E G M Q F D R G Y L S Q
603/201
TAC ATG GTG ACA GAT AGC GAA AAA ATG GTG GCT GAC CTT GAA AAT CCG TAC ATT TTG ATT
Y M V T D S E K M V A D L E N P Y I L I
663/221
ACA GAC AAG AAA ATT TCC AAT ATC CAA GAA ATC TTG CCA CTT TTG GAA AGC ATT CTC CAA
T D K K I S N I Q E I L P L L E S I L Q
723/241
AGC AAT CGT CCA CTC TTG ATT ATT GCG GAT GAT GTG GAT GGT GAG GCT CTT CCA ACT CTT
S N R P L L I I A D D V D G E A L P T L
783/261
GTT TTG AAC AAG ATT CGT GGA ACC TTC AAC GTA GTA GCA GTC AAG GCA CCT GGT TTT GGT
V L N K I R G T F N V V A V K A P G F G
843/281
GAC CGT CGC AAA GCC ATG CTT GAA GAT ATC GCC ATC TTA ACA GGC GGA ACA GTT ATC ACA
D R R K A M L E D I A I L T G G T V I T
903/301
GAA GAC CTT GGT CTT GAG TTG AAA GAT GCG ACA ATT GAA GCT CTT GGT CAA GCA GCG AGA
E D L G L E L K D A T I E A L G Q A A R
963/321
GTG ACC GTG GAC AAA GAT AGC ACG GTT ATT GTA GAA GGT GCA GGA AAT CCT GAA GCG ATT
V T V D K D S T V I V E G A G N P E A I
1023/341
TCT CAC CGT GTT GCG GTT ATC AAG TCT CAA ATC GAA ACT ACA ACT TCT GAA TTT GAC CGT
S H R V A V I K S Q I E T T T S E F D R
1083/361
GAA AAA TTG CAA GAA CGC TTG GCC AAA TTG TCA GGT GGT GTA GCG GTT ATT AAG GTC GGA
E K L Q E R L A K L S G G V A V I K V G
1143/381
GCC GCA ACT GAA ACT GAG TTG AAA GAA ATG AAA CTC CGC ATT GAA GAT GCC CTC AAC GCT
A A T E T E L K E M K L R I E D A L N A
1203/401
ACT CGT GCA GCT GTT GAA GAA GGT ATT GTT GCA GGT GGT GGA ACA GCT CTT GCC AAT GTG
T R A A V E E G I V A G G G T A L A N V

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FIG. 15A

SUBSTITUTE SHEET (RULE 26)

22/37

1263/421
 ATT CCA GCT GTT GCT ACC TTG GAA TTG ACA
 I P A V A T L E L T
 1323/441
 CTC CGT GCT TTG GAA GAA COT GTT CGT CAA
 L R A L E E P V R Q
 1383/461
 ATC GTT ATC GAT CGT TTG AAA AAT GCT GAG
 I V I D R L K N A E
 1443/481
 GAG TGG GTT AAC ATG ATT GAT CAA GGT ATC
 E W V N M I D Q G I
 1503/501
 CTA CAA AAT GCA GCA TCT GTA GCC AGC TTG
 L Q N A A S V A S L
 1563/521
 AAA CCA GAA CCA GTA GCC CCA GCT CCA GCA
 K P E P V A P A P A
 1623/541
 GGA GCT AGC atg cat gga gat aca cct aca
 G A S M H G D T P T
 1683/561
 gag aca act gat ctc tac tgt tat gag caa
 E T T D L Y C Y E Q
 1743/581
 ata gat ggt cca gct gga caa gca gaa ccg
 I D G P A G Q A E P
 1803/601
 tgt tgc aag tgt gac tct acg ctc cgg ttg
 C C K C D S T L R L
 1863/621
 act ttg gaa gac ctg tta atg ggc aca cta
 T L E D L L M G T L
 1923/641
 cca TAA
 P .

1293/431
 GGA GAT GAA GCA ACA GGA CGT AAT ATT GTT
 G D E A T G R N I V
 1353/451
 ATT GCT CAC AAT GCA GGA TTT GAA GGA TCT
 I A H N A G F E G S
 1413/471
 CTT CGT ATA GGA TTC AAC GCA GCA ACT GGC
 L G I G P N A A T G
 1473/491
 ATT GAT CCA GTT AAA GTG AGT CGT TCA GCC
 I D P V K V S R S A
 1533/511
 ATT TTG ACA ACA GAA GCA GTC GTA GCC AAT
 I L T T E A V V A N
 1593/531
 ATG GAT CCA AGT ATG ATG GGT GGA ATG GGC
 H D P S M M G G M G
 1653/551
 ttg cat gaa tat atg tta gat ttg caa cca
 L H E Y M L D L Q P
 1713/571
 tta aat gac agc tca gag gag gag gat gaa
 L N D S S E E E D E
 1773/591
 gac aga gcc cat tac aat att gta acc ttt
 D R A H Y N I V T F
 1833/611
 tgc gta caa agc aca cac gta gac att cgt
 C V Q S T H V D I R
 1893/631
 gga att gtg tgc ccc atc tgt tct cag aaa
 G I V C P I C S Q K

FIG. 15B

23/37

4/1
 ATG AAA GAG CTC AAG TTC GGT GTC GAA GCC CGT GCT CAG CTC CTC AAG GGT GTT GAC ACT
 M X E L K F G V E A R A Q L L K G V D T
 64/21
 CTG GCC AAG GCC GTG ACT TCG ACT CTT GGT CCT AAG GGT CGT AAC GTC CTT ATC GAG TCT
 L A K A V T S T L G P K G R N V L I E S
 124/41
 CCC TAT GGC TCC CCT AAG ATC ACC AAG GAT GGT GTC TCT GTT GCC AAG GCC ATC ACT CTC
 P Y G S P K I T K D G V S V A K A I T L
 184/61
 CAA GAC AAG TTC GAG AAC CTC GGT GCT CGC CTC CTC CAG GAT GTC GCT TCT AAG ACC AAC
 Q D K F E N L G A R L L Q D V A S K T N
 244/81
 GAG ATT GCT GGT GAC GGT ACC ACC ACC GCT ACC GTC CTT GCC CGT GCC ATC TTC TCT GAG
 E I A G D G T T T A T V L A R A I F S E
 304/101
 ACC GTG AAG AAT GTT GCT GCT GGC TGC AAC CCC ATG GAT CTG CGC CGC GGT ATC CAG GCT
 T V K N V A A G C N P M D L R R G I Q A
 364/121
 GCT GTT GAT GCT GTC GTC GAC TAC CTC CAG AAG AAC AAG CGT GAC ATC ACC ACC GGT GAG
 A V D A V V D Y L Q K N K R D I T T G E
 424/141
 GAG ATC GCT CAG GTT GCT ACT ATC TCC GCT AAC GGT GAC ACC CAC ATT GGT AAG CTG ATC
 E I A Q V A T I S A N G D T H I G K L I
 484/161
 TCC ACC GCC ATG GAG CGT GTT GGC AAG GAG GGT GTC ATC ACT GTC AAG GAG GGC AAG ACC
 S T A M E R V G K E G V I T V K E G K T
 544/181
 ATT GAG GAT GAG CTC GAG GTC ACT GAG GGT ATG CGC TTC GAC CGT GGA TAC ACC TCC CCC
 I E D E L E V T E G M R F D R G Y T S P
 604/201
 TAC TTC ATC ACC GAT ACC AAG TCC CAG AAG GTT GAG TTC GAG AAG CCT CTG ATT CTG CTG
 Y F I T D T K S Q K V E F E K P L I L L
 664/221
 TCT GAG AAG AAG ATC TCT GCC GTT CAG GAC ATC ATC CCC GCC CTT GAG GCC TCC ACC ACC
 S E K K I S A V Q D I I P A L E A S T T
 724/241
 CTC CGC CGC CCC CTG GTT ATT ATC GCA GAG GAC ATT GAG GGT GAG GCT CTC GCC GTC TGC
 L R R P L V I I A E D I E G E A L A V C
 784/261
 ATT CTG AAC AAG CTT CGT GGC CAG CTG CAG GTC GCT GCT GTC AAG GCT CCT GGA TTC GGT
 I L N K L R G Q L Q V A A V K A P G F G
 844/281
 GAC AAC CGC AAG AGC ATC CTG GGC GAT CTT GCC GTC CTT ACC AAC GGT ACC GTC TTC ACT
 D N R K S I L G D L A V L T N G T V F T
 904/301
 GAT GAG CTC GAC ATC AAA CTC GAG AAG CTT ACC CCC GAT ATG CTT GGT TCC ACC GGC GCC
 D E L D I K L E K L T P D M L G S T G A
 964/321
 ATC ACC ATC ACC AAG GAG GAC ACC ATC ATC CTG AAC GGG GAG GGC AGC AAG GAC GCC ATT
 I T I T K E D T I I L N G E G S K D A I
 1024/341
 GGC CAG CGC TGC GAG CAG ATT CGC GGT GTC ATG GCG GAC CCC AGC ACC TCC GAA TAC GAG
 A Q R C E Q I R G V M A D F S T S E Y E
 1084/361
 AAG GAG AAG CTC CAG GAG CGT CTA GCT AAG CTC TCT GGC GGT GTT GCC GTC ATC AAG GTC
 K E K L Q E R L A K L S G G V A V I K V
 1144/381
 GGT GGT GCC TCC GAG GTT GAG GTC GGT GAG AAG AAG GAC CGT GTT GTC GAT GCT CTC AAT
 G G A S E V E V G E K K D R V V D A L N
 1204/401
 GCT ACC CGT GCT GCT GTT GAG GAG GGT ATC CTC CCC GGT GGT GGT ACC GCC CTT CTC AAG
 A T R A A V E E G I L P G G G T A L L K

FIG. 16A

SUBSTITUTE SHEET (RULE 26)

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1264/421	1294/431
GCC GCC GCC AAC GGC CTT GAC AAT GTC AAG	CCC GAG AAC TTC GAC CAG CAA CTC GGT GTG
A A A N G L D N V K	P E N F D Q Q L G V
1324/441	1354/451
AGC ATC ATC AAG AAT GCC ATC ACC CCC CCC	GCT CGC ACC ATT GTT GAG AAC GCC GGC CTC
S I I K N A I T R P	A R T I V E N A G L
1384/461	1414/471
GAG GGC AGC GTC ATT GTC GGC AAG CTG ACC	GAC GAG TTC GCC AAG GAC TTC AAC CGC GGT
E G S V I V G K L T	D E F A K D F N R G
1444/481	1474/491
TTC GAC AGC TCC AAG GGC GAG TAC GTC GAC	ATG ATC TCC AGC GGT ATC CTC GAT CCC CTC
F D S S K G E Y V D	M I S S G I L D P L
1504/501	1534/511
AAG GTT GTT CGC ACC GCT CTG CTC GAC GCC	AGC GGT GTC GCC TCC CTG CTC GGT ACC ACT
K V V R T A L L D A	S G V A S L L G T T
1564/521	1594/531
GAG GTC GCT ATT GTT GAG GCC CCT GAG GAG	AAG GGC CCC GCT GCT CCT GGC ATG GGT GGT
E V A I V E A P E E	K G P A A P G M G G
1624/541	1654/551
ATG GGT GGT ATG GGC GGC ATG GGC GGC ATG	CAT GGA GAT ACA CCT ACA TTG CAT GAA TAT
M G G M G G M G G M	H G D T P T L H E Y
1684/561	1714/571
ATG TTA GAT TTG CAA CCA GAG ACA ACT GAT	CTC TAC TGT TAT GAG CAA TTA AAT GAC AGC
M L D L Q P E T T D	L Y C Y E Q L N D S
1744/581	1774/591
TCA GAG GAG GAG GAT GAA ATA GAT GGT CCA	GCT GCA CAA GCA GAA CCG GAC AGA GCC CAT
S E E E D E I D G P	A G Q A E P D R A H
1804/601	1834/611
TAC AAT ATT GTA ACC TTT TGT TGC AAG TGT	GAC TCT ACG CTT CGG TTG TGC GTA CAA AGC
Y N I V T F C C K C	D S T L R L C V Q S
1864/621	1894/631
ACA CAC GTA GAC ATT CGT ACT TTG GAA GAC	CTG TTA ATG GGC ACA CTA GGA ATT GTG TGC
T H V D I R T L E D	L L M G T L G I V C
1924/641	
CCC ATC TGT TCT CAG AAA CCA TAG	
P I C S Q K P *	

FIG. 16B

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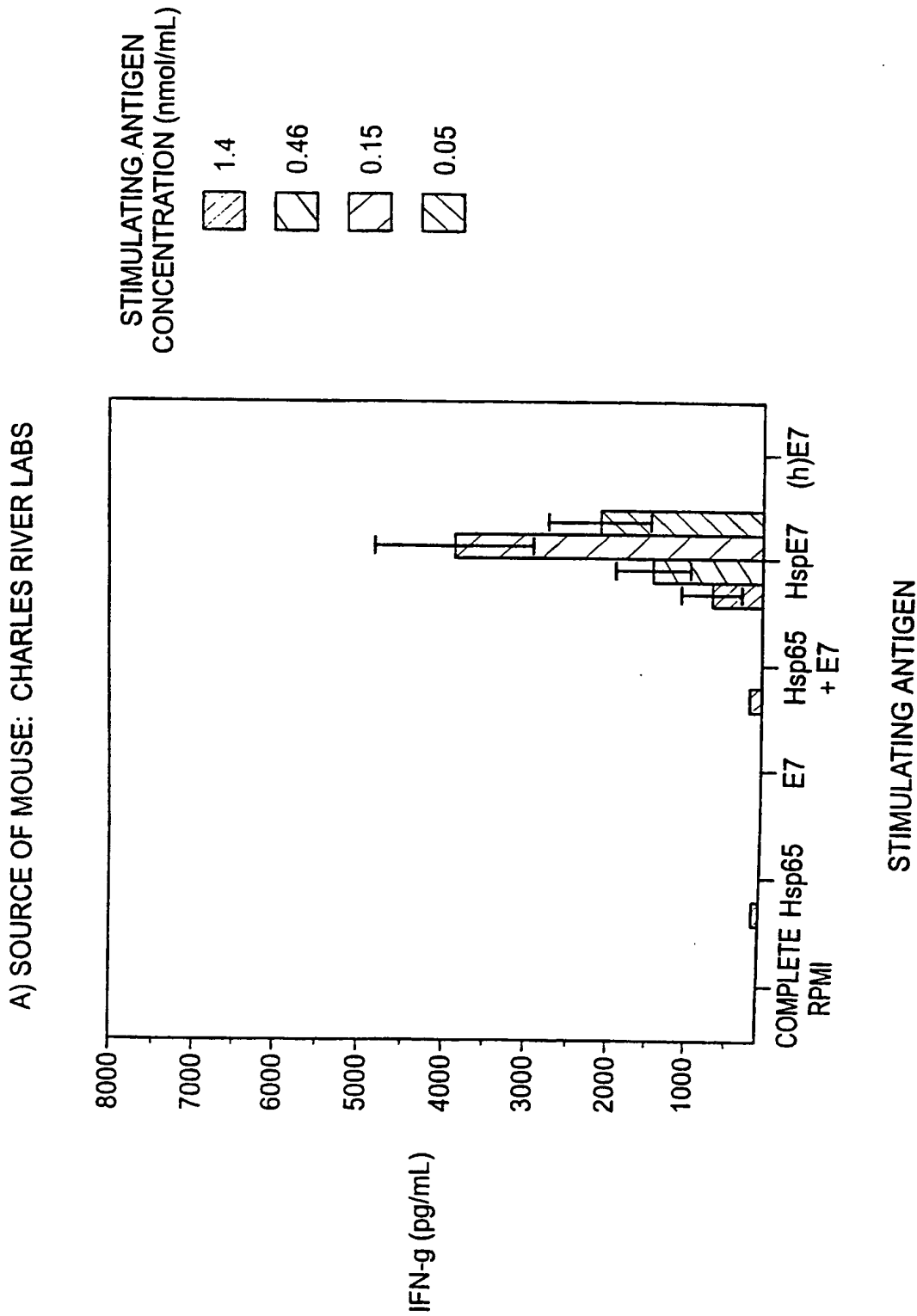


FIG. 17A

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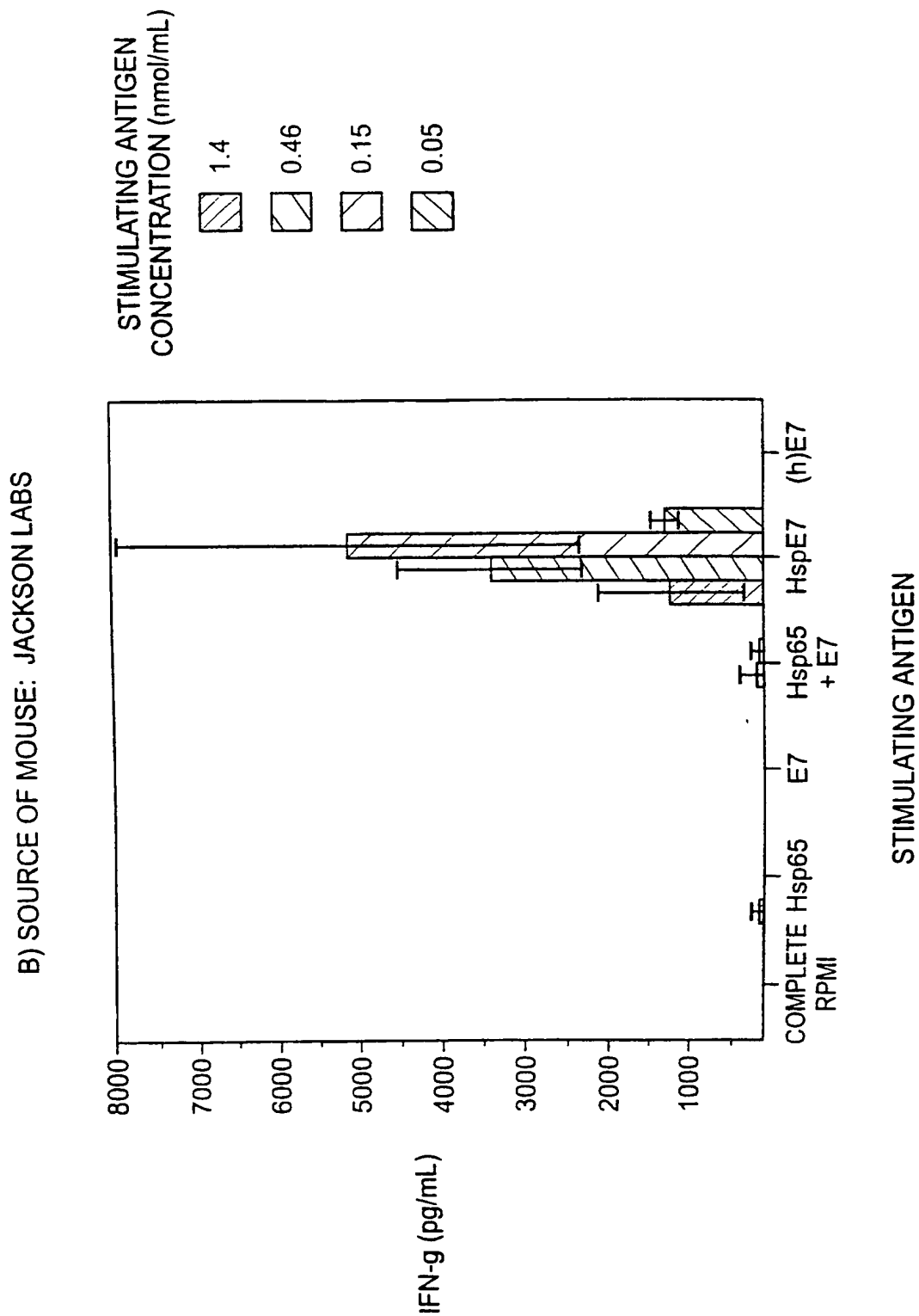


FIG. 17B

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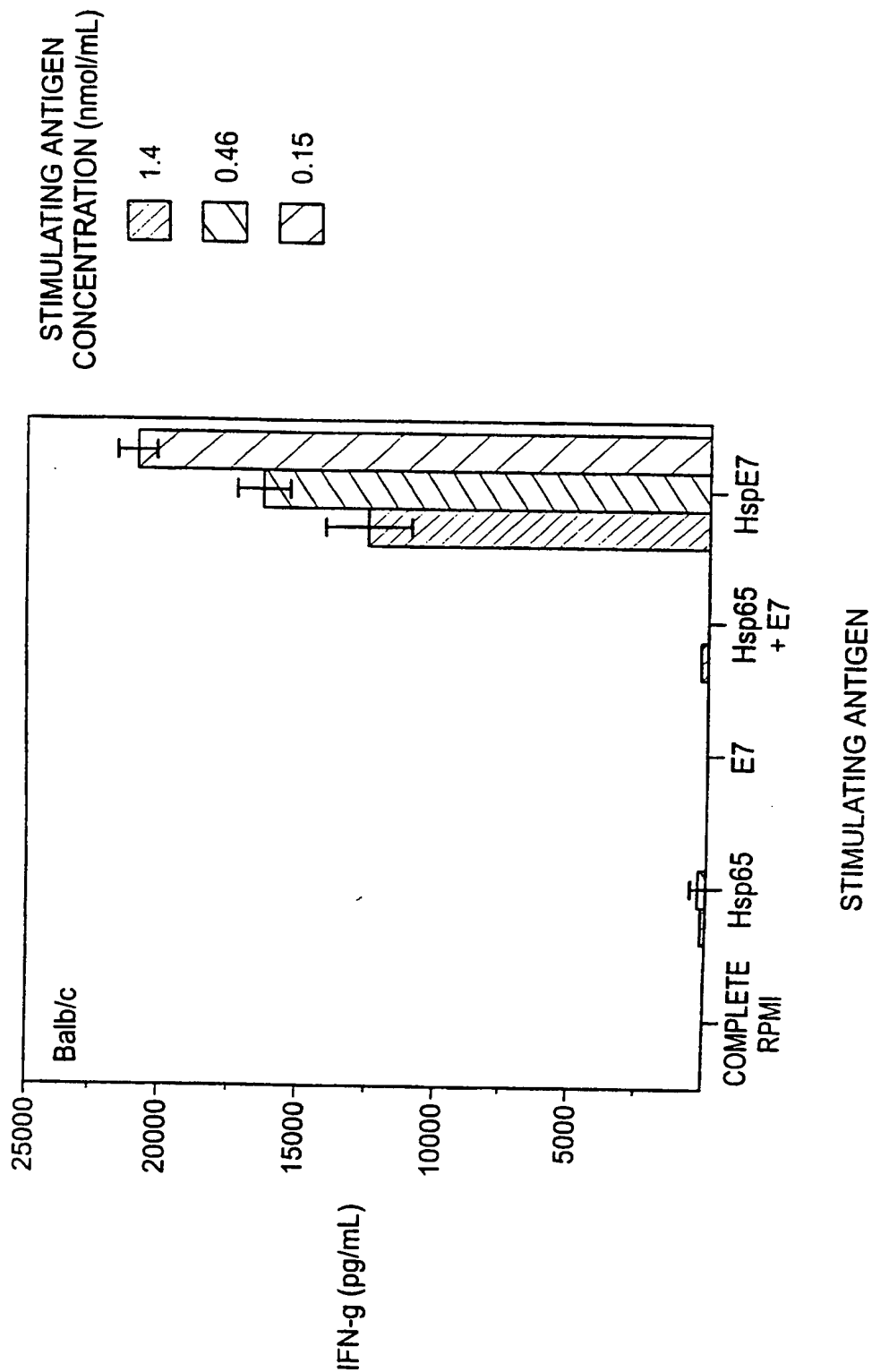


FIG. 18A

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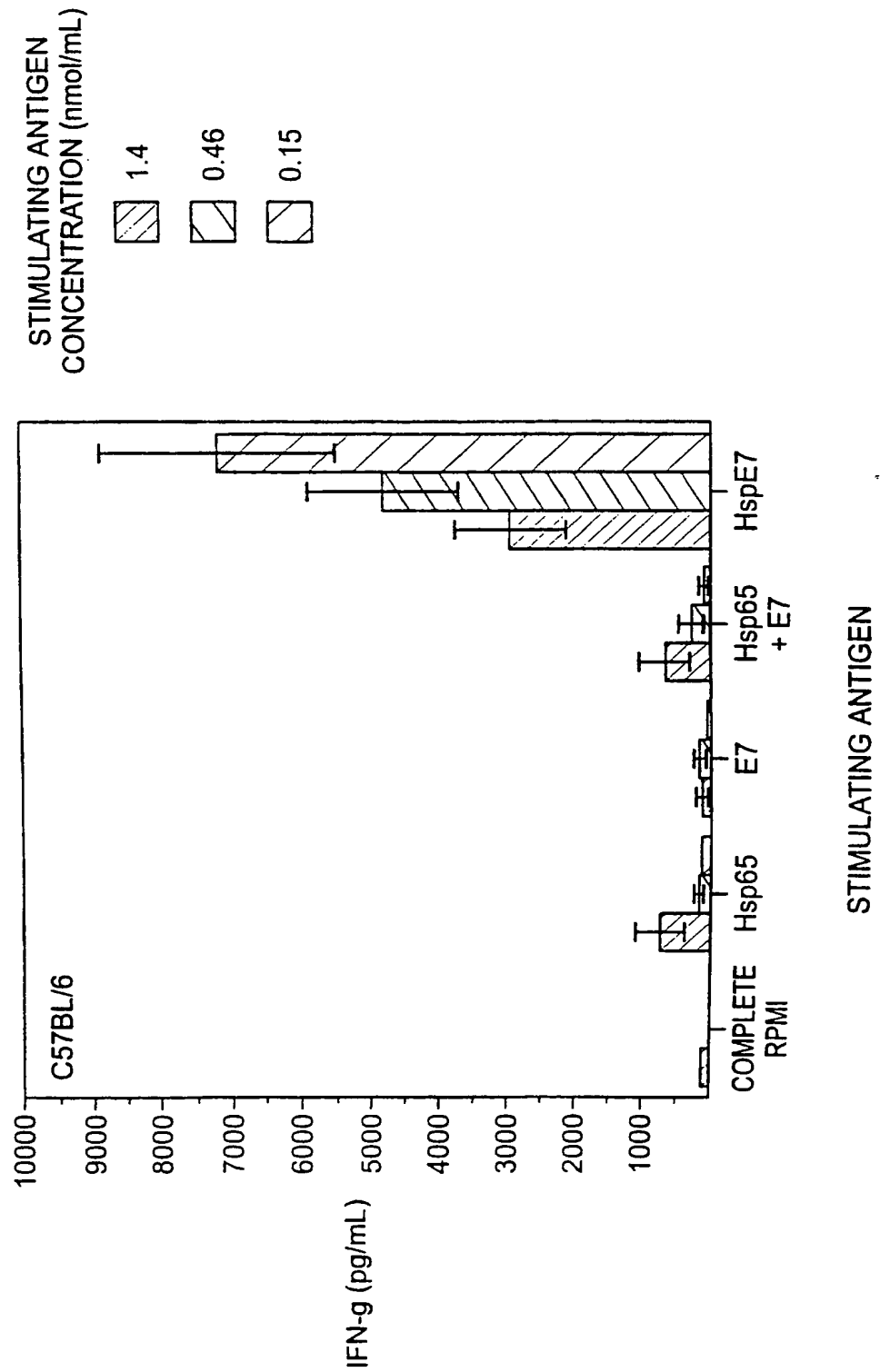


FIG. 18B

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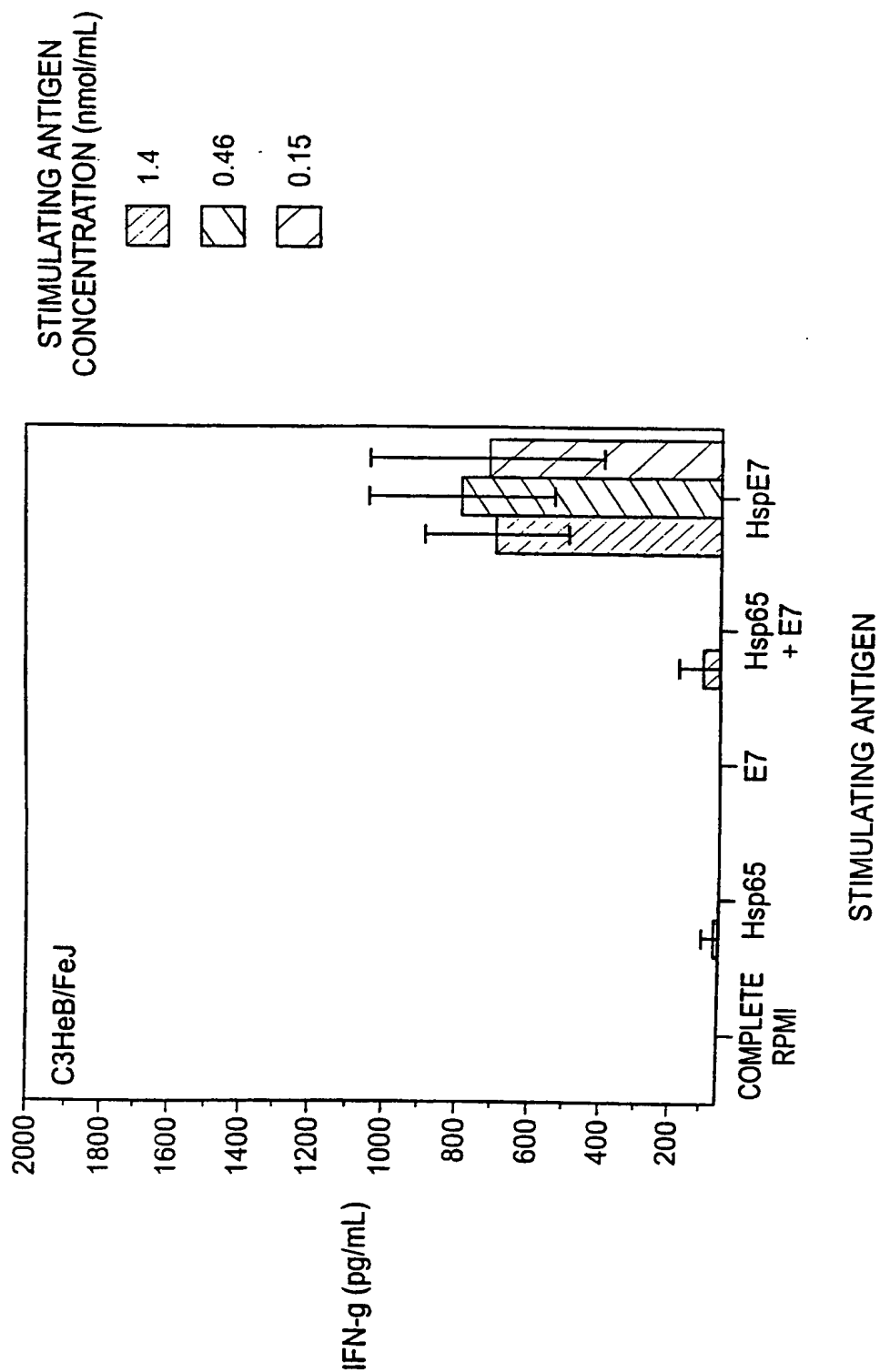


FIG. 18C

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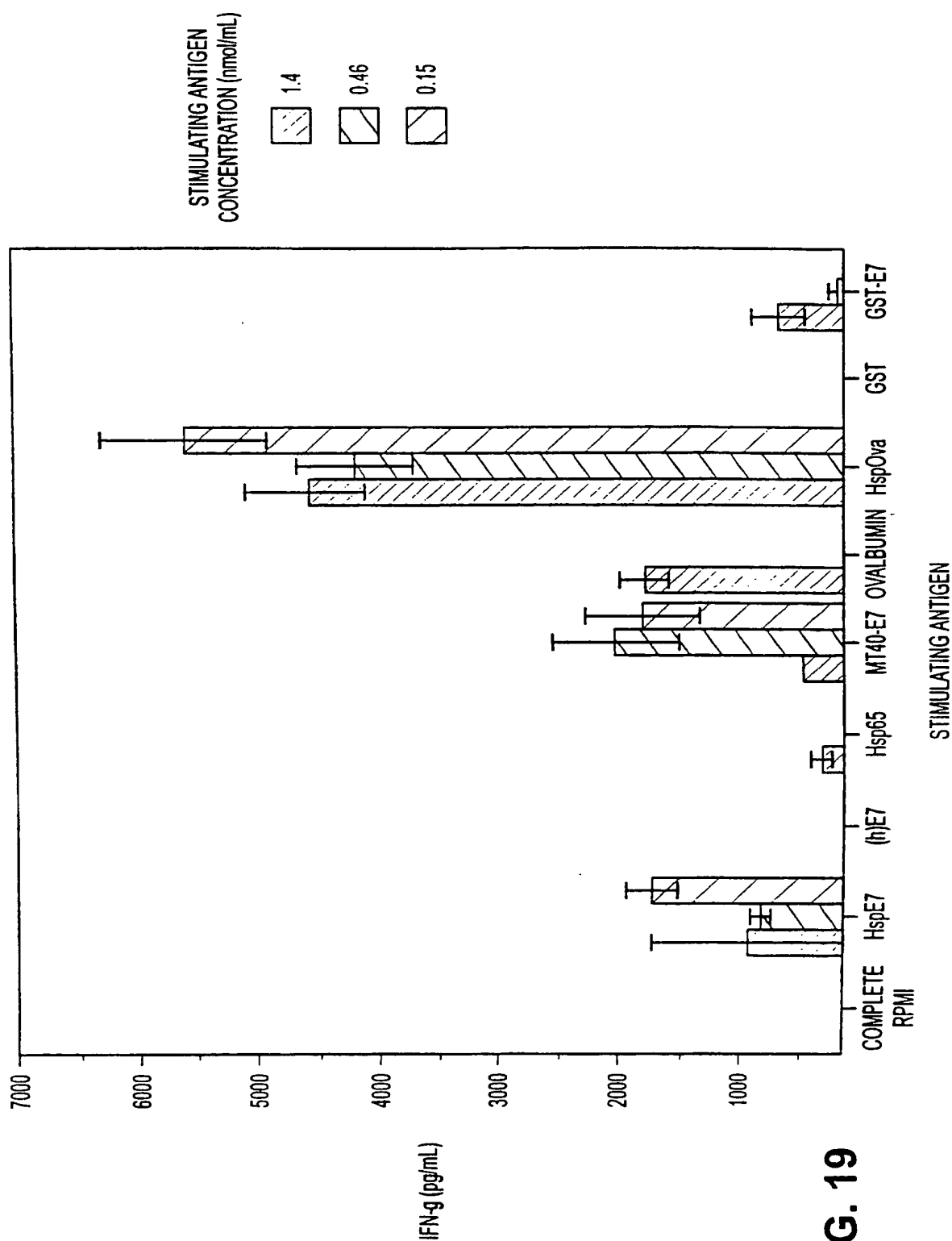


FIG. 19

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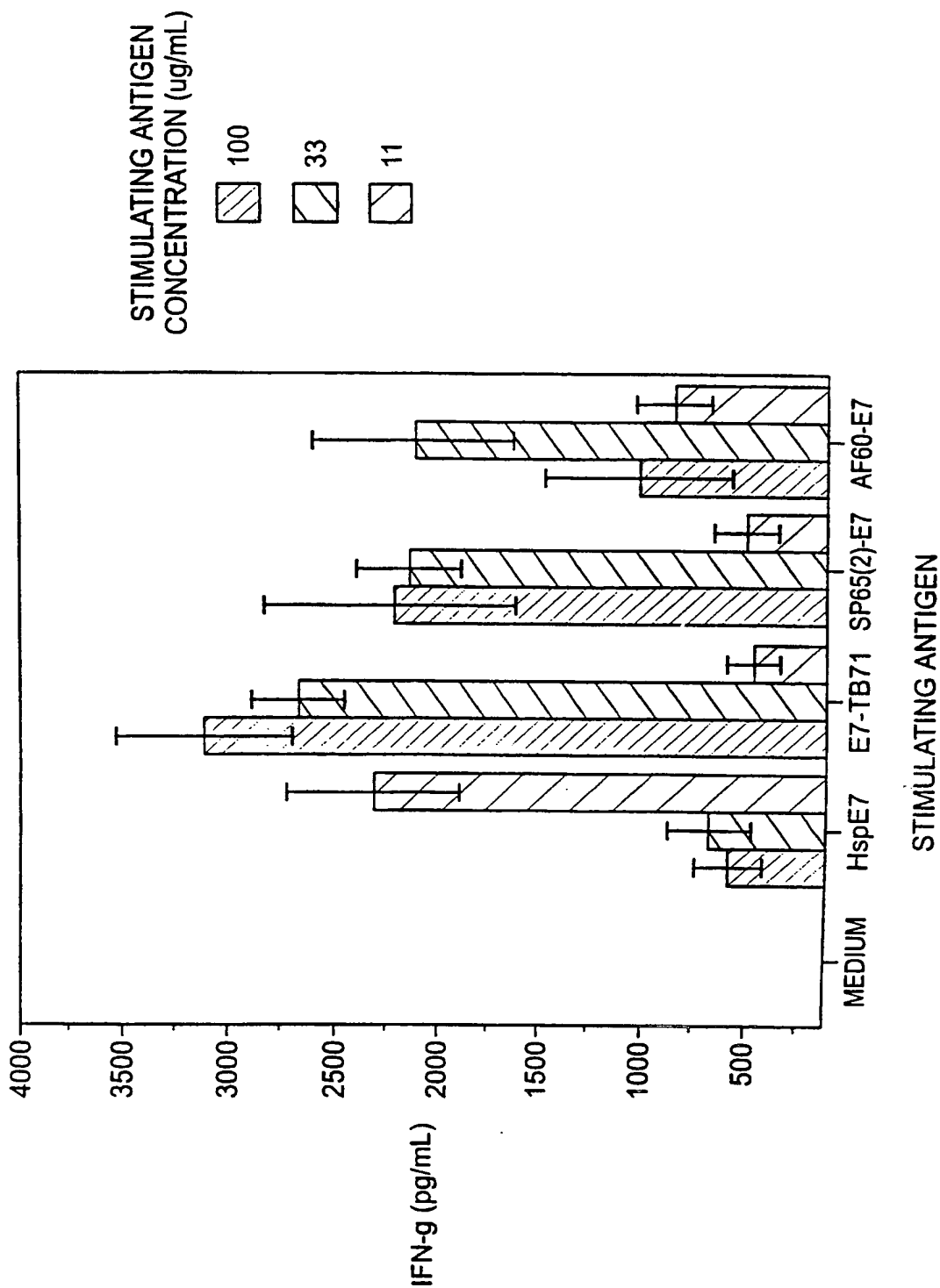


FIG. 20A

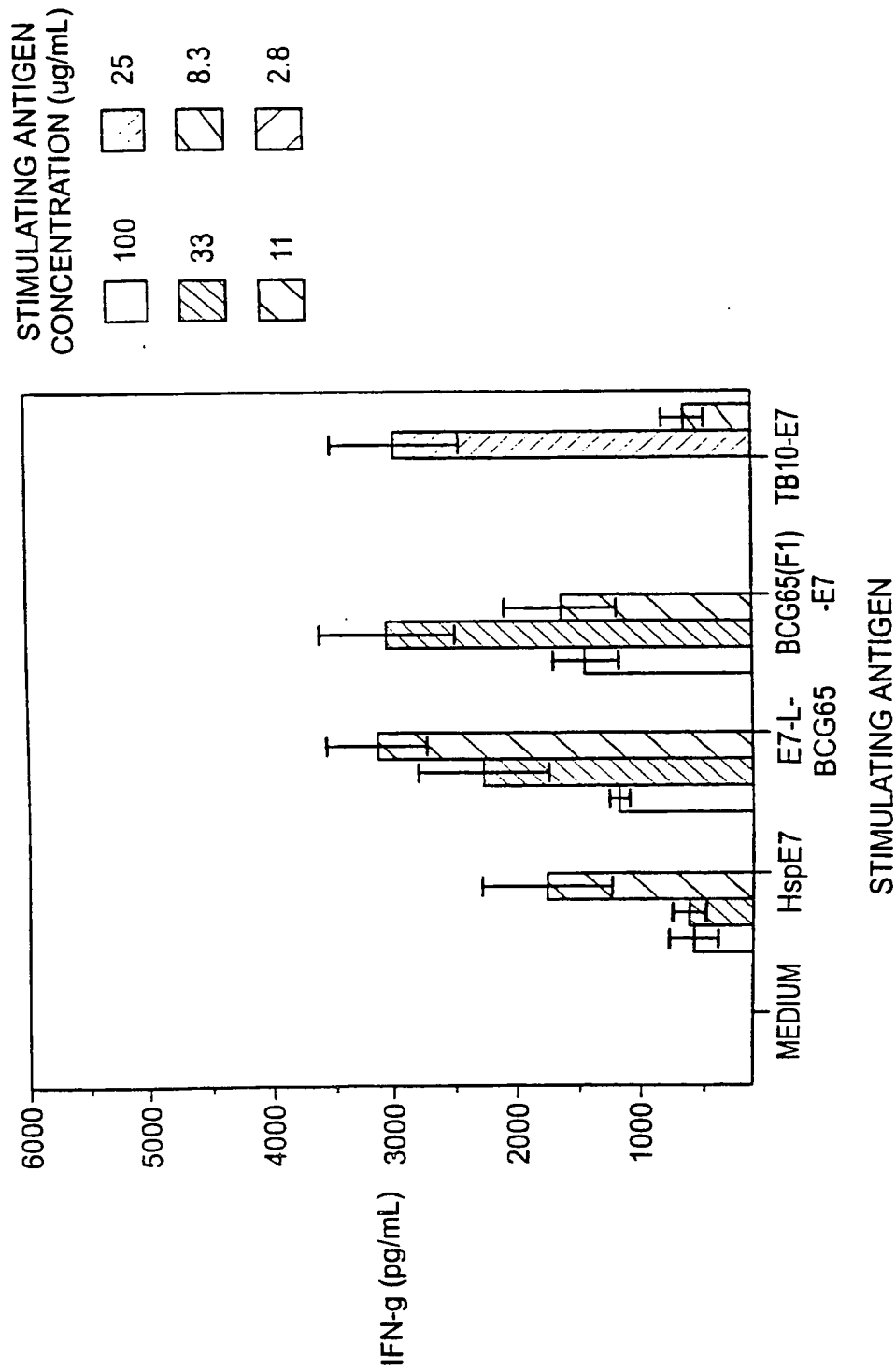


FIG. 20B

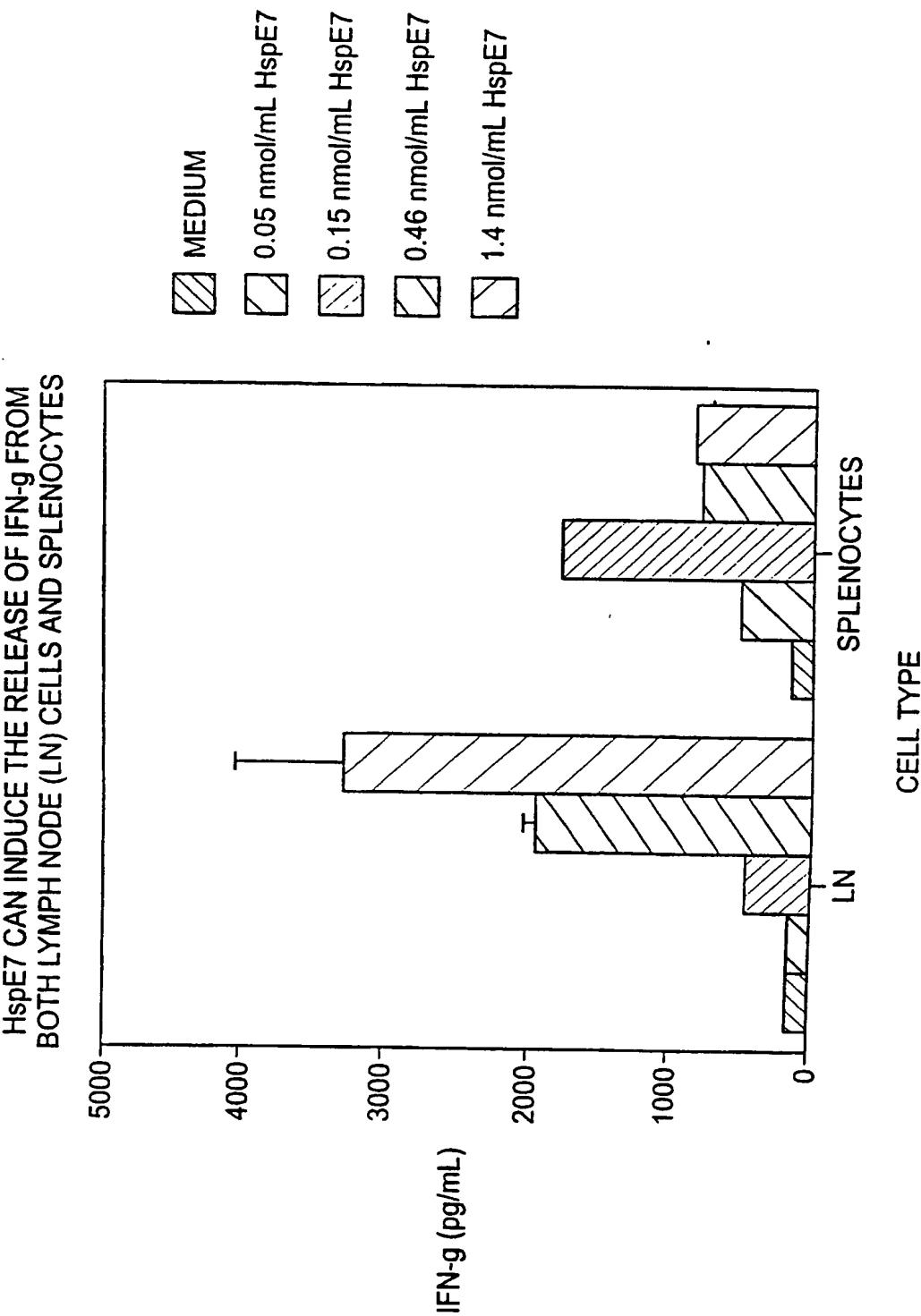


FIG. 21

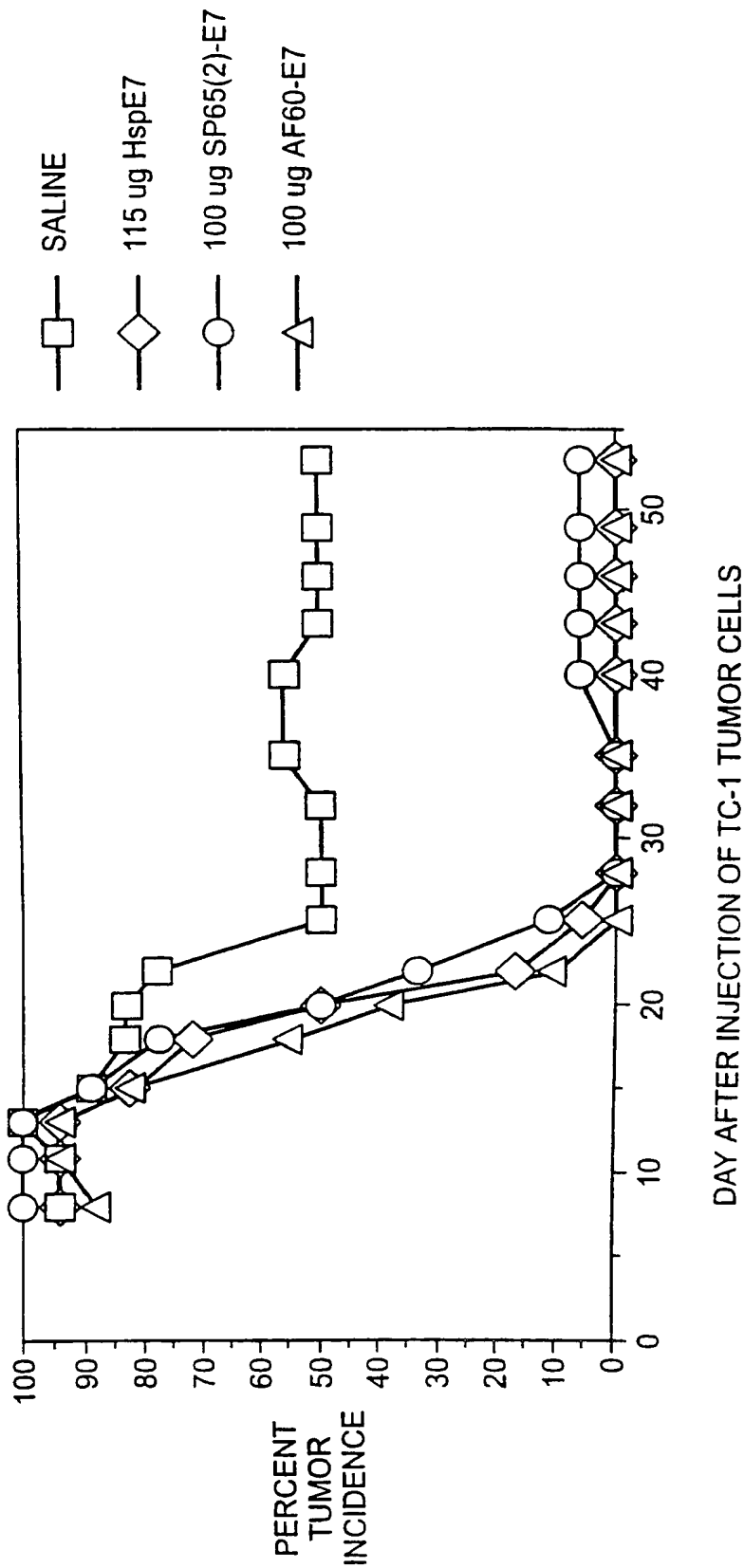


FIG. 22A

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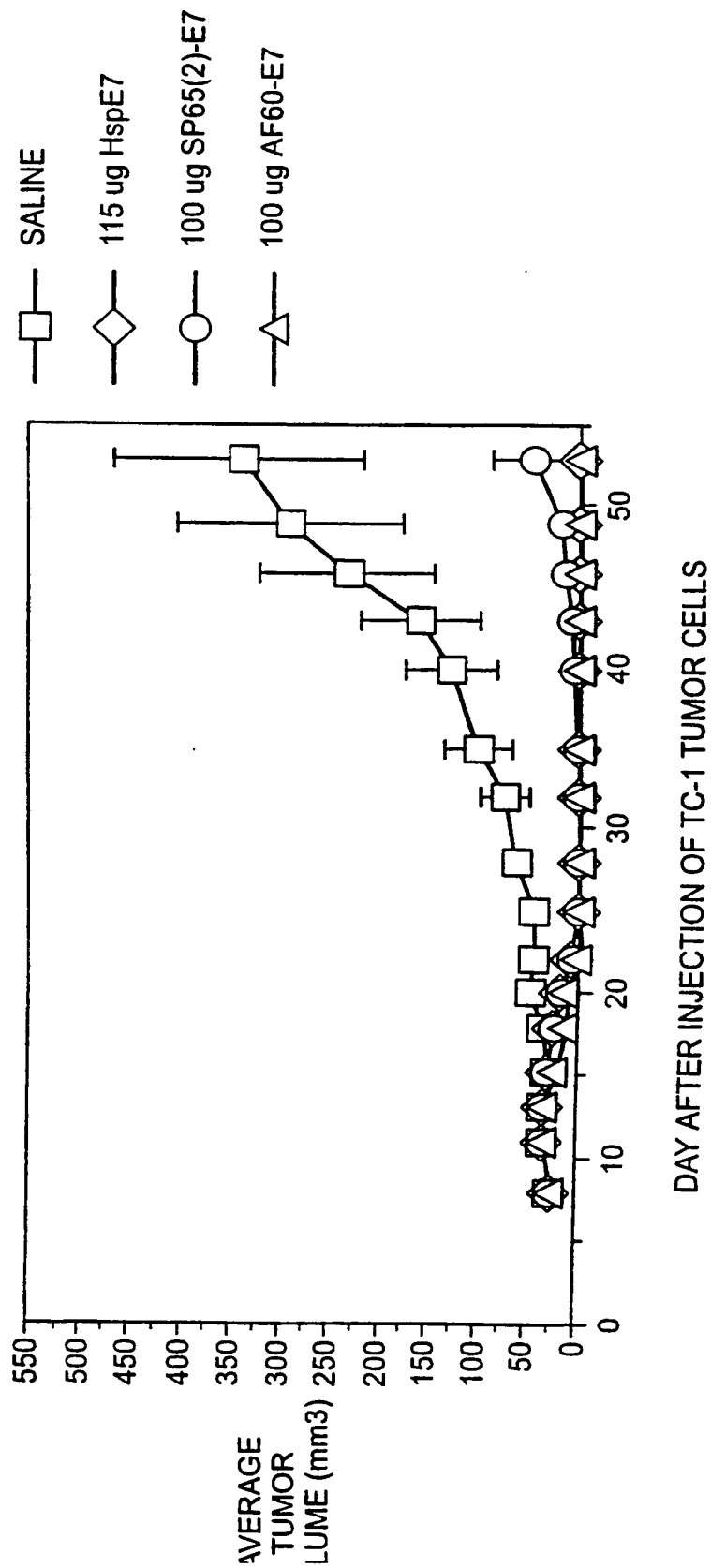


FIG. 22B

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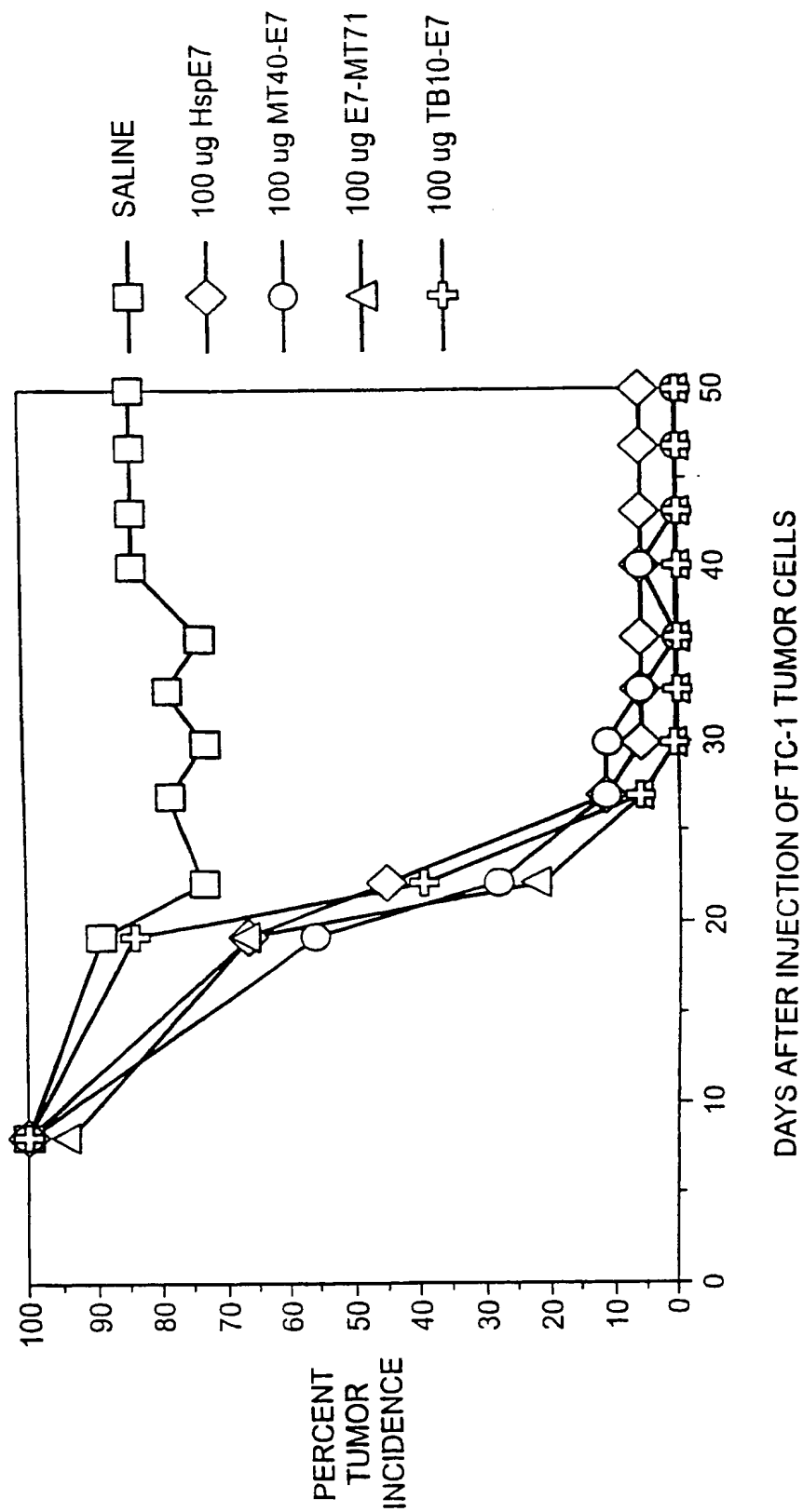


FIG. 23A

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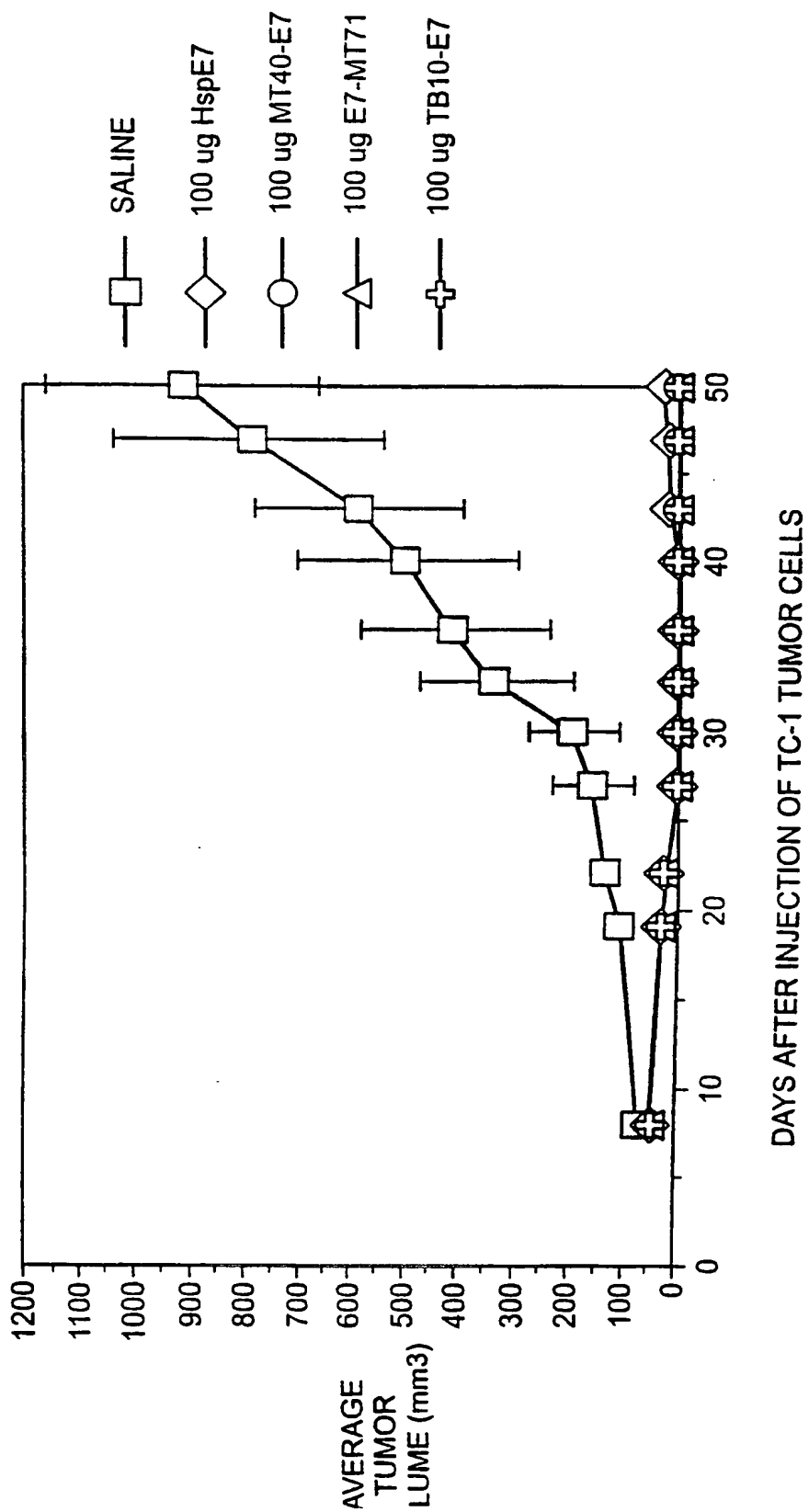


FIG. 23B

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Filed on 8 July 1999 (08.07.1999)(71) Applicant (for all designated States except US):
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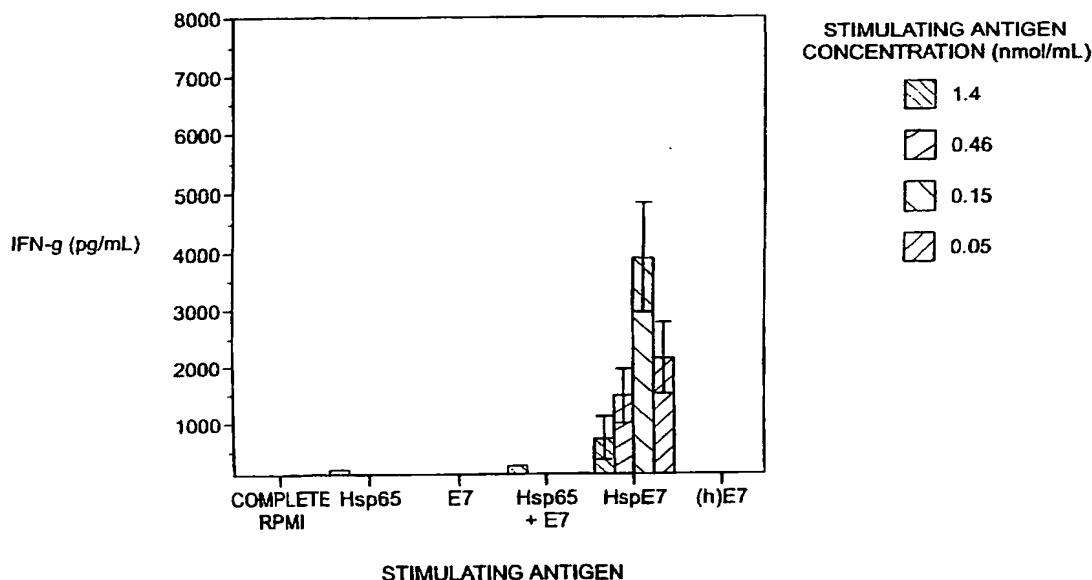
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Franklin Street, Boston, MA 02110-2804 (US).(81) Designated States (national): AE, AG, AL, AM, AT, AU,
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HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR,
LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ,
NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM,
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patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE,

[Continued on next page]

(54) Title: INDUCTION OF A TH1-LIKE RESPONSE IN VITRO

A) SOURCE OF MOUSE: CHARLES RIVER LABS

(57) Abstract: The invention provides compositions and methods for stimulating a Th1-like response *in vitro*. Compositions include fusion proteins and conjugates that contain at least a portion of a heat shock protein. A Th1-like response can be elicited by contacting *in vitro* a cell sample containing naive lymphocytes with a fusion protein or conjugate of the invention. The Th1-like response can be detected by measuring IFN-gamma produced by the cell sample.

WO 01/04344 A3



IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

(88) Date of publication of the international search report:
15 November 2001

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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

INTERNATIONAL SEARCH REPORT

International Application No

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A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 G01N33/50 C07K14/35 C07K14/025 C12N15/62

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

BIOSIS, MEDLINE, EPO-Internal, WPI Data, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>SUZUE K ET AL: "ADJUVANT-FREE HSP70 FUSION PROTEIN SYSTEM ELICITS HUMORAL AND CELLULAR IMMUNE RESPONSES TO HIV-1" JOURNAL OF IMMUNOLOGY, US, THE WILLIAMS AND WILKINS CO. BALTIMORE, vol. 156, 1996, pages 873-879, XP002070468 ISSN: 0022-1767 abstract page 873, right-hand column, line 6 - line 16 page 877, right-hand column, line 41 - line 50</p> <p>---</p> <p>-/--</p>	1-34, 39-42, 64

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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"A" document defining the general state of the art which is not considered to be of particular relevance

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"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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"&" document member of the same patent family

Date of the actual completion of the international search

4 December 2000

Date of mailing of the international search report

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INTERNATIONAL SEARCH REPORT

Inte. Jnal Application No
PCT/US 00/18828

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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A	<p>--- BARRIOS C ET AL: "Heat shock proteins as carrier molecules: In vivo helper effect mediated by Escherichia-coli GroEL and DnaK proteins requires cross-linking with antigen." CLINICAL AND EXPERIMENTAL IMMUNOLOGY, vol. 98, no. 2, 1994, pages 229-233, XP000965591 ISSN: 0009-9104 abstract</p>	1-34, 39-42,64
A	<p>--- WO 96 19496 A (CSL LTD ; UNIV QUEENSLAND (AU); EDWARDS STIRLING JOHN (AU); COX JOH) 27 June 1996 (1996-06-27) claims 7,8</p>	1
A	<p>--- WO 95 31994 A (YEDA RES & DEV ; COHEN IRUN R (IL); FRIDKIN MATITYAHU (IL); KONEN W) 30 November 1995 (1995-11-30) claim 16</p>	1-34, 39-42,64
A	<p>--- WO 93 17712 A (SCLAVO BIOGINE SPA) 16 September 1993 (1993-09-16) abstract; claim 3</p>	1-34, 39-42,64
A	<p>--- ANTHONY L S D ET AL: "Priming of CD8+ CTL effector cells in mice by immunization with a stress protein-influenza virus nucleoprotein fusion molecule" VACCINE, GB, BUTTERWORTH SCIENTIFIC. GUILDFORD, vol. 17, no. 4, 1 February 1999 (1999-02-01), pages 373-383, XP004144750 ISSN: 0264-410X abstract</p>	1-34, 39-42,64
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Int'l. Application No

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,A	<p>DATABASE WPI Section Ch, Week 200038 Derwent Publications Ltd., London, GB; Class B04, AN 2000-431995 XP002154481 & CN 1 248 631 A (ZHOU G), 29 March 2000 (2000-03-29) abstract</p>	<p>1-34, 39-42,64</p>
T	<p>CHU N R·ET AL: "Immunotherapy of a human papillomavirus (HPV) type 16 E7-expressing tumour by administration of fusion protein comprising Mycobacterium bovis bacille Calmette-Guerin (BCG) hsp65 and HPV16 E7." CLINICAL AND EXPERIMENTAL IMMUNOLOGY, vol. 121, no. 2, August 2000 (2000-08), pages 216-225, XP000965423 ISSN: 0009-9104 the whole document</p>	<p>1-34, 39-42,64</p>

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International Application No

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